Spatial versus Temporal Structure: Implications of Inter-haul Variation and Relatedness in the North East Atlantic Spurdog *Squalus acanthias*.

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# Running title: Genetic structure Squalus acanthias NEA

1 Abstract:

1) Micro-population processes, such as gene flow, operating within geographic regions
are often poorly understood despite their potential to impact stock structure and
sustainability. This is especially true for highly mobile species, such as elasmobranchs,
where the potential for spatial overlap of regional populations is increased due to
higher movement capabilities. A lack of information on these processes means
management plans rarely consider spatio-temporal structure.

2) Spurdog (*Squalus acanthias*) are globally distributed throughout temperate regions
yet there is an apparent lack of gene flow between ocean basins. In the North-east
Atlantic, there is little work on gene flow within the region which is currently managed
as a single stock that is estimated to be at 19% compared to 1905. Some evidence from
this region suggests population processes which have the potential to cause
structuring.

3) Population structure of NE Atlantic spurdog was investigated using an 828-bp
fragment of the mitochondrial DNA control region and seven focal polymorphic
microsatellite markers. Samples from 295 individuals from eight locations throughout
UK waters were used in this study.

4) Overall, mitochondrial sequences suggested some regional differentiation. Genetic
diversity was comparable with that reported in previous studies of spurdog. Haplotype
diversity (0.782 - 1) is amongst the highest observed for an elasmobranch.
Microsatellite markers suggested a high level of relatedness was responsible for
regional population structuring. There was no apparent spatial structure after removal
of 'full sibling' relationships.

5) Inter-haul variation from Celtic Sea samples is suggestive of sub-populations and
aggregation events which may have important implications for fisheries conservation
of this, and other, elasmobranch species.

Key words: coastal, ocean, distribution, fish, fishing, elasmobranch management,
genetic structure, relatedness, social interactions

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# 30 1.Introduction

A fundamental challenge to the conservation of marine organisms is defining an 31 appropriate spatial scale at which to manage a species, a conundrum which has led to 32 the grouping of exploited marine organisms into stocks. As a general rule, stocks are 33 defined as populations that encounter no genetic or demographic overlap with each 34 other (Waples, 1998). The idealised concept of a genetically isolated stock arises when 35 a population becomes divided and gene flow ceases. Having achieved demographic 36 independence, the two populations will then begin to gradually diverge, until each is 37 characterized by a distinctive genetic signature (Palsbøll, Bérubé, & Allendorf, 2007; 38 Waples & Gaggiotti, 2006). In the aquatic environment, demographic independence 39 does not necessarily equate to spatial isolation and two genetically distinct populations 40 may be sympatric with spatial overlap, as has been shown in freshwater fish (Verspoor 41 & Cole, 1989) and marine mammals (Hoelzel & Dover, 1991). In both cases, the genetic 42 differentiation was caused by different behaviours, either migratory vs residency 43 (Verspoor & Cole, 1989) or foraging strategies (Hoelzel & Dover, 1991). 44

45 Complete genetic isolation in vagile marine species is rare as they experience few 46 physical barriers to gene flow (Palumbi, 2003; Waples, 1998). This often results in 47 species with high dispersal potential showing only slight genetic differentiation 48 throughout their range (Palumbi, 2003). Due to the need to define stocks for

management purposes, various threshold levels of gene flow have been proposed to 49 define a population and/or a management unit (Reiss, Hoarau, Dickey-Collas, & Wolff, 50 2009), one of which has been to identify separate stocks if a significant and 51 reproducible genetic differentiation can be detected (Reiss et al., 2009). In some cases, 52 this significant differentiation may be very small and the results of combined genetic 53 and tagging studies suggest that in teleost fish, small FsT values of 0.0037 can 54 represent effective regional isolation with a dispersal rate of less than 1% (Knutsen et 55 56 al., 2011).

Such considerations may be especially important in interpreting genetic 57 58 differentiation in elasmobranchs that, due to slow rates of evolution (Martin et al., 1992), can have even lower genetic variability than widespread teleost fish (Heist & 59 Gold, 1999). Genetic differentiation in elasmobranch species appears to be highly 60 correlated with dispersal ability (Veríssimo, McDowell, & Graves, 2010) with some 61 cosmopolitan species displaying very little global genetic differentiation (Schmidt et 62 al., 2009). This may suggest that genetic investigations into regional stocks would 63 yield little in the way of spatial resolution. There are however certain behaviours that 64 have been documented in elasmobranch populations which may promote genetic 65 differentiation, even in wide-ranging species. 66

Population segregation based on age or sex, as is common in many elasmobranch
species (Alonso et al., 2002; Hurst, Bagley, McGregor, & Francis, 1999), may allow one
of the sexes, more commonly females (Pardini et al., 2001; Portnoy, McDowell, Heist,
Musick & Graves, 2010; Daly-Engel et al., 2012), to maintain site fidelity. This,
especially when in conjunction with reproductive behaviour (i.e. natal philopatry (Hueter, Heupel, Heist, & Keeney, 2005)), can create genetic differentiation between
regions that are not separated by physical barriers to gene flow (Dudgeon et al., 2012;

Feldheim et al., 2002; Jorgensen et al., 2010; Pratt & Carrier, 2001). Hence, there is the potential to create genetically distinct sympatric populations, highlighting a difference between spatially and genetically distinct stocks. Anthropogenic exploitation also has the potential to impact the genetic composition of a species within a region; a significant population decline caused by overexploitation can cause range contraction and population fragmentation (Kenchington, 2003), which may encourage the formation of small, isolated sub-populations.

Spurdog (Squalus acanthias) are generally considered to be highly mobile (Gauld and 81 Macdonald, 1982; Templeman, 1976; Templeman, 1984) and are distributed 82 83 worldwide throughout temperate continental shelf seas (Camhi, Valenti, Fordham, Fowler, & Gibson, 2009). No regional genetic differentiation has been found to date 84 (Hauser, Franks, Vega, & Gallucci, 2007) and it appears that oceanic water depths do 85 86 not act as a migratory barrier for this species as no differentiation has been found between the southern Atlantic and Pacific, nor between the eastern and western North 87 88 Atlantic; a concept supported by tagging evidence indicating transoceanic movements (Holden, 1967; Templeman, 1976). However, taking into account that just one 89 breeding migrant per generation may be enough to prevent, or greatly reduce, genetic 90 differentiation between typically discrete geographic populations (Slatkin, 1987; 91 Spieth, 1974), this is perhaps not surprising. Gene flow does, however, appear to be 92 restricted across equatorial waters (Hauser et al., 2007). This barrier to gene flow has 93 caused divergence to the point whereby the North-eastern Pacific spurdog has been 94 re-classified as S. suckleyi (Ebert et al., 2010). Squalus suckleyi has markedly different 95 population dynamics to S. acanthias, with a later age of maturity, larger maximum 96 size and larger length at maturity (Hauser et al., 2007; Ketchen, 1972; Saunders & 97 98 McFarlane, 1993).

In contradiction to the genetic data, there is evidence suggesting more limited regional 99 movements in some populations. Based on mark and recapture tagging, some spurdog 100 appear to maintain a certain level of site association, often within coastal areas, where 101 sub-units appear to restrict movements to the coastal shelf and partially enclosed 102 water bodies (Campana et al., 2009; Carlson et al., 2014; Ketchen, 1986; Templeman, 103 1984; Thorburn, Neat, Bailey, Noble, & Jones, 2015). In contrast, offshore units appear 104 highly migratory (Campana et al., 2009; Ketchen, 1986; McFarlane & King, 2003). 105 Such differentiation may be a consequence of populations partitioning by age or sex, a 106 feature of many other elasmobranch species (Alonso et al., 2002; Hurst et al., 1999). 107

108 Spurdog in the NE Atlantic are currently viewed as a single stock (Aasen, 1964; Gauld & Macdonald, 1982; Vince, 1991), a designation based on mark and recapture tagging 109 data demonstrating wide-ranging movements throughout this region (Gauld & 110 Macdonald, 1982; Templeman, 1976, Templeman, 1984; Vince, 1991). In 2010 The NE 111 Atlantic spurdog biomass was estimated to be about 23% of 1955 numbers and 19% 112 compared to 1905 (Oliveira, Ellis, & Dobby, 2013), but appear to have stabilized over 113 the last decade (ICES, 2016). There is limited evidence suggesting that some spurdog 114 in this region display winter residency within semi-enclosed coastal regions (Thorburn 115 et al., 2015), however, it is unknown if these individuals represent an isolated group or 116 if the area is used by a proportion of the wider population. Historical exploitation and 117 the subsequent reduction in biomass of the NE Atlantic spurdog places importance on 118 the appropriate management of the remaining population. There are management 119 schemes, such as spatial and temporal measures, currently (2015 - 2019) under trial 120 in the Celtic Sea; the Spurdog By-catch Avoidance Programme (Hetherington, 121 Nicholson, & O'Brien, 2016). These schemes aim to actively avoid spurdog 122 aggregations and reduce spurdog by-catch and discards, reducing overall fishing 123

mortality using information supplied by the fishing industry in real time. However, 124 management also needs to be on a wider scale for which it is essential we understand 125 the full range of movements and connectivity in the species. Beyond this, it is vital to 126 understand the impact of commercial fisheries on elasmobranch populations as, 127 beyond direct population reduction, fisheries exploitation can have significant direct 128 and indirect effects on communities (Jennings & Kaiser, 1998) by targeting specific 129 age and sex classes. In elasmobranchs, these processes can lead to changes in the age 130 structure and size of individuals within a population (Hutchings, 2005; Stevens, 131 Bonfil, Dulvy, & Walker, 2000) as well as changes to the community structure of 132 elasmobranchs within a given area (Walker & Hislop, 1998). 133

A prerequisite for sustainable management is the matching of biological processes and 134 management action (Reiss et al., 2009). As recent studies have shown, some 135 individuals display strong site association in coastal regions (Thorburn et al., 2015), 136 suggesting investigation of population genetic structure in the spurdog population 137 around the UK may indicate if there is partitioning suggestive of site 138 fidelity/philopatric behaviours. This would have implications for population 139 management. To explore this, an 828-bp fragment of the mitochondrial DNA 140 (mtDNA) control region and seven focal polymorphic microsatellite markers (µSAT) 141 were used to determine levels of gene flow and relatedness in spurdog around the UK. 142

### 143 **2. Methods**

## 144 **2.1 Tissue sample collection**

Samples from 295 individuals from eight locations throughout UK waters (Figure 1,
central Latitude and Longitude points for each sampling area is given in
Supplementary table 1) collected between 2010 and 2013 were used in this study.

148 Additional information on individuals sampled can be found in Supplementary table 1. Collections were from research trawls (North Scotland, Rockall, and Jura) 149 undertaken by Marine Scotland (MS), the offshore gill net fishery (also deploying long-150 lines opportunistically) (Celtic Sea), inshore longline fishery (Southern North Sea, 151 Wales) and recreational fishery (rod and line) (Wales) with sampling undertaken by 152 the Centre for Environment, Fisheries & Aquaculture Science (CEFAS) and the 153 recreational fishing community (rod and line) (Loch Sunart, Loch Etive). All tissue 154 samples were collected from the trailing edge of the first dorsal fin and immediately 155 preserved in 70-95% ethanol. Additionally, total length (TL) (snout to the tip of the 156 terminal dorsal lobe on the tail fin), and girth for most, but not all, of the spurdog 157 (around the central thorax, in front of the first dorsal fin and behind the pectoral fins) 158 159 were recorded. All licensed scientific sampling was carried out under the Animals (Scientific Procedures) Act 1986, using the appropriate project and personal licences 160 authorized by the United Kingdom Home Office. Genomic DNA was extracted from 161 352 tissue samples using phenol-chloroform protocols (Sambrook & Russell, 2006) 162 163 with 100% ethanol precipitation.

# 164 2.2 Mitochondrial DNA Sequencing

PCR amplification of an approximately 900 base pair (bp) section of the mitochondrial 165 166 control region (CR) conducted using primers TMTF1 (5'was CCCACATACCCTAATATACCA) and TMTR1 (5'-CATCTTCAGTGCCATACTCT) 167 designed using Primer Premier vers. 5 (Premier Biosoft International). A 50µL PCR 168 master mix was used which included approximately 25-50ng DNA, 0.5µM of each 169 primer, 10µl Bioline 5xMyTaq reaction buffer, 0.25µl Bioline MyTaqHS (Hotstart Taq) 170 and UV sterilized water. PCR conditions were: initial denaturation at 95°C for 8min 171 followed by 36 cycles of 95°C for 60sec, 53°C for 60sec and 72°C for 90sec with a final 172

extension step at 72°C for 8min using Biometra TGradient PCR machines. Products
were purified using Qiagen PCR purification kits (www.qiagen.com) prior to
commercial sequencing (Beckman Coulter Genomics).

### 176 **2.3 PCR amplification of the microsatellite regions**

Seven focal nuclear microsatellite markers were used in this study, all of which have 177 been previously published; four loci from McCauley et al., (2004): DF T289, DF J451, 178 DF U285, DF J445 and three from Veríssimo et al., (2010): SACA GA11, SACA 3853 179 180 and SACA 6396. An 11 µL PCR master mix included approximately 1ng DNA, 0.2µL 0.5µM of each primer, 2µL Bioline 5x MyTaq reaction buffer solution, 0.15µL Bioline 181 182 MyTaq, and UV sterilized water. After testing, all seven loci were found to amplify well 183 under the same PCR conditions with an initial denaturation step at 94°C for 5min followed by 35 cycles of 94°C for 60sec, 53°C for 30sec and 72°C for 30sec, with a final 184 185 extension step at 72°C for 7min. Samples were sent for commercial genotyping (MRC 186 PPU DNA Sequencing and Services, Dundee).

## 187 2.4 Mitochondrial control region data analysis

188 Sequences were aligned manually and edited using MEGA vs 6 (Tamura, Stecher, Peterson, Filipski, & Kumar, 2013), providing sequences of 828bp. Diversity indices 189 190 for mtDNA sequences (number of haplotypes (H), haplotype (h) and nucleotide (n)diversity) were calculated in DNASP version 5 (Librado & Rozas, 2009). Regional 191 genetic differentiation was estimated in ARLEQUIN version 3.5.1.2 (Excoffier & 192 Lischer, 2010) using genetic-distance based pairwise  $\Phi_{ST}$ , with 10,000 permutations 193 to estimate significance and the Kimura 2P model as selected by JModeltest (Darriba, 194 Taboada, Doallo, & Posada, 2012). 195

196 To examine genealogical relationships among haplotypes, a neighbour-joining network was created in PHYML 3.1 (Guindon, Delsuc, Dufayard, & Gascuel, 2009) and 197 visualized using HAPVIEW (Salzburger et al., 2011). Demographic history was 198 explored using neutrality tests (Tajima's D test (Tajima, 1989) and Fu's Fs test (Fu 199 1996)), implemented in ARLEQUIN with p-values generated from 10,000 200 simulations. Significant negative Tajima D or Fu's F values are indicative of recent 201 population expansion. A mismatch analysis was also undertaken in ARLEQUIN to 202 explore further demographic history; a smooth, unimodal peaked distribution with a 203 skew towards zero is suggestive of a historical population expansion event 204 (Harpending, 1994; Rogers & Harpending, 1992) while a ragged, erratic distribution 205 is suggestive of population stability (Harpending, 1994; Rogers & Harpending, 1992). 206

# 207 2.5 Microsatellite data analysis

The total number of alleles, observed heterozygosity (H<sub>0</sub>), expected heterozygosity 208 (He), allelic richness (A), Wright's inbreeding coefficient (FIS; 10000 bootstrap 209 replicates), linkage disequilibrium and deviation from Hardy Weinberg Expectations 210 (HWE) (Monte Carlo replicates 10000) were estimated in the R package routine 211 diveRsity (Keenan, McGinnity, Cross, Crozier, & Prodöhl, 2013), and GENEPOP ver. 212 4.3 (Rousset, 2008). Regional genetic differentiation was estimated by pairwise FsT in 213 ARLEQUIN. Significant Fst differences after Bonferroni sequential correction were 214 visualized by a factorial correspondence plot, calculated in GENETIX ver. 4.05 215 (Belkhir, Borsa, Chikhi, Raufaste, & Bonhomme, 2004), and plotted using the R 216 package routine GGplot2 (Wickham & Chang, 2009). Regional population 217 differentiation was also explored using STRUCTURE 2.3.4 (Pritchard, Stephens, & 218 Donnelly 2000). Maximum number of populations (K) 1-8 was tested based on the 219

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number of sampling sites. Each run consisted of a 1,000,000 run burn-in period and 1,000,000 MCMC iterations, both with and without prior location information. 221

Levels of relatedness, both inter and intra regionally and between temporal sampling 222 episodes from the same site, were investigated using GENALEX (Peakall & Smouse, 223 2006) and MLRELATE (Kalinowski et al., 2006). Exploration of temporal variation 224 was only possible for Loch Etive, Loch Sunart, and the Celtic Sea due to the limited 225 number of temporal samples at other sites. In GENALEX, Lynch & Ritland (1999) and 226 Queller & Goodnight (1989) estimators of relatedness were used to investigate 227 pairwise relationships and mean levels of population relatedness; using 10000 228 permutations and 10000 bootstraps in each simulation. MLRELATE was used to 229 calculate the maximum likelihood estimates of relatedness and relationships taking 230 null alleles into account, using 10000 bootstraps. The significant number of full sibling 231 (FS values =>0.5) relationships identified by MLrelate may have influenced estimates 232 of intra-regional structuring. Therefore, microsatellite regional differentiation tests 233 were re-rerun removing the individuals with the highest pairwise FS relationships 234 within the population. This ensured retention of the maximum number of individuals 235 per population. Related individuals were not removed for mtDNA analysis due to low 236 levels of haplotype differentiation in initial investigations. 237

Sex-biased dispersal was explored in FSTAT (Goudet, 2001) by comparing 238 microsatellite fixation index (FsT), inbreeding coefficient (FIS), levels of relatedness (r), 239 mean assignment indices (mAIc) and variance of assignment index (vAIc) between 240 males and females from each sampling site. Different levels of dispersal in each sex 241 will, in principle, lead to unequal levels of gene flow. This may create a Wahlund effect, 242 producing higher FIS and vAIC values, lower FST and relatedness values, and negative 243 mAIc values in the dispersing sex; full descriptions of tests are given in Goudet et al., 244

(2002). Only mature individuals were used for analysis as tests assume sampled
individuals have already had the opportunity to disperse (Goudet et al., 2002). Due to
the low numbers of mature spurdog of each sex from some sampling locations, only
five regions were used in these analyses (Celtic Sea, Loch Etive, Loch Sunart, North
Scotland and Southern North Sea).

250 3. Results

## 251 3.1 Genetic Diversity

Alignment of 828 bp mtDNA control region sequences from 228 individuals gave 49
haplotypes based on 25 polymorphic sites, 16 of which were parsimoniously
informative. Of the 49 haplotypes, 26 were singletons. As many singletons can imply
PCR and sequencing errors (Johnson & Slatkin, 2006), 10% of samples had PCR rerun and were re-sequenced. Error rate was found to be <5% based on re-sequencing.</li>
Haplotype diversities ranged from 0.782 (Wales) to 1.00 (North Scotland) (Table 1).
Nucleotide diversities ranged from 0.0015 (Wales) to 0.0035 (Rockall) (Table 1).

A total of 295 individuals were genotyped for seven microsatellite loci. To establish 259 PCR or genotyping errors, 10% of samples had PCRs re-run and were re-genotyped 260 261 establishing the error rate to be <5%. Prior to FS relationships being removed from 262 the full dataset, significant departures from HWE were detected at five loci, DF T289, SACA 6396, DF J451, DF J445 and SACA 3853, within five populations, Loch Etive, 263 Sound of Jura, southern North Sea, Wales, and the Celtic Sea. The observed 264 heterozygosity ranged from 0.58 (Etive) - 0.7 (N.Sco) (Table 2; full population 265 summaries per locus in supplementary Table 2. With FS relationships removed, SACA 266 267 6396 in both the Celtic Sea and Southern North Sea and SACA 3853 in the Southern North Sea population were out of HWE (Table 2; full population summaries per locusin Supplementary Table 1;). All loci were in linkage equilibrium.

# 270 3.2 Population differentiation

### 271 3.2.1 Spatial structure

Of the 49 haplotypes identified the mtDNA haplotype network showed that six common haplotypes are shared between most regions, indicating little evidence of population structure around the UK as haplotypes did not appear to group by sample location. A minor exception to this is Jura, which is not represented on several of the multi-haplotype branches (Figure. 2). As expected, other less frequent haplotypes, mainly singletons (33), branch from the common haplotypes.

278 Mitochondrial analyses showed evidence of weak spatial differentiation, with 279 significant (p<0.05) pairwise  $\Phi_{ST}$  differences detected between Rockall and the Celtic 280 Sea ( $\Phi_{ST}$  0.1551), Etive ( $\Phi_{ST}$  0.1727), Jura ( $\Phi_{ST}$  0.2681), Southern North Sea ( $\Phi_{ST}$ 281 0.1587), North Scotland ( $\Phi_{ST}$  0.1917); and Jura and Sunart ( $\Phi_{ST}$  0.0617), Wales ( $\Phi_{ST}$ 282 0.1138) and the Celtic Sea ( $\Phi_{ST}$  0.0605) following FDR correction (Table 3). After more 283 stringent Bonferroni sequential correction, however, only Jura and Rockall ( $\Phi_{ST}$ 284 0.2681) remained significantly differentiated (Table 3).

Including all individuals, microsatellite evidence suggested there was some genetic differentiation, with significant pairwise  $F_{ST}$  between Wales and Jura ( $F_{ST}$  0.0305) following FDR correction. After Bonferroni correction, FST remained significant between Wales and the Celtic Sea ( $F_{ST}$  0.029), Loch Etive ( $F_{ST}$  0.038), Loch Sunart ( $F_{ST}$  0.041) and the southern North Sea ( $F_{ST}$  0.034) (Table 3). Factorial correspondence analysis shows that while the population mean of Wales is discrete from those of other populations there is some overlap of individuals (Figure 3). After removing full sibling (FS) relatives (as identified by MLrelate) from each sampling site no significant genetic differentiation between regional populations was apparent (F<sub>ST</sub> range -0.01867 - 0.01113) (Table 3). Results from STRUCTURE both with and without priors showed no regional differences, with K= 1 for both and no discernible pattern in either case (data not shown).

## 297 3.2.2 Temporal structure

298 No temporal variation was observed in samples from Loch Etive or Loch Sunart, however, it should be noted that samples from these regions comprised largely 299 individuals taken opportunistically from angling trips where typically low numbers of 300 fish are taken at a single spatiotemporal sampling point. Samples taken from 301 commercial fishing vessels in the Celtic Sea at four separate time points 2010, 2010a, 302 2011, 2011a (Table 4) showed significant (p<0.05) differentiation after FS had been 303 removed between 2010 and 2010a (Fst 0.028) following FDR, and between 2010a and 304 2011 (Fst 0.037), following Bonferroni correction (Table 5). Factorial correspondence 305 analysis showed discrete population means, but admixture of individuals is evident 306 (Figure. 4). There was no significant variation in the number of FS relationships 307 308 between samples from each sampling occasion (Table 4). The ratio of full sibling relationships was higher in the temporally split Celtic Sea groups (Table 4) than it was 309 for the Celtic Sea as a whole (Table 2). Sex ratios varied between the four commercial 310 fishing trips. In 2010a, only one male was caught out of 18 spurdog; in 2010 and 2011, 311 numbers of males and females were approximately equal; while in 2011a, there were 312 twice as many females caught as males (Table 4). 313

# 314 3.3 Relatedness

All regions had a similar proportion of FS relationships (relatedness values >0.5)
between sampling sites (range 0.028 (N. Sco) – 0.034 (Rockall)) (Table 2). Samples

from Wales had a considerably higher number of intra-region FS relationships (0.368) compared to other sampling sites which had similar, low levels of FS relationships (mean = 0.033, range 0.000 (N.Sco) – 0.0071 (Rockall)) (Table 2). Further investigation into relatedness using GenAlEx showed samples from Wales to have a significantly higher mean relatedness (r = 1.12, p<0.05) than expected (Figure. 5). No other populations had significantly high or low levels of relatedness. All spurdog caught in Wales were adult females, TL 94 – 111 cm.

#### 324 3.4 Sex biased dispersal

There was limited evidence of male dispersal when comparing of male and female microsatellite data, which showed that males had lower F<sub>IS</sub> and F<sub>ST</sub> values, a lower level of relatedness, lower mAIC and higher vAIC values. However, only vAIC showed a significant difference (Table 6).

329 3.5 Demographic history

All sites except Rockall displayed significant negative Fu's FS values (Table 1). No sampling sites had significant Tajima's D values. The mismatch distribution showed a smooth unimodal distribution with a heavy left skew towards zero (Figure. 6). Due to the low number of samples sequenced, neutrality tests could not be performed on sequences from North Scotland.

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- 336 4. Discussion
- 337 4.1 Genetic Diversity

Haplotype diversity (0.782 – 1.0) was comparable to that found for *S. suckleyi* in the
NE Pacific (0.966; (Hauser et al., 2007)), but higher than that previously reported for *S. acanthias* in the North-east Atlantic (0.56 – 0.71; Veríssimo et al., 2010). The

nucleotide diversities (0.0015 - 0.0035) compare to previously published values for *S. acanthias* from sites within the NE Atlantic from the west of Ireland (0.0024) and the Irish Sea (0.0016) (Veríssimo et al., 2010), yet lower than those reported for *S. suckleyi* in the NE Pacific (0.0072; Hauser et al., 2007). The diversity values for *S. suckleyi* in the NE Pacific only came from a small sample size (Hauser et al., 2007) so may not be fully representative of the true genetic diversity of the NE Pacific spurdog popualtion.

The genetic variation observed in the mtDNA in this study is among the highest 348 recorded of any elasmobranch (e.g. Blower, Pandolfi, Bruce, GomezCabrera, & 349 Ovenden, 2012; Keeney, Heupel, Hueter, & Heist, 2005; Stow et al., 2006; Tavares et 350 al., 2013; Valsecchi, Vacchi, & Di Sciara, 2005). This is surprising as spurdog in the 351 NE Atlantic are known to have undergone a severe decline in biomass (Oliveira et al., 352 2013) which might be expected to erode much of the genetic diversity (Kenchington, 353 2003). While this may be indicative of a mutational hotspot in the Control Region, as 354 observed in other elasmobranch species (Tavares et al., 2013), this cannot be shown 355 by this data and should be explored further by sequencing the entire mitogenome. 356

Another possibility for the observed haplotype diversity would be some form of 357 aggregating behaviour. Aggregations may occur in response to high levels of 358 exploitation, as previously observed in both teleost's (Rose & Kulka, 1999) and 359 elasmobranchs (Ellis et al., 2008). Spurdog are known to be an aggregating species 360 (Fordham, Fowler, Coelho, Goldman, & Francis, 2006) and if aggregations are formed 361 following exploitation, these would represent convergence of several different sub-362 populations thus increasing genetic diversity. As the UK lies at the junction of several 363 biogeographic zones (e.g. Johnson et al., 2014) where spurdog are known to occur, it 364 is possible that following population reduction, individuals from these different zones 365

366 have come together. These would, potentially, bring unique haplotypes with them and 367 produce the high haplotype diversity observed. However regional genetic distinction 368 was not observed in this study so cannot support this theory. Increased diversity may also come from the NW Atlantic. No genetic differentiation has been found between 369 these two regions (Veríssimo et al., 2010), yet as only one migrant per generation can 370 prevent populations becoming genetically differentiated (Slatkin, 1987; Spieth, 1974), 371 this does not necessarily prove regular admixture. The initial evidence of weak 372 differentiation between Rockall and the rest of the sampling sites observed in the 373 mtDNA does tentatively offer the possibility that this offshore region is slightly 374 different from the rest of the UK, potentially influenced by spurdog from the NW 375 Atlantic. However, small sample sizes and weak differentiation, that is removed with 376 multiple analysis corrections, does not support this hypothesis. Tagging studies have 377 shown some connectivity between the NE and NW Atlantic (Templeman, 1976), yet 378 not in great numbers. If connectivity is low, this could be a means of introducing new 379 haplotypes to the NE Atlantic from the NW Atlantic, increasing diversity while 380 381 maintaining heterogeneity. The level of connectivity between the NE and NW Atlantic, 382 is worthy of further investigation as this has important implications for spurdog stock 383 maintenance and management, because their observed decline in the NE Atlantic may 384 be buffered by numbers and genetic admixture from the NW Atlantic.

Some aggregations are attributed to life history events (Carlson et al., 2014; Pawson, including mating (Carlson et al., 2014) and may not be a result of overexploitation, but a population strategy to allow for genetic mixing helping to retard inbreeding and increase overall genetic diversity (Newby, Darden, Bassos-Hull, & Shedlock, 2014). Significant spurdog by-catch events of approximately 10 tonnes in Celtic Sea fisheries (Hetherington et al., 2016) are not uncommon and demonstratesthe aggregative tendency of spurdog in the UK.

Identification of aggregation behaviour, is important because it can lead to an over
estimation of population numbers, as Catch Per Unit Effort remains high and stable at
aggregation sites while overall abundance declines (Erisman et al., 2011; Rose & Kulka,
1999).

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## 397 **4.2 Population differentiation**

Overall, no structuring was observed in the mtDNA haplotype network, and while 398 significant  $\Phi$ ST values were observed between Rockall and other regions, sample 399 numbers for Rockall were too low to draw firm conclusions regarding genetic 400 differentiation with other regions. The mtDNA also offered no evidence of natal 401 philopatry in spurdog (Hueter et al., 2005). Microsatellite analysis initially showed 402 403 some significant differentiation between Wales and other regions. This appeared to be caused by high numbers of individuals with above-average levels of relatedness in 404 samples from Wales; with these individuals removed there was spatial homogeneity 405 across all sampling locations. However, further intra-regional investigations using 406 407 trawls taken on different dates in the same area did reveal temporal heterogeneity in the Celtic Sea. This may be attributed to genetically distinct sympatric populations 408 using the same region (McPherson, Stephenson, & Taggart, 2003) which has been 409 shown to create similar levels of genetic variation at the same location between 410 samples collected at different times as that across widely separated geographic points 411 (Reiss et al., 2009). As spurdog are known to aggregate, it may be that, following such 412 an event, spurdog split into genetically distinct sub-units, as shown in teleosts 413

(Hutchinson, Carvalho, & Rogers, 2001) and suggested by the discrete trawls in theCeltic Sea.

416 It may be that sub-groups are associated with geographic regions. The British Isles can be considered to lie near the intersection of several biogeographic provinces (e.g. 417 Johnson et al., 2014) as discussed in the diversity section of this paper, the lack of natal 418 philopatry and unambiguous spatial genetic differentiation found in this study does 419 not support the suggestion of geographically separate stocks associated with these 420 regions, despite initial weak differentiation between Rockall and other sites. 421 Furthermore, such geographic separation has not been shown in previous tagging 422 studies which show spatial homogeneity in the NE Atlantic with spurdog being wide 423 ranging and cosmopolitan (Gauld & Macdonald, 1982; McFarlane & King, 2003; 424 Templeman, 1976; Templeman, 1984). 425

Sub-grouping may be a consequence of social structuring. Social relationships have 426 been shown in other free-ranging species of shark which form communities based on 427 428 individual preference to other individuals (Mourier, Vercelloni, & Planes, 2012; Newby et al., 2014). There is evidence of this in some elasmobranch species within spatially 429 limited nursery areas where juveniles and sub-adults appear to remain in brood 430 groups within these areas (Larson et al., 2010). Similarly, some teleosts display a 431 preference for remaining in kin groups (Olsén, Petersson, Ragnarsson, Lundqvist, & 432 Järvi, 2004; Selkoe, Gaines, Caselle, & Warner, 2006; Sikkel & Fuller, 2010), showing 433 aggression towards unrelated individuals (Sikkel & Fuller, 2010), suggesting both 434 recognition of siblings and an active choice to maintain brood relationships. 435

436 Small genetic differences have been observed between interannual cohorts of
437 elasmobranchs at the same nursery site as well as between nursery sites within the
438 same year (Sampio, 2014). There have been observations of neonatal spurdog in

partially enclosed loch systems in Scotland (Thorburn et al., 2015) suggesting multiple pupping sites throughout the UK. The small genetic differentiation observed in the samples from Jura, which comprised of a high level of juvenile individuals, may be indicative of this in spurdog, although it was not observed in other areas where juveniles were sampled. If social cohesion occurs and lends structure to sub-groups of spurdog based on nursery site and these bonds are maintained throughout life, then this could create genetically distinct sub-units within a population.

Due to the high level of relatedness observed in the samples from Wales, such bonds 446 may be formed with related individuals. Yet, as spurdog aggregate, it is likely some 447 females give birth in the same locale, therefore any social drivers for maintenance of 448 relationships would occur between unrelated individuals as well as within broods, 449 450 which may explain why relatedness was within expected ranges for all other sampling locations including the temporally split Celtic Sea samples. The difference in sizes 451 between the groups samples in the Celtic Sea do suggest that they may be formed from 452 interannual cohorts, which may be causative of the differentiation between groups 453 observed. 454

It is worth noting that the markers used in this study are unlikely to be powerful enough to definitively detect siblings, whilst the lack of a common haplotype amongst samples identified as FS negates them all being full siblings. However, the above average level of relatedness does suggest some level of genetic relationship.

Suggested drivers for segregation in elasmobranch populations refer to differences between either the sexes (Economakis & Lobel, 1998; Hanchet, 1991; Klimley, 1987) or life stages (Lucifora, García, Menni, & Escalante, 2006); (Wearmouth & Sims, 2010). There is, therefore, no physiological reason why same sex individuals cannot remain associated throughout life from birth, assuming they are born at the same spatial-temporal point, an opportunity increased in live bearing species like spurdog (Hanchet, 1988), as drivers of segregation will act on these individuals concurrently.
The limited evidence of male-biased dispersal, a common feature of elasmobranch
populations (Daly-Engel et al., 2012; Pardini et al., 2001; Portnoy et al., 2010),
observed in the microsatellite data may give further opportunities for females to
maintain social groups.

The overall spatial homogeneity does suggest that even if spurdog do exist as 470 genetically distinct sub-populations, these groups are highly mobile, moving 471 throughout much of their range. Furthermore, it does not appear that the NE Atlantic 472 population experience behavioural separation, as sites where there is evidence of site 473 fidelity in spurdog are not genetically distinct. In fact, Loch Etive, a site where there is 474 limited evidence of site fidelity (Thorburn et al., 2015) had the highest number of 475 observed haplotypes, including all major UK haplotypes and the second highest 476 haplotype diversity and allelic richness, increasing the conservation potential of such 477 sites for the species. 478

What is unclear is if this spatial homogeneity is caused by complete admixture within 479 480 the spurdog population, or sub-groups of the population moving throughout the entirety of the species NE Atlantic range and a factor of the sampling regime. A lack of 481 temporal sampling, while the only option available over most of the area covered in 482 this study, may have led to a perception of homogeneity, as temporal genetic variation 483 can promote spatial homogeneity across sampling sites (Hedgecock, 1994). 484 485 Consequently a comprehensive understanding of spatial differentiation is contingent upon a more extensive and intensive sampling regime with synoptic sampling 486 487 throughout the region may produce more spatial heterogeneity to clarify this. A wider sampling area may also help investigations into Biogeographic regionality. 488

489

### 490 4.3 The Origin of NE Atlantic Spurdog

The presence of several common haplotypes suggests the contemporary population of 491 spurdog around the UK was derived from several populations, while the low levels of 492 sequence divergence suggest historical population expansion from a small effective 493 population size following a bottleneck, with no long-term geographic separation 494 (Avise, 2000). This contention is further supported by negative Tajima D and Fu's FS 495 values for each region (Fu 1997; Joshi, Salar, Banerjee, & others, 2013; Okello, 496 Nyakaana, Masembe, Siegismund, & Arctander, 2005; Tajima, 1989) and a smooth, 497 unimodal mismatch distribution (Harpending, 1994; Rogers & Harpending, 1992). 498 Such population formation may have occurred during rapid recolonization of the NE 499 Atlantic shelf during the current interglacial period. Interestingly, the weak genetic 500 differentiation observed between Rockall and other areas, coupled with a non-501 significant FS value tentatively suggest that there may have been a historically stable 502 offshore population of spurdog (or again, one buffered by the NW Atlantic). However, 503 the small samples size from Rockall does prevent any firm conclusions being drawn. 504 The geographic location of Rockall and the small irregularities observed in the genetic 505 data from this study suggest it is an area worthy of further investigation. 506

507

# 508 **5. Conclusions**

We suggest there is genetic mixing of spurdog in UK waters, supporting the notion that, for management purposes, spurdog should be treated as a single large stock unit. However, we suggest this assumption is treated with some caution as, while unambiguous spatial evidence to the contrary is unavailable, there are indications of population processes at a temporal and regional level that cause some smaller

genetically distinct groups. This suggests local management may be additionally 514 beneficial to the population rather than a single blanket management strategy. Local 515 spatial management at apparent key habitats could contribute towards the 516 conservation of the species as it would eliminate any bycatch of the species in that area. 517 It may also be important for contributing to the conservation of the species on a much 518 wider spatial scale as it appears some locations would offer protection to a 519 considerable proportion of the apparent NE Atlantic spurdog genetic resource. On a 520 more local scale, spatial closures could provide an opportunity to conserve all age 521 classes of both sexes (Thorburn et al., 2015) which is key to ensuring population 522 survival in this late maturing species (Kinney & Simpfendorfer, 2009). 523

Tools such as temporal closures may disproportionality affect some genetic sub-524 populations due to the type of temporal variation observed in the Celtic Sea and could 525 have significant implications for the region's genetic diversity. The effect of genetic 526 variation over time may be of importance to temporary closures and should be further 527 investigated in line with temporal management strategies, with special focus on the 528 effect of seasonality which, might indicate times when temporal management may be 529 most effective. Future genetic investigation should be conducted sampling locations at 530 multiple synoptic temporal points to clarify regional temporal genetic variation of 531 spurdog and the occurrence of groups of high relatedness. Identifying the times of 532 greatest stock integrity. 533

534 Our findings have implications for all elasmobranch fisheries, as the presence of 535 related groups and temporal variability in spurdog may be indicative of population 536 processes creating sub-structuring in other species. Therefore, until movement and 537 genetic studies including temporal samples can be integrated to define population 538 processes, a precautionary approach to management measures should address the 539 possibility of temporal stock variability and temporal differentiation should be 540 considered equally important as regional differentiation in management decisions.

One such measure could, theoretically, be, if commercial catch limits are reinstated, 541 the addition of a temporal element to the catch limit that spreads numbers taken over 542 a certain time to take individuals from several different genetic sub-units in an effort 543 maintain the high genetic diversity observed around the UK. This may, upon further 544 investigation, prove to be more beneficial to many other species of elasmobranch that 545 display similar genetic sub-grouping. There is also the suggestion that there may be 546 benefits from reassessment of historical tagging studies, with each spatial area looked 547 at on a temporal level. 548

549

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865 Table 1: Mitochondrial control region genetic diversity indices and results of neutrality 866 test (Tajima's D and Fu's FS) for all samples from each population of Squalus 867 acanthias (no neutrality test results for North Scotland due to low sample numbers). 868 Sampling locations as seen in Figure. 3.1: CELTIC = Celtic Sea, ETIVE = Loch Etive, JURA = Sound of Jura, S. N. SEA = Southern North Sea, N.SCO = North Scotland, 869 870 ROCKALL = Rockall, SUNART = Loch Sunart, WALES = Wales. N = number of 871 individuals, H = number of haplotypes, h = haplotype diversity, n = nucleotide diversity. Tajima's D = Tajima value with associated P-value in column to the left, Fu's 872 FS = Fu's FS value with associated P-value in the column to the left. 873

	Ν	Н	h	n	Tajima's D	P-value	Fu's FS	P-value
CELTIC	48	13	0.838	0.0022	-1.171	0.115	-5.64	0.006
Ετινε	65	23	0.936	0.0028	-0.745	0.249	-15.505	0
JURA	22	11	0.909	0.0024	-0.625	0.295	-5.551	0.001
S. N. SEA	33	17	0.917	0.0030	-0.738	0.252	-10.785	0
N.Sco	7	7	1	0.0026	-	-	-	-
ROCKALL	7	5	0.905	0.0035	0	0.519	-0.803	0.239
SUNART	33	17	0.939	0.0029	-0.414	0.385	-9.747	0
WALES	13	6	0.782	0.0015	-1.244	0.111	-2.375	0.021

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876 Table 2: Nuclear microsatellite (averaged over all loci) genetic diversity indices and 877 summary of the regional proportions of full sibling (FS) pairwise relationships between individuals for all samples from each population of Squalus acanthias. 878 879 Microsatellite diversity values with FS relationships (as identified by MLrelate) 880 removed are also shown (italic). Sampling locations as seen in Figure. 3.1: CELTIC = 881 Celtic Sea, ETIVE = Loch Etive, JURA = Sound of Jura, S. N. SEA = Southern North Sea, N.SCO = North Scotland, ROCKALL = Rockall, SUNART = Loch Sunart, WALES 882 883 = Wales. N = number of individuals,  $H_0$  = observed heterozygosity,  $H_e$  = expected heterozygosity, A = mean allelic richness, HWE = Hardy Weinberg Expectation P 884 values, F<sub>IS</sub> = Wright's inbreeding coefficient. Proportion of FS relationships between 885 886 different sampling regions (B), and between individuals from the same sampling region (W). Relationships where estimated using the Maximum likelihood method in 887 888 MLRelate.

	٨	V	Н	l <sub>o</sub>	h	le	A		HV	VE	F	IS	В	W
Celtic	76	64	0.59	0.6	0.64	0.65	4.35	4.5	0.01	0.091	0.088	0.073	0.030	0.032
Etive	69	40	0.58	0.57	0.64	0.66	4.46	4.6	0.000*	0.287	0.1	0.141	0.029	0.030
Jura	31	24	0.63	0.61	0.65	0.65	4.36	4.4	0.025	0.362	0.032	0.07	0.029	0.026
S. N. Sea	48	29	0.6	0.59	0.66	0.68	4.36	4.5	0.000*	0.000*	0.083	0.133	0.029	0.048
N.Sco	13	13	0.7	0.7	0.68	0.68	4.78	4.8	0.519	0.519	-0.03	-0.03	0.028	0
Rockall	8	7	0.65	0.65	0.62	0.62	3.91	3.9	0.697	0.697	-0.03	-0.03	0.034	0.071
Sunart	33	26	0.6	0.62	0.64	0.65	4.2	4.3	0.017	0.206	0.06	0.047	0.029	0.025
Wales	1 <u>66</u>	1 <u>00</u>	0.69	0.64	0.62	0.66	3.8	4.1	0.000*	0.147	-0.11	0.024	0.030	0.368

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890 Table 3: Mitochondrial  $\Phi$ ST (top panel, below the diagonal), nuclear microsatellite 891 FST (middle panel, below the diagonal) and nuclear microsatellite FST with 'FS' 892 relationships (as identified by MLrelate) removed (bottom panel, below the diagonal) of genetic differentiation between areas removed among 8 sampling localities of 893 spurdog Squalus acanthias from around the UK. Associated p-values are above the 894 895 diagonal in all panels. \*\*significant after Bonferroni sequential correction. \*significant after fdr correction. Significant FST values and related p-valeus are highlighted in bold 896 text. Sample regions legend: JURA = Sound of Jura; CELTIC = Celtic Sea; SUNART = 897 898 Loch Sunart; ROCKALL = Rockall; ETIVE = Loch Etive; N.SCO = North Scotland; WALES = Wales; S.N.SEA = Southern North Sea. 899

	CELTIC	ETIVE	JURA	S.N.SEA	N.SCO	ROCKALL	SUNART	WALES
CELTIC		0.1265	0.0141*	0.1743	0.4809	0.0052*	0.4039	0.3434
ETIVE	0.0113		0.1479	0.1758	0.9780	0.0024*	0.2986	0.0435
JURA	0.0605	0.0162		0.2942	0.6400	0**	0.0132*	0.0125*
S.N.SEA	0.0108	0.0093	0.0062		0.6913	0.0049*	0.1014	0.1890
N.SCO	-	-0.0602	-0.0325	-0.0302		0.0127*	0.6545	0.0659
ROCKALL	0.1551	0.1727	0.2681	0.1587	0.1917		0.0296	0.0378
SUNART	-	0.0033	0.0617	0.0199	-0.0268	0.0987		0.2413
WALES	0.0044	0.0552	0.1138	0.0215	0.1048	0.1068	0.0145	
CELTIC		0.1622	0.9203	0.0844	0.7426	0.6495	0.7793	0.0015**
ETIVE	0.0029		0.1939	0.2719	0.2559	0.1328	0.8194	0.0003**
JURA	-	0.0041		0.1815	0.6878	0.4047	0.7989	0.0022*
S.N.SEA	0.0051	0.0023	0.0044		0.6023	0.5235	0.4570	0.0005**
N.SCO	-	0.0059	-0.0042	-0.0014		0.9311	0.5911	0.0826
ROCKALL	-	0.0162	0.0024	0.0001	-0.0187		0.5943	0.0350
SUNART	-	-0.0026	-0.0037	0.0007	-0.0015	-0.0029		0.0006**
WALES	0.0285	0.0378	0.0305	0.0340	0.0146	0.0323	0.0412	
CELTIC		0.7877	0.9740	0.4369	0.8997	0.6607	0.9710	0.6329
ETIVE	-		0.6774	0.841	0.6585	0.2686	0.9998	0.4115
JURA	- 0.0068	-0.0012		0.6538	0.8695	0.7202	0.8662	0.7263
S.N.SEA	0.0013	-0.0029	-0.0013		0.8028	0.6215	0.9075	0.5714
N.SCO	- 0.0076	-0.0011	-0.0081	-0.0048		0.9315	0.7706	0.8116
ROCKALL	- 0.0035	0.0111	-0.0064	-0.0011	-0.0187		0.5527	0.4510
SUNART	- 0.0061	-0.0107	-0.0056	-0.0057	-0.0054	-0.0020		0.4471
WALES	- 0.0023	0.0051	-0.0054	0.0004	-0.0103	0.0034	0.0020	

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Table 4: Temporal sampling details for the four hauls form the Celtic Sea. Including
group name, Date of haul, sex ratio and size range of caught spurdog. Details from
Celtic sea temporal sampling incidences. FS: Full sibling relationships, HS: Half
sibling relationships, PO: Parental offspring and U: unrelated.

Group	Date	Males	Females	Size range	FS	HS	PO	U
				(cm)				
2010	03 Oct 2010	10	8	75 - 109	0.046	0.177	0.052	0.725
2010a	12 Dec 2010	1	18	93 - 116	0.039	0.137	0.078	0.745
2011	21 Aug 2011	79	58	58 – 107	0.040	0.110	0.053	0.797
2011a	21 Sep 2011	5	10	64 - 108	0.038	0.124	0.067	0.777

Table 5: FST value (below diagonal) and associated P-values of genetic differentiation
between the temporal sampling occasions within the Celtic sea with FS relationships
removed. \*\*Significant after Bonferroni sequential correction, \*significant after fdr
correction. For details of samples from each group, refer to Table 7.

	CEL2010	CEL2010a	CEL2011	CEL2011a
CEL2010		0.0076**	0.3754	0.0599
CEL2010a	0.03722		0.0109*	0.7568
CEL2011	0.00353	0.02827		0.5429
CEL2011a	0.02073	-0.00629	-0.00029	

Table 6: Comparisons of inbreeding coefficient (FIS), fixation index FST, relatedness (REL), =
mean assignment indices (mAIc) and variance of assignment index (vAIc) between
male (M) and female (F) spurdog from UK waters as indicators of sex biased dispersal.
Significant differences are indicated by a p-value.

920		FIS	FST	REL	mAIC	vAIC	
921	М	0.024	-0.01	-0.019	-0.15778	10.00471	
000	F	0.1285	0.0045	0.0079	0.12335	5.15851	
922 923	Р	0.9818	0.1596	0.1512	0.3224	0.0185	

Figure 1: Sampling locations for spurdog around the UK. Insert shows the three sampling sites from the west coast of Scotland. Total number of samples from which DNA was extracted from each sampling site: Celtic Sea = **48**, *76*; Loch Etive = **65**, *69*; Sound of Jura = **22**, *31*; Southern North Sea = **33**, *48*; North Scotland = *7*, *13*; Rockall = *7*, *8*; Loch Sunart = **33**, *33*; Wales = **12**, *17*. Total = **228**, *295*. Bold numbers indicate samples sequenced for mitochondrial analysis, italics indicate number of samples genotyped for microsatellites.

Figure 2: Haplotype network of the mitochondrial DNA control region sequences for 931 the spiny dogfish Squalus acanthias indicating lack of spatial population structure in 932 the NE Atlantic. Haplotypes are represented by circles with sizes proportional to 933 absolute frequency in the total sample. Colour codes are given of sample regions: 934 JURA = Sound of Jura; CELTIC = Celtic Sea; SUNART = Loch Sunart; ROCKALL = 935 936 Rockall; ETIVE = Loch Etive; N.SCO = North Scotland; WALES = Wales; S.N.SEA = Southern North Sea. All branches correspond to one nucleotide substitution between 937 haplotypes. Blue dots represent missing haplotypes. 938

Figure 3: Factorial correspondence analysis plot of significant FST differences using
all 7 µSAT loci before related individuals were removed (Table 3). All samples are
connected to the population mean. Sample regions legend: JURA = Sound of Jura;
CELTIC = Celtic Sea; SUNART = Loch Sunart; ETIVE = Loch Etive; WALES = Wales;
S.N.SEA = Southern North Sea.

Figure 4: Factorial correspondence analysis plot of spurdog µSAT data from Table 5with FS relationships removed. All points are linked to the populations mean.

946 Figure 5: Mean relatedness (r) within 8 populations of spurdog from NE Atlantic based

947 on pairwise comparisons. Mean values refer to the mean level of relatedness within

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949	'No difference' across the populations. Sample regions legend: JURA = Sound of Jura;
950	CELTIC = Celtic Sea; SUNART = Loch Sunart; ROCKALL = Rockall; ETIVE = Loch
951	Etive; N.SCO = North Scotland; WALES = Wales; S.N.SEA = Southern North Sea.
952	Figure 2: Observed and expected mismatch distribution of mtDNA haplotypes from
953	samples of spurdog ( <i>Squalus acanthias</i> ) in UK waters.
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the population. U & L refer to the 95% confidence intervals around the null hypothesis