ORIGINAL RESEARCH

Revised: 31 October 2018

WILEY Ecology and Evolution

Variations in age- and sex-specific survival rates help explain population trend in a discrete marine mammal population

Mònica Arso Civil¹ Barbara Cheney² | Nicola J. Quick^{1,3} | Valentina Islas-Villanueva^{1,4} | Jeff A. Graves⁵ | Vincent M. Janik¹ | Paul M. Thompson² | Philip S. Hammond¹

¹Sea Mammal Research Unit, Scottish Oceans Institute, University of St Andrews, St Andrews, UK

²Lighthouse Field Station, Institute of Biological and Environmental Sciences, University of Aberdeen, Cromarty, UK

³Duke University Marine Laboratory, Nicholas School of the Environment, Beaufort, North Carolina

⁴CONACYT, Universidad del Mar, Instituto de Genética, Ciudad Universitaria, Oaxaca, México

⁵Centre for Biological Diversity, University of St Andrews, St Andrews, UK

Correspondence

Arso Civil Mònica, Sea Mammal Research Unit, Scottish Oceans Institute, University of St Andrews, St Andrews, UK. Email: mac64@st-andrews.ac.uk

Funding information Department of Energy and Climate Change

Abstract

Understanding the drivers underlying fluctuations in the size of animal populations is central to ecology, conservation biology, and wildlife management. Reliable estimates of survival probabilities are key to population viability assessments, and patterns of variation in survival can help inferring the causal factors behind detected changes in population size. We investigated whether variation in age- and sex-specific survival probabilities could help explain the increasing trend in population size detected in a small, discrete population of bottlenose dolphins Tursiops truncatus off the east coast of Scotland. To estimate annual survival probabilities, we applied capture-recapture models to photoidentification data collected from 1989 to 2015. We used robust design models accounting for temporary emigration to estimate juvenile and adult survival, multistate models to estimate sex-specific survival, and age models to estimate calf survival. We found strong support for an increase in juvenile/ adult annual survival from 93.1% to 96.0% over the study period, most likely caused by a change in juvenile survival. Examination of sex-specific variation showed weaker support for this trend being a result of increasing female survival, which was overall higher than for males and animals of unknown sex. Calf survival was lower in the first than second year; a bias in estimating third-year survival will likely exist in similar studies. There was some support first-born calf survival being lower than for calves born subsequently. Coastal marine mammal populations are subject to the impacts of environmental change, increasing anthropogenic disturbance and the effects of management measures. Survival estimates are essential to improve our understanding of population dynamics and help predict how future pressures may impact populations, but obtaining robust information on the life history of long-lived species is challenging. Our study illustrates how knowledge of survival can be increased by applying a robust analytical framework to photoidentification data.

KEYWORDS

calf survival, capture-recapture, mortality rate, population dynamics, sex-specific survival, *Tursiops*

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2018 The Authors. Ecology and Evolution published by John Wiley & Sons Ltd.

1 | INTRODUCTION

Knowing whether populations are increasing or declining and understanding the drivers behind such fluctuations are important issues in ecology, conservation biology and wildlife management (Eberhardt, 1985: Galimberti, Sanvito, Boitani, & Fabiani, 2001: Williams, Nichols, & Conroy, 2002). Patterns in the variation of survival rates are important indicators in population dynamics (e.g., Kraus et al., 2005; Altwegg, Crawford, Underhill, & Williams, 2008), and reliable estimates of survival probabilities are required to assess and determine the viability of populations (Doak, Kareiva, & Klepetka, 1994; White, Franklin, & Shenk, 2002). Mammalian age-specific survival is typified by a U-shaped mortality curve, characterized by high rates in young animals and low rates in adults that increase toward the maximum age (Caughley, 1966). Males tend to have higher mortality rates than females (Trivers, 1972). Variations in age- and sex-specific survival rates modulate population dynamics; adult survival typically has a greater effect on population trends than juvenile survival (Eberhardt, 2002; Fowler, 1987).

In a conservation context, monitoring to determine trends in population size is crucial to inform assessments of wildlife populations (e.g., IUCN, 2017). The cause of a trend may be understood from knowledge of human-induced mortality, for example, but in other cases the cause may not be so easily revealed. In such cases, estimates of fecundity and survival can play an important role in helping to understand which element(s) of life history may be responsible for changes in population size (e.g., Gaillard, Festa-Bianchet, & Yoccoz, 1998; Currey et al., 2011).

Long-term individual-based studies are an effective tool to estimate population parameters from discrete populations in which individuals can be repeatedly captured over time (Clutton-Brock and Sheldon, 2010), and are particularly important in long-lived species for which the ability to detect changes in demographic parameters may require decades of data. Capture-recapture analyses of individual-based data have been used extensively in ecology (Burnham, Anderson, White, Brownie, & Pollock, 1987; Cormack, 1964) to estimate survival probabilities (e.g., Gaillard et al., 1998; Altwegg et al., 2008). Obtaining robust estimates of survival probabilities in cetacean populations remains challenging, and estimates of age- and sex-specific survival are scarce (e.g., Bradford et al., 2006; Currey et al., 2009; Ramp, Bérubé, Palsboll, Hagen, & Sears, 2010).

Our focus here is the east coast of Scotland bottlenose dolphin (*Tursiops truncatus*) population (Figure 1), which long-term photoidentification monitoring since 1989 indicates is increasing, especially since around 2000 (Cheney, Graham, Barton, Hammond, & Thompson, 2018; Wilson, Hammond, & Thompson, 1999; Wilson, Reid, Grellier, Thompson, & Hammond, 2004). Survival rates have been estimated for this population using eight (Sanders-Reed, Hammond, Grellier, & Thompson, 1999) and 13 (Corkrey et al., 2008) years of data collected in the 1990s to early 2000s. Data for both studies indicated a greater probability of a population decline than of an increase. However, this population expanded its distributional range during the 1990s (Wilson et al., 2004), meaning that these



FIGURE 1 Bottlenose dolphins from the east coast of Scotland (photograph by Mònica Arso Civil, taken during this study)

estimated declines are likely to be confounded with temporary emigration (Corkrey et al., 2008).

Here, we use a 27 year-long dataset of individual capture histories to investigate variations in age- and sex-specific survival rates in this small, discrete population of bottlenose dolphins. We explore whether changes in juvenile/adult survival could help explain the observed increase in population size and investigate variation in calf survival. We first estimate annual survival of juvenile/adult dolphins using robust design capture-recapture models, which incorporate the estimation of temporary emigration, including assessing the support for a trend over time. We then explore whether there is evidence for sex-specific survival using multistate capture-recapture models. We estimate calf survival during the first 3 years of life for a subset of dolphins followed since their year of birth using agespecific models, and investigate whether survivorship of first-born calves was different from calves born subsequently.

2 | MATERIALS AND METHODS

2.1 | Photoidentification data

Boat-based surveys were conducted off the east coast of Scotland (Figure 2) between May and September every year from 1989 to 2015 to collect photoidentification data of bottlenose dolphins following standardized protocols (Cheney et al., 2014; Islas-Villanueva, 2009; Quick & Janik, 2008). Sampling effort occurred in two main areas at opposite ends of the population's main distributional range; in the Moray Firth Special Area of Conservation (SAC) effort was consistent over the study period (Cheney et al., 2014), while in St Andrews Bay and the Tay estuary effort was variable from 1997 to 2007 and consistent from 2009 onwards (Chenev et al., 2013: Islas-Villanueva, 2009; Quick & Janik, 2008). Individual dolphins were identified from high-quality photographs (Wilson et al., 1999) based on unique markings in the dorsal fin (Würsig & Jefferson, 1990), and matched against a catalogue of previously identified bottlenose dolphins from the east coast of Scotland. Because bottlenose dolphin calves do not tend to have permanent marks that can be tracked across years, individuals in the first 3 years of life were aged and identified based on body size, skin coloration, presence of fetal



FIGURE 2 Map showing the two main study areas (circles) located in the Moray Firth Special Area of Conservation (SAC; dashed area) and in St Andrews Bay and the Tay estuary

folds and repeated association with a known adult dolphin (i.e., the mother; Grellier, Hammond, Wilson, Sanders-Reed, & Thompson, 2003; Arso Civil, Cheney, Quick, Thompson, & Hammond, 2017). Capture histories of marked juveniles (at least 4 years of age) and adults (i.e., dolphins with distinctive long-lasting nicks on the trailing edge of the dorsal fin) and of calves were constructed to model survival probabilities. The sex of individual dolphins was determined from photographs of the genital area, molecular analysis of biopsy samples (Islas-Villanueva, 2009) or the identification of mother-calf pairs.

2.2 | Modeling age- and sex-specific survival

The open population Cormack-Jolly-Seber (CJS) model is typically used to estimate survival probabilities from capture-recapture data (e.g., Pollock, Nichols, Brownie, & Hines, 1990). Capture-recapture models rely on a number of assumptions about the captured individuals and about the probability of capture, which can generate bias in the estimated model parameters if violated (Hammond, 2010; Pollock et al., 1990). Conventional open population models assume, among other things, that any emigration from the study population is permanent (Kendall, Nichols, & Hines, 1997), an assumption that is violated when animals transit through an area and are not available for capture after first release (Pradel, Hines, Lebreton, & Nichols, 1997) or when animals range beyond the study area and are unavailable for capture on some occasions (i.e., temporary emigration; Kendall et al., 1997).

Our study population expanded its distributional range in the 1990s (Wilson et al., 2004), but sampling effort only gradually expanded outside the initial study area from the end of the 1990s. WILEY_Ecology and Evolution

Combined with variability in movement patterns among individuals in the population (Cheney et al., 2013), this means that temporary emigration is likely to be a feature of our study, as suggested by Corkrey et al. (2008). Failure to account for this may lead to bias. Methods to determine the sex of individuals can also be a source of bias; in cetaceans assigning sex reliably typically requires repeatedly resighting individuals, positively biasing the survival of sexed individuals simply because they have been sighted for longer than unsexed animals (Nichols, Kendall, Hines, & Spendelow, 2004).

Prior to survival modeling, the key assumptions about the probability of capture (Lebreton, Burnham, Clobert, & Anderson, 1992) were explored in each dataset using goodness-of-fit tests in program U-CARE (Choquet, Lebreton, Gimenez, Reboulet, & Pradel, 2009). Assumptions were satisfied for the calf and the multistate (sex-specific) juvenile/adult datasets, but showed evidence for trap-dependence and transience in the single-state juvenile/adult dataset (see below for dataset details), stressing the need to use a modeling approach which could incorporate temporary emigration and heterogeneity in the capture probabilities. A small level of overdispersion was detected in the single-state juvenile/adult dataset, and model statistics were adjusted by the estimated overdispersion factor (ĉ = 1.629; Choquet et al., 2009). Model selection was based on the Akaike Information Criterion (AIC; Akaike, 1973) adjusted for small samples (AICc) for the calf and the multistate juvenile/adult datasets, and on the Quasi Akaike Information Criterion (QAIC_c) for the single-state juvenile/adult dataset (Burnham & Anderson, 2002). Model structures and parameters were specified and run using the package RMark (Laake, 2013) in R (R Core Team, 2016), and program MARK (White & Burnham, 1999).

2.2.1 | Juvenile/Adult survival: Robust design models

Robust design (RD) models (Kendall & Nichols, 1995; Kendall et al., 1997; Pollock, 1982) were used to estimate survival probabilities of juvenile/adult dolphins (i.e., non-calf), combining open and closed population models with estimators that incorporate temporary emigration. Each annual field season represented a primary sampling occasion and months within each season were treated as secondary sampling occasions. Parameters estimated by RD models include the probabilities of survival, temporary emigration, capture and recapture. Survival was first set to be constant in all models to obtain an average survival estimate for juveniles/adults.

The probability of temporarily emigrating was modeled as the probability that an animal was unavailable for capture during a primary period (i.e., a given year), given that it was available (γ ") or unavailable (γ ") in the previous primary period. Random temporary emigration is characterized by the probability of emigrating not depending on whether or not an animal was previously available (γ " = γ '). Markovian temporary emigration occurs when the probability of emigrating depends on whether an animal was previously available or not (γ " $\neq \gamma$ '). Models with random or Markovian temporary emigration were fitted, with probabilities allowed to be constant or

time-dependent, in which case constraints were applied to allow identifiability of parameters (Kendall et al., 1997). No-movement models (γ '' = γ ' = 0) were included for comparison.

Recapture probability was assumed to equal capture probability for all models because photoidentification does not involve handling of the animals and is not expected to cause any behavioral changes in the captured individuals. Capture probabilities were allowed to vary over both primary and secondary periods in all models because preliminary analysis showed less support for models restricting capture probabilities to vary between years but not within year, or not to vary at all. Models incorporating individual heterogeneity were also considered (π ; Pledger, 2000), in which the population was assumed to comprise a mixture of two types of individual with different probability of capture, with probability π of being in the first type. Using the most supported model regarding capture and temporary emigration probabilities, a trend in survival probability over the study period was then explored.

2.2.2 | Sex-specific survival: Multistate models

To account for uncertainty in sex assignment and thus reduce bias in estimates of survival (Nichols et al., 2004; Pradel et al., 2008), multistate (MS) models (e.g., Hestbeck, Nichols, & Malecki, 1991; Brownie, Hines, Nichols, Pollock, & Hestbeck, 1993) were used to estimate the sex-specific probability of survival of juvenile/adult dolphins. Sightings of individuals made during the same year were pooled together in one sampling occasion. States were defined as male (M), female (F), or unknown (U) if sex had not been determined. When first sighted, an individual was recorded as unknown (U) until the sex was determined (M or F), and then remained in that state for all following re-sightings. Parameters estimated in the MS models included the probabilities of recapture and survival by state (i.e., by sex) and transition probabilities between states. Individuals could only transition from state U to either state M or F, fixing all other transition probabilities at zero. Transition probabilities in years when no males or females were sexed were fixed at zero. A set of candidate models was created to investigate the effects of no variation, time-dependence, time trend, and state (sex-specific: $M \neq F \neq U$, or the combinations $M = U \neq F$, F = U≠ M) on survival, recapture and transition probabilities. Models with constant transition probabilities over time were not considered because the number of animals sexed varied markedly among years.

2.2.3 | Calf survival: Age-specific CJS models

Age-specific CJS models were used to estimate the probability of survival of dolphins during the first 3 years of life. Because the probability of sighting a calf was highly dependent on sighting the mother (Grellier et al., 2003), we only included calves with a known mother and year of birth, and whose mothers were seen in the year of birth and in the three subsequent years. Recapture probabilities were allowed to be constant or to vary among



FIGURE 3 Time trend in juvenile/ adult estimated survival probability (with associated 95% confidence intervals) from the most supported RD model

years. Survival was modeled as constant for all ages, separately for 1st, 2nd and 3rd year, or for 1st year and 2nd/3rd year as a single parameter. Models were also fitted to investigate whether survivorship of first-born calves was different from calves born subsequently.

3 RESULTS

3.1 | Juvenile/Adult survival

In total, 205 iuvenile/adult marked bottlenose dolphins were identified between 1989 and 2015. Model selection favored models incorporating Pledger (2000) heterogeneity mixture parameters over those without it. Including a trend in the probability of survival (model 1 in Supporting Information Table S1) greatly improved the model fit (Δ QAICc = 9.6, compared to the equivalent model with time-invariant survival, model 2). This model had Markovian temporary emigration with a very low constant probability of emigrating (γ ") of 0.017 (95% CI: 0.006–0.047) and a high constant probability of remaining an emigrant (γ') of 0.712 (95% CI: 0.358-0.915). The estimated survival probability increased from 0.931 (95% CI 0.886-0.958) in 1990 to 0.960 (95% CI 0.932-0.977) in 2015 (Figure 3). The next best-supported model incorporated a constant survival probability of 0.948 (95% CI 0.933-0.959; model 2 in Supporting Information Table S1) and very similar estimates of Markovian emigration to the model with a time trend in survival.

To investigate if the detected trend in survival was more likely to be a result of a change in adult or juvenile survival, the top two most supported models in Supporting Information Table S1 were fitted to a restricted dataset of adults only. This dataset excluded the first 6 years of sightings for those dolphins classified as juveniles when first seen, and the first 3 years of sightings for those classified as subadults (based on body size). For this adult-only dataset, the model with constant survival probability received more support $(\Delta QAICc = 4.4)$ compared to the one that included a trend in survival.

3.2 | Sex-specific survival

The marked juvenile/adult dolphins identified between 1989 and 2015 included 43 males, 66 females and 96 animals of unknown sex. Multistate (MS) models that allowed transition and capture probabilities to vary among years but not between sexes had better support from the data than other models (Supporting Information Table S2). In the top eight MS models, all within a \triangle AICc < 4 indicating some support from the data, survival was parameterized in a variety of ways including constant, sex-specific, and with and without a temporal trend. Within this suite of models, there was most support for survival being different between females and males/unknown sex but the model with constant survival also received considerable support (Supporting Information Table S2). Models incorporating a sex-specific temporal trend in survival had relatively weak support (Supporting Information Table S2) but showed a more pronounced trend in survival for females than for males and animals of unknown sex (Figure 4). Estimated survival was higher for females (0.962; 95% CI 0.941-0.976) than males (0.942; 95% CI 0.904-0.966; Table 1).

3.3 | Calf survival

In total, 116 calves with known year of birth and known mother were identified between 1990 and 2015. Of these, 56 calves had mothers sighted in the 3 years following birth. Models restricting the recapture probability to be invariant received a lot more support than those allowing it to vary among years and ages, resulting in a very high recapture probability of 0.954 (95% CI: 0.914-0.975, model 1 in Supporting Information Table S3). Models in which survival probability was constant, different for each age class or different between age 1 and ages 2/3 combined were all well supported by the data (models 1-3).





FIGURE 4 Time trend in sex-specific estimated survival probabilities with associated 95% confidence intervals for juvenile/adult bottlenose dolphins. The estimated parameters are derived from multistate model number 5 in Supporting Information Table S2

Survival probabilities of calves were lowest in the 1st year, increased in the 2nd year and decreased in the 3rd year (Table 1). Models in which the survival of first-born calves was different from calves born subsequently received considerable support from the data but not as much as models that did not incorporate this (models 4 and 5 in Supporting Information Table S3). Estimated survival of first-born calves was lower than that of later-born calves at any age (Table 1).

4 | DISCUSSION

In this analysis of age- and sex-specific survival in a small, discrete coastal bottlenose dolphin population, we present evidence for variation in juvenile/adult survivorship over two and a half decades that is consistent with an increasing population. We also provide the first estimates of calf survival probabilities for this population, adding to the very few reliable estimates available for this or any other cetacean species.

4.1 | Increasing juvenile/adult survival

Our estimate of time-invariant survival probability for juveniles/ adults in the Scottish east coast bottlenose dolphin population (0.948, 95% CI: 0.933-0.960) is comparable to estimates reported for these age classes in other coastal bottlenose dolphin populations (Currey et al., 2011; Smith, Pollock, Waples, Bradley, & Bejder, 2013; Speakman, Lane, Schwacke, Fair, & Zolman, 2010). Our estimate is higher than those previously reported for our population by Sanders-Reed et al. (1999; 0.942, SE = 0.015) and Corkrey et al. (2008) (0.93, SE = 0.029), using data from 1990-1997 and 1990-2002, respectively; both of these studies suggested that a population decline was more probable than an increase. However, any prediction of decline was likely confounded with temporary emigration (Corkrey et al., 2008) because of the population's range expansion (Wilson et al., 2004). Our analysis is based on a much larger dataset (27 years compared to 13 years) that encompasses changes in the population's range and the subsequent changes in sampling effort. By explicitly

Age class	Group	Modeling approach	Survival	95% CI
Juvenile/adult	All	Robust design	0.948	0.933-0.959
	Female	Multistate	0.962	0.941-0.976
	Male		0.942	0.904-0.966
	Unknown sex		0.939	0.923-0.952
Calf—1st year	All	Age-CJS	0.865	0.785-0.919
Calf—2nd year			0.981	0.797-0.998
Calf—3rd year			0.883	0.708-0.959
Calf—1st year	First-born		0.836	0.697-0.918
	Subsequently born		0.880	0.789-0.935
Calf—2nd year	First-born		0.976	0.744-0.998
	Subsequently born		0.983	0.814-0.999
Calf–3rd year	First-born		0.853	0.619-0.954
	Subsequently born		0.894	0.725-0.964

TABLE 1 Estimates of time-invariant survival probability with associated 95% Confidence Interval (CI) from the best-supported models of age-specific survival, sex-specific survival and the survival of first- or subsequently born calves (model details given in Supporting Information Table S1–S3)

-WILEY

modeling temporary emigration, we minimized bias in the estimates of survival probability that could result by ignoring this.

It is generally accepted that in slow-growing populations of longlived species, adult survival has more influence on population growth than reproduction (Crone, 2001; Oli & Dobson, 2003). In a number of cetacean populations, trends in probabilities of survival have been linked to changes in population size (e.g., Kraus et al., 2005; Ford, Ellis, Olesiuk, & Balcomb, 2010; Currey et al., 2011; Gormley et al., 2012). Our results showed very strong support for an increase in juvenile/adult survival over two and a half decades, most likely caused by a change in juvenile survival, which translates into a 45% decrease in mortality rate (from 6.9% in 1990 to 4.0% in 2015). This change is likely to be responsible, at least partially, for the increase in population size identified by Cheney et al. (2014).

However, some studies highlight the importance of reproduction versus adult survival in shaping population dynamics (Manlik et al., 2016; Mitchell, Pacifici, Grand, & Powell, 2009). Arso Civil et al. (2017) estimated a fecundity rate for the study population similar to those reported for other stable bottlenose dolphin populations but Cheney (2017) found an increasing trend in reproductive rates using a restricted dataset of reproductive females utilizing the Moray Firth SAC between 2001 to 2016. Therefore, a parallel change in fecundity rate cannot be ruled out.

What could have caused the observed increase in survival probability? A decline in the use of illegal gill nets in the inner Moray Firth, which was an identified cause of mortality for this population during the 1990s (Butler et al., 2017), could have contributed to the increased survival, as identified for Hector's dolphins (Cephalorhynchus hectori) in New Zealand (Gormley et al., 2012). Wilson et al. (2004) hypothesized that the population's range expansion in the 1990s was more likely to be explained by changes in prey resources than an increase in population size, partially because of the apparent negative population growth at the time. This is supported by more recent studies that have identified high use areas, including foraging areas, outside the inner Moray Firth, which was the main area of distribution before the range expansion (Arso Civil, 2014; Pirotta et al., 2013). Also, dolphin movements across the distributional range are variable, with lower rates of exchange between the most geographically distant areas (Cheney et al., 2013). If, as seems likely, the range expansion has resulted in more prey resources being available to the population, this could have led to improved fitness of individual dolphins, and, consequently, an improved probability of survival in the population.

Demographic parameters such as survival and reproductive rates may change in other populations of mobile species as a consequence of environmental change, disturbance or management action, which may result in changes in the availability of or access to prey resources and/or a reduction in anthropogenic mortality sources. A reduction in foraging activity due to human pressure (e.g., Pirotta, Merchant, Thompson, Barton, & Lusseau, 2015, Williams, Lusseau, & Hammond, 2006) or a displacement of individuals from foraging areas (Pirotta et al., 2013) that affects their energy intake could have long-term population consequences through changes in individual vital rates (New et al., 2013). Changes in prey availability caused by environmental variability have been linked to changes in the survival and reproduction rates of killer whales (*Orcinus orca*) (e.g., Ford et al., 2010, Ward, Holmes, & Balcomb, 2009), while anthropogenic modifications to habitat have been linked to in increased age-specific mortality rates in another population of bottlenose dolphins (e.g., Currey et al., 2011). A decrease in mortality pressure due to management actions can result in significant changes in survival (e.g., Gormley et al., 2012) and increases in population size, as in the case of some whale populations recovering from past overexploitation as they reoccupy territories and exploit existing resources (Branch, Matsuoka, & Miyashita, 2004; Pallin et al., 2018).

4.2 | Sex-specific survival

Available estimates of sex-specific survival/mortality for bottlenose dolphins are scarce but all suggest higher mortality rates for males (e.g., Scott, Wells, & Irvine, 1990; Stolen & Barlow, 2003; Currey et al., 2009). Our estimate of male mortality rate (5.8% per annum) was almost double that for females (3.8% p.a.; Table 1) but, although sexspecific survival models were most supported by the data, support for them was not particularly strong. Mortality rate, calculated here as 1 - apparent survival rate, is a combination of the true mortality and any permanent emigration confounded with mortality. However, there is no evidence of permanent emigration in this population. Comparison of photoidentification data between the east and west coasts of Scotland showed no matches, suggesting no permanent or temporary movement of animals between those two areas (Cheney et al., 2013). Long-distance movements reported by Robinson et al. (2012) between the east and west coast of Scotland and Ireland were of an isolated group of individuals never seen in the east coast previously or subsequently to their encounter and were thus never part of the east coast of Scotland catalogue. These results are in accordance with those from genetic analysis which show some but not complete isolation between animals in this population and those found elsewhere in Britain (Parsons, Noble, Reid, & Thompson, 2002; Thompson et al., 2011).

Sex-differentiated mortality rates in mammals have been mainly attributed to sexual selection costs (Promislow, 1992; Trivers, 1972). In cetacean species, these include costs associated with male selection in polygynous mating systems (Ralls, Brownell, & Ballou, 1980), sex differences in ranging patterns (Stolen & Barlow, 2003), and sex differences in the accumulation of toxic burdens with age (Schwacke et al., 2002). Male bottlenose dolphins in the study population show a significantly higher percentage of body scarring and dorsal fin nicks compared to females (Marley, Cheney, & Thompson, 2013), which has been related to male-male competition (Scott, Mann, Watson-capps, Sargeant, & Connor, 2005). If males move spatially and temporally more than females, as observed in other bottlenose dolphin populations (Möller & Beheregaray, 2004; Scott et al., 1990), it could compromise their fitness and hence survival, assuming an increase in energetic costs. Jepson et al. (2016) report higher levels of polychlorinated biphenyls (PCBs) from stranded and biopsied male -WILEV_Ecology and Evolution

ARSO CIVIL ET AL.

bottlenose dolphins from the NE Atlantic, compared to females, which transfer part of their toxic burden to their offspring through lactation, although the exact role of contaminants in the survival of marine mammals remains unclear (Ross, 2002).

Our results also showed some support for a female-driven increase in juvenile/adult survival, which suggests that the hypothesized improved fitness in juveniles/adults following the range expansion was more pronounced in females compared to males and dolphins of unknown sex. The latter group is likely to include a higher proportion of juveniles and adult males than females because of sex assignment based on photographs. Estimating sexual selection costs is challenging but sex differences in ranging patterns could be investigated to improve understanding of why there may have been a sex-differentiated trend in survival in this population.

4.3 | Calf survival

First-year survival estimates are available for a few well-studied bottlenose dolphin populations (Currey et al., 2009; Mann, Connor, Barre, & Heithaus, 2000; Stolen & Barlow, 2003; Wells & Scott, 1990) and compare well with our estimate (0.865, 95% CI: 0.785-0.919). We found that second-year survival increased to 0.981 but third-year survival decreased to 0.708, which would not be expected if the age-specific mortality curve that is characteristic of long-lived mammals were strictly U-shaped (Caughley, 1966). This drop in estimated survival in the third year could be a result of increased mortality when calves are weaned and become independent from their mothers, which starts to occur in their third year of life (Grellier et al., 2003; Mann et al., 2000). Alternatively, it could be a result of mark loss. Young bottlenose dolphins have typically not yet acquired permanent marks; a three-year old calf associating less with its mother (Grellier et al., 2003) could be lost from the marked population because it was unable to be identified. Three-year inter-birth intervals occur less than 20% of the time in this population (Arso Civil et al., 2017) but the mothers of nine of the sixteen calves not captured in their third year of life were sighted that year accompanied by a new calf. The extent to which our estimate of third-year survival is negatively biased by mark loss is unknown but obtaining estimates of age-specific survival probabilities in older calves is likely to be problematic in other studies relying on photoidentification to recognize individuals.

Censoring the data to include only calves whose mothers were seen during the 3 years after calf birth could overestimate calf survival, as calves are very likely to die if the mother dies in their first year, and more likely to die if the mother dies in their second year and before weaning (Noren & Edwards, 2007). The maximum bias in first- or second-year calf survival would be equivalent to the mortality rate of adult females (3.8%), which would result in a first-year calf survival as low as 0.832.

Cheney, Wells, Barton, and Thompson (2017) have shown that first-born calves in the study population are shorter in length than calves born subsequently and found weak evidence for fitness consequences on first-year calf survival. Our results also suggest, although not strongly, that first-born calves have lower survival rates than later-born siblings, especially during the first year of life. This is consistent with the available information from a limited number of other well-studied populations of bottlenose dolphins (Henderson, Dawson, Currey, Lusseau, & Schneider, 2014; Mann et al., 2000). Suggested reasons behind a differentiated mortality in first-born calves include poor calf condition (Cheney et al., 2017; Mann & Watson-Capps, 2005; Rowe, Currey, Dawson, & Johnson, 2010), individual differences among mothers in successfully rearing calves (Henderson et al., 2014) and the detrimental effect of organochlorine compounds transferred from the mother, which is higher in firstborn calves (Hall et al., 2006; Wells et al., 2005).

4.4 | Applications

Our results inform assessment of the conservation status of this population, a requirement under the European Habitats Directive (Council Directive 92/43/EEC). As mentioned above, one cause of the increasing survival rates could have been the decline in the use of gillnets in the inner Moray Firth in the 1990s. Knowledge of how survival rates have changed in the past will inform our understanding and assessment of how future pressures, such as the development of offshore wind energy, may impact the east coast population of bot-tlenose dolphins. Population viability analysis (PVA; e.g., Lacy, 2000) is a useful framework for exploring such impacts. The availability of new information on survival presented here and on reproductive rates (Arso Civil et al., 2017) provides an excellent basis for revisiting the PVA that was attempted 20 years ago (Sanders-Reed et al., 1999).

More generally, information on survival will become an increasingly important part of the assessment of conservation status and the impact of human activities on coastal cetaceans. Information of sufficient robustness and detail, and over a sufficient time period, has hitherto been sparse (e.g., Currey et al., 2009; Gormley et al., 2012). But as time series of photoidentification data increase in intensity and length in ongoing studies, results from survival analyses will become more available for input into the assessment process (such as through PVAs). Our study serves as a useful indication of the information that can be made available with sufficient data and a robust analytical framework. Coastal marine mammal populations are globally subject to the impacts of environmental change and increasing anthropogenic disturbance but also to the effects of management measures to reduce human impact. Obtaining robust estimates of age- and sex-specific survival will help understand how populations are responding to these changes.

5 | CONCLUSIONS

In generating robust estimates of survival for a small and discrete population of bottlenose dolphins, we showed very strong support for an increase over time in juvenile/adult survival, fairly strong support for age-specific calf survival and sex-specific juvenile/adult

541

-WILEY-

survival, and weaker support for females driving the trend in juvenile/adult survival and an impact of birth order on calf survival. These results inform our understanding of reasons for an observed change in population size and provide essential information to parameterize future population assessments (PVAs) to explore the impact of human activities. Our study serves as a useful indication of the information that can be made available with sufficient data and a robust analytical framework, to ultimately help understand how populations may respond to the impacts of environmental changes, anthropogenic pressure, as well as management measures implemented to reduce human impacts.

ACKNOWLEDGMENTS

Funding to MAC was provided by the Department for Energy and Climate Change (DECC; now Department for Business, Energy and Industrial Strategy; UK). Annual surveys in the Moray Firth were funded by BES, ASAB, Greenpeace Environmental Trust, SNH, Scottish Government, Whale and Dolphin Conservation, Talisman Energy (UK) Ltd., DECC, Chevron, NERC and the University of Aberdeen. VMJ was funded by a Royal Society University Research Fellowship. St Andrews Bay surveys were funded by the Royal Society, NERC, the CONACYT, DECC, SNH and the University of St Andrews. All fieldwork was carried out under Scottish Natural Heritage Animal Scientific Licenses to PMT, PSH, and VMJ. Biopsy samples were taken under Home Office license number 603135 to VMJ. The authors have no conflict of interest to declare. We thank three anonymous reviewers for their useful comments on the manuscript.

CONFLICT OF INTEREST

None declared.

AUTHOR CONTRIBUTIONS

MAC and PSH conceived the initial ideas and designed methodology; MAC, NQ, BC, VI, and VMJ collected the photo-ID data; VMJ and VI collected biopsies for sexing; VI and JAG conducted the analysis of sex animals; MAC, BC, VI, and NQ processed all photo-ID data; PMT contributed to the interpretation of data for the work and to the analysis of the data; MAC analyzed the data; MAC and PSH led the writing of the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

DATA ACCESSIBILITY

Data available from the Dryad Digital Repository: https://doi. org/10.5061/dryad.8qm8r4m.

ORCID

Mònica Arso Civil ២ https://orcid.org/0000-0001-8239-9526

REFERENCES

- Altwegg, R., Crawford, R. J., Underhill, L. G., & Williams, A. T. J. (2008). Long-term survival of de-oiled Cape gannets Morus capensis after the Castillo de Bellver oil spill of 1983. Biological Conservation, 141, 1924–1929. https://doi.org/10.1016/j.biocon.2008.04.030
- Arso Civil, M. (2014). Population ecology of bottlenose dolphins (*Tursiops truncatus*) off the east coast of Scotland. University of St Andrews, UK. Unpublished PhD thesis.
- Arso Civil, M., Cheney, B., Quick, N. J., Thompson, P. M., & Hammond, P. S. (2017). A new approach to estimate fecundity rate from inter-birth intervals. *Ecosphere*, 8, e01796.
- Bradford, A. L., Wade, P. R., Weller, D. W., Burdin, A. M., Ivashchenko, Y. V., Tsidulko, G. A., ... Brownell, R. L. (2006). Survival estimates of western gray whales *Eschrichtius robustus* incorporating individual heterogeneity and temporary emigration. *Marine Ecology-Progress Series*, 315, 293–307. https://doi.org/10.3354/meps315293
- Branch, T. A., Matsuoka, K., & Miyashita, T. (2004). Evidence for increases in Antarctic blue whales based on bayesian modelling. *Marine Mammal Science*, 20, 726–754. https://doi.org/10.1111/j.1748-7692.2004. tb01190.x
- Brownie, C., Hines, J. E., Nichols, J. D., Pollock, K. H., & Hestbeck, J. B. (1993). Capture-recapture studies for multiple strata including non-Markovian transitions. *Biometrics*, 1(49), 1173–1187. https://doi. org/10.2307/2532259
- Burnham, K. P., & Anderson, D. R. (2002). Model selection and inference: A practical information theoretic approach, 2nd ed. New York, NY: Springer.
- Burnham, K. P., Anderson, D. R., White, G. C., Brownie, C., & Pollock, K. H. (1987). Design and analysis of fish survival experiments based on release-recapture data. *American Fisheries Society Monograph*, *5*, 1–437.
- Butler, J. R. A., McKelvey, S. A., McMyn, I. A. G., Leyshon, B., Reid, R. J., & Thompson, P. M. (2017). Does community surveillance mitigate by-catch risk to coastal cetaceans? Insights from salmon poaching and bottlenose dolphins in Scotland. Oceanography & Fisheries Open Access Journal, 3(1), 555603.
- Caughley, G. (1966). Mortality patterns in mammals. *Ecology*, 47, 906– 918. https://doi.org/10.2307/1935638
- Cheney, B. (2017). Temporal variation in the demographics and dynamics of a bottlenose dolphin population.University of Aberdeen, UK. Unpublished PhD thesis.
- Cheney, B., Graham, I. M., Barton, T. R., Hammond, P. S., & Thompson, P. M. (2018). Site Conditioning Monitoring of bottlenose dolphins within the Moray Firth Special Area of Conservation: 2014–2016. Scottish Natural Heritage Research Report No. 1021.
- Cheney, B., Wells, R. S., Barton, T. R., & Thompson, P. M. (2017). Laser photogrammetry reveals variation in growth and early survival in free-ranging bottlenose dolphins. *Animal Conservation*, https://doi. org/10.1111/acv.12384.
- Cheney, B., Corkrey, R., Durban, J. W., Grellier, K., Hammond, P. S., & Islas-Villanueva, ... Thompson, P. M. (2014). Long-term trends in the use of a protected area by small cetaceans in relation to changes in population status. *Global Ecology and Conservation*, *2*, 118–128. https://doi.org/10.1016/j.gecco.2014.08.010
- Cheney, B., Thompson, P. M., Ingram, S. N., Hammond, P. S., Stevick, P. T., Durban, J. W., ... Wilson, B. (2013). Integrating multiple data sources to assess the distribution and abundance of bottlenose dolphins *Tursiops truncatus* in Scottish waters. *Mammal Review*, 43, 71–88.
- Choquet, R., Lebreton, J.-D., Gimenez, O., Reboulet, A.-M., & Pradel, R. (2009). U-CARE: Utilities for performing goodness of fit tests and manipulating Capture-Recapture data. *Ecography*, 32, 1071–1074.
- Corkrey, R., Brooks, S., Lusseau, D., Parsons, K., Durban, J. W., Hammond, P. S., & Thompson, P. M. (2008). A bayesian capture-recapture population model with simultaneous estimation of heterogeneity. *Journal*

of the American Statistical Association, 103, 948–960. https://doi. org/10.1198/016214507000001256

- Cormack, R. M. (1964). Estimates of survival from the sighting of marked animals. *Biometrika*, 51, 429–438. https://doi.org/10.1093/biomet/51.3-4.429
- Crone, E. E. (2001). Is survivorship a better fitness surrogate than fecundity? *Evolution*, 55, 2611–2614. https://doi. org/10.1111/j.0014-3820.2001.tb00773.x
- Currey, R. J. C., Dawson, S. M., Schneider, K., Lusseau, D., Boisseau, O. J., Haase, P. A., & Slooten, E. (2011). Inferring causal factors for a declining population of bottlenose dolphins via temporal symmetry capture-recapture modeling. *Marine Mammal Science*, 27, 554–566. https://doi.org/10.1111/j.1748-7692.2010.00417.x
- Currey, R. J. C., Dawson, S. M., Slooten, E., Schneider, K., Lusseau, D., Boisseau, O. J., ... Williams, J. A. (2009). Survival rates for a declining population of bottlenose dolphins in Doubtful Sound, New Zealand: An information theoretic approach to assessing the role of human impacts. Aquatic Conservation-Marine and Freshwater Ecosystems, 19, 658–670. https://doi.org/10.1002/aqc.1015
- Doak, D., Kareiva, P., & Klepetka, B. (1994). Modeling population viability for the desert tortoise in the western Mojave Desert. *Ecological Applications*, 4, 446–460. https://doi.org/10.2307/1941949
- Eberhardt, L. (1985). Assessing the dynamics of wild populations. Journal of Wildlife Management, 49, 997-1012. https://doi. org/10.2307/3801386
- Eberhardt, L. L. (2002). A paradigm for population analysis of long-lived vertebrates. *Ecology*, 83, 2841–2854. https://doi.org/10.1890/0012-9658(2002)083[2841:APFPAO]2.0.CO;2
- Ford, J. K. B., Ellis, G. M., Olesiuk, P. F., & Balcomb, K. C. (2010). Linking killer whale survival and prey abundance: Food limitation in the oceans' apex predator? *Biology Letters*, 6, 139–142. https://doi. org/10.1098/rsbl.2009.0468
- Fowler, C. W. (1987). A review of density dependence in populations of large mammals. In H. H. Genoways (Ed.), *Current mammalogy* (pp. 401–441). Boston, MA: Springer.
- Gaillard, J.-M., Festa-Bianchet, M., & Yoccoz, N. G. (1998). Population dynamics of large herbivores: Variable recruitment with constant adult survival. *Trends in Ecology & Evolution*, 13, 58–63. https://doi. org/10.1016/S0169-5347(97)01237-8
- Galimberti, F., Sanvito, S., Boitani, L., & Fabiani, A. (2001). Viability of the southern elephant seal population of the Falkland Islands. Animal Conservation, 4, 81–88. https://doi.org/10.1017/ S1367943001001093
- Gormley, A. M., Slooten, E., Dawson, S. M., Barker, R. J., Rayment, W., du Fresne, S., & Brager, S. (2012). First evidence that marine protected areas can work for marine mammals. *Journal of Applied Ecology*, 49, 474–480. https://doi.org/10.1111/j.1365-2664.2012.02121.x
- Grellier, K., Hammond, P. S., Wilson, B., Sanders-Reed, C. A., & Thompson, P. M. (2003). Use of photoidentification data to quantify mother-calf association patterns in bottlenose dolphins. *Canadian Journal of Zoology-Revue Canadienne De Zoologie*, 81, 1421–1427. https://doi. org/10.1139/z03-132
- Hall, A. J., McConnell, B. J., Rowles, T. K., Aguilar, A., Borrell, A., Schwacke, L., ... Wells, R. S. (2006). Individual-based model framework to assess population consequences of polychlorinated biphenyl exposure in bottlenose dolphins. *Environmental Health Perspectives*, 114, 60–64. https://doi.org/10.1289/ehp.8053
- Hammond, P. S. (2010). Estimating the abundance of marine mammals. In I. L. Boyd, W. D. Bowen, & S.Iverson (Eds.), Marine mammal ecology and conservation: A handbook of techniques (pp. 42–67). Oxford, UK: Oxford University Press.
- Henderson, S. D., Dawson, S. M., Currey, R. J. C., Lusseau, D., & Schneider, K. (2014). Reproduction, birth seasonality, and calf survival of bottlenose dolphins in Doubtful Sound, New Zealand. *Marine Mammal Science*, 30, 1067–1080. https://doi.org/10.1111/mms.12109

- Hestbeck, J. B., Nichols, J. D., & Malecki, R. A. (1991). Estimates of movement and site fidelity using mark-resight data of wintering Canada geese. *Ecology*, 72, 523–533. https://doi.org/10.2307/2937193
- Islas-Villanueva, V. (2009). Genetic characterisation and social structure of the East Scotland population of bottlenose dolphins (*Tursiops truncatus*). University of St Andrews, UK. Unpublished PhD thesis.
- IUCN. (2017) The IUCN Red List of Threatened Species. Version 2017-3. http://www.iucnredlist.org/.
- Jepson, P. D., Deaville, R., Barber, J. L., Aguilar, À., Borrell, A., Murphy, S., ... Law, R. J. (2016). PCB pollution continues to impact populations of orcas and other dolphins in European waters. *Scientific Reports*, *6*, 18573. https://doi.org/10.1038/srep18573
- Kendall, W. L., & Nichols, J. D. (1995). On the use of secondary capturerecapture samples to estimate temporary emigration and breeding proportions. *Journal of Applied Statistics*, 22, 751–762. https://doi. org/10.1080/02664769524595
- Kendall, W. L., Nichols, J. D., & Hines, J. E. (1997). Estimating temporary emigration using capture-recapture data with Pollock's robust design. *Ecology*, 78, 563–578.
- Kraus, S. D., Brown, M. W., Caswell, H., Clark, C. W., Fujiwara, M., Hamilton, P. K., Kenney, R. D., ... Rolland, R. M. (2005). North Atlantic right whales in crisis. *Science*, 309(5734), 561–562.
- Laake, J. L. (2013). RMark: An R Interface for Analysis of Capture-Recapture Data with MARK. AFSC Processed Rep 2013-01, 25p. Alaska Fish. Sci. Cent., NOAA, Natl. Mar. Fish. Serv., 7600 Sand Point Way NE, Seattle WA 98115.
- Lacy, R. C. (2000). Structure of the VORTEX simulation model for population viability analysis. *Ecological Bulletins*, 48, 191–203.
- Lebreton, J.-D., Burnham, K. P., Clobert, J., & Anderson, D. R. (1992). Modeling survival and testing biological hypotheses using marked animals: A unified approach with case studies. *Ecological Monographs*, 62, 67–118. https://doi.org/10.2307/2937171
- Manlik, O., McDonald, J. A., Mann, J., Raudino, H. C., Bejder, L., Krützen, M., ... Sherwin, W. B. (2016). The relative importance of reproduction and survival for the conservation of two dolphin populations. *Ecology* and Evolution, 6, 3496–3512. https://doi.org/10.1002/ece3.2130
- Mann, J., Connor, R. C., Barre, L. M., & Heithaus, M. R. (2000). Female reproductive success in bottlenose dolphins (*Tursiops sp.*): Life history, habitat, provisioning, and group-size effects. *Behavioral Ecology*, 11, 210–219. https://doi.org/10.1093/beheco/11.2.210
- Mann, J., & Watson-Capps, J. J. (2005). Surviving at sea: Ecological and behavioural predictors of calf mortality in Indian Ocean bottlenose dolphins, *Tursiops sp. Animal Behaviour*, 69, 899–909. https://doi. org/10.1016/j.anbehav.2004.04.024
- Marley, S., Cheney, B., & Thompson, P. M. (2013). Using tooth rakes to monitor population and sex differences in aggressive behaviour in bottlenose dolphins (*Tursiops truncatus*). Aquatic Mammals, 39, 107– 115. https://doi.org/10.1578/AM.39.2.2013.107
- Mitchell, M. S., Pacifici, L. B., Grand, J. B., & Powell, R. A. (2009). Contributions of vital rates to growth of a protected population of American black bears. Ursus, 20, 77–84. https://doi. org/10.2192/1537-6176-20.2.77
- Möller, L. M., & Beheregaray, L. B. (2004). Genetic evidence for sex-biased dispersal in resident bottlenose dolphins (*Tursiops* aduncus). Molecular Ecology, 13, 1607–1612. https://doi. org/10.1111/j.1365-294X.2004.02137.x
- New, L. F., Harwood, J., Thomas, L., Donovan, C., Clark, J. S., Hastie, G., ... Lusseau, D. (2013). Modelling the biological significance of behavioural change in coastal bottlenose dolphins in response to disturbance. *Functional Ecology*, 27, 314–322. https://doi. org/10.1111/1365-2435.12052
- Nichols, J. D., Kendall, W. L., Hines, J. E., & Spendelow, J. A. (2004). Estimation of sex-specific survival from capture-recapture data when sex is not always known. *Ecology*, 85, 3192–3201. https://doi. org/10.1890/03-0578

542

- Noren, S. R., & Edwards, E. F. (2007). Physiological and behavioral development in delphinid calves: Implications for calf separation and mortality due to tuna purse-seine nets. *Marine Mammal Science*, 23, 15–29.
- Oli, M. K., & Dobson, F. S. (2003). The relative importance of life-history variables to population growth rate in mammals: Cole's prediction revisited. *The American Naturalist*, 161, 422-440. https://doi. org/10.1086/367591
- Pallin, L. J., Baker, C. S., Steel, D., Kellar, N. M., Robbins, J., Johnston, D. W., ... Friedlaender, A. S. (2018). High pregnancy rates in humpback whales (*Megaptera novaeangliae*) around the Western Antarctic Peninsula, evidence of a rapidly growing population. *Royal Society Open Science*, 5;180017.
- Parsons, K. M., Noble, L. R., Reid, R. J., & Thompson, P. M. (2002). Mitochondrial genetic diversity and population structuring of UK bottlenose dolphins (*Tursiops truncatus*): Is the NE Scotland population demographically and geographically isolated? *Biological Conservation*, 108, 175–182. https://doi.org/10.1016/ S0006-3207(02)00103-9
- Pirotta, E., Laesser, B. E., Hardaker, A., Riddoch, N., Marcoux, M., & Lusseau, D. (2013). Dredging displaces bottlenose dolphins from an urbanised foraging patch. *Marine Pollution Bulletin*, 74, 396–402. https://doi.org/10.1016/j.marpolbul.2013.06.020
- Pirotta, E., Merchant, N. D., Thompson, P. M., Barton, T. R., & Lusseau, D. (2015). Quantifying the effect of boat disturbance on bottlenose dolphin foraging activity. *Biological Conservation*, 181, 82–89. https:// doi.org/10.1016/j.biocon.2014.11.003
- Pledger, S. (2000). Unified maximum likelihood estimates for closed capture-recapture models using mixtures. *Biometrics*, *56*, 434–442. https://doi.org/10.1111/j.0006-341X.2000.00434.x
- Pollock, K. H. (1982). A capture-recapture design robust to unequal probability of capture. *Journal of Wildlife Management*, 46, 752–757. https://doi.org/10.2307/3808568
- Pollock, K. H., Nichols, J. D., Brownie, C., & Hines, J. E. (1990). Statistical inference for capture-recapture experiments. Wildlife Monographs, 107, 1–97.
- Pradel, R., Hines, J. E., Lebreton, J. D., & Nichols, J. D. (1997). Capture-recapture survival models taking account of transients. *Biometrics*, 53, 60–72. https://doi.org/10.2307/2533097
- Pradel, R., Maurin-Bernier, L., Gimenez, O., Genovart, M., Choquet, R., & Oro, D. (2008). Estimation of sex-specific survival with uncertainty in sex assessment. *The Canadian Journal of Statistics*, 36, 29–42. https:// doi.org/10.1002/cjs.5550360105
- Promislow, D. E. (1992). Costs of sexual selection in natural populations of mammals. Proceedings of the Royal Society of London. Series B: Biological Sciences, 247, 203–210.
- Quick, N. J., & Janik, V. M. (2008). Whistle rates of wild bottlenose dolphins (*Tursiops truncatus*): Influences of group size and behavior. *Journal of Comparative Psychology*, 122, 305–311. https://doi. org/10.1037/0735-7036.122.3.305
- R Core Team (2016). R: A language and environment for statistical computing. R Foundation for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing. https://www.R-project. org/.
- Ralls, K., Brownell, R. L., & Ballou, J. (1980). Differential mortality by sex and age in mammals, with specific reference to the sperm whale. Reports of the International Whaling Commission (Special Issue, 2), 233–243.
- Ramp, C., Bérubé, M., Palsboll, P., Hagen, W., & Sears, R. (2010). Sexspecific survival in the humpback whale *Megaptera novaeangliae* in the Gulf of St. Lawrence, Canada. *Marine Ecology Progress Series*, 400, 267–276. https://doi.org/10.3354/meps08426
- Robinson, K. P., O'Brien, J. M., Berrow, S. D., Cheney, B., Costa, M., Eisfeld, S. M., ... Whooley, P. (2012). Discrete or not so discrete : Long distance movements by coastal bottlenose dolphins in UK

and Irish waters. Journal of Cetacean Research and Management, 12, 365–371.

- Ross, P. S. (2002). The role of immunotoxic environmental contaminants in facilitating the emergence of infectious diseases in marine mammals. *Human and Ecological Risk Assessment*, 8, 277–292. https://doi. org/10.1080/20028091056917
- Rowe, L. E., Currey, R. J., Dawson, S. M., & Johnson, D. (2010). Assessment of epidermal condition and calf size of Fiordland bottlenose dolphin *Tursiops truncatus* populations using dorsal fin photographs and photogrammetry. *Endangered Species Research*, 11, 83–89. https://doi. org/10.3354/esr00256
- Sanders-Reed, C. A., Hammond, P. S., Grellier, K., & Thompson, P. M. (1999). Development of a population model for bottlenose dolphins. Scottish Natural Heritage Research, Survey and Monitoring Report No. 156.
- Schwacke, L. H., Voit, E. O., Hansen, L. J., Wells, R. S., Mitchum, G. B., Hohn, A. A., & Fair, P. A. (2002). Probabilistic risk assessment of reproductive effects of polychlorinated biphenyls on bottlenose dolphins (*Tursiops truncatus*) from the Southeast United States Coast. *Environmental Toxicology and Chemistry*, 21, 2752–2764.
- Scott, E. M., Mann, J., Watson-capps, J. J., Sargeant, B. L., & Connor, R. C. (2005). Aggression in bottlenose dolphins: Evidence for sexual coercion, male-male competition, and female tolerance through analysis of tooth-rake marks and behaviour. *Behaviour*, 142, 21–44. https:// doi.org/10.1163/1568539053627712
- Scott, M. D., Wells, R. S., & Irvine, A. B. (1990). A long-term study of bottlenose dolphins on the west coast of Florida. In S. Leatherwood, & R. R. Reeves (Eds.), *The bottlenose dolphin* (pp. 235–244). San Diego, CA: Academic Press.
- Smith, H. C., Pollock, K., Waples, K., Bradley, S., & Bejder, L. (2013). Use of the robust design to estimate seasonal abundance and demographic parameters of a coastal bottlenose dolphin (*Tursiops aduncus*) population. *PLoS ONE*, *8*, e76574.
- Speakman, T. R., Lane, S. M., Schwacke, L., Fair, P. A., & Zolman, E. S. (2010). Mark-recapture estimates of seasonal abundance and survivorship for bottlenose dolphins (*Tursiops truncatus*) near Charleston, South Carolina, USA. Journal of Cetacean Research and Management, 11, 153–162.
- Stolen, M. K., & Barlow, J. (2003). A model life table for bottlenose dolphins (*Tursiops truncatus*) from the Indian river lagoon system. *Marine Mammal Science*, 19, 630–649.
- Thompson, P. M., B. Cheney, S. Ingram, P. T. Stevick, B. Wilson, & P. S. Hammond (Eds.) (2011). Distribution, abundance, and population structure of bottlenose dolphins in Scottish waters. Scottish Government and Scottish Natural Heritage funded report. Scottish Natural Heritage Commissioned Report No. 354.
- Trivers, R. (1972). Parental investment and sexual selection. In B. Campbell (Ed.), *Sexual selection and the descent of man* (pp. 136–179). Chicago, IL: Aldine.
- Ward, E. J., Holmes, E. E., & Balcomb, K. C. (2009). Quantifying the effects of prey abundance on killer whale reproduction. *Journal of Applied Ecology*, 46, 632–640. https://doi. org/10.1111/j.1365-2664.2009.01647.x
- Wells, R. S., & Scott, M. D. (1990). Estimating bottlenose dolphin population parameters from individual identification and capture-release techniques. Reports of the International Whaling Commission (Special Issue, 12), 407–415.
- Wells, R. S., Tornero, V., Borrell, A., Aguilar, A., Rowles, T. K., Rhinehart, H. L., ... Sweeney, J. C. (2005). Integrating life-history and reproductive success data to examine potential relationships with organochlorine compounds for bottlenose dolphins (*Tursiops truncatus*) in Sarasota Bay, Florida. *Science of the Total Environment*, 349, 106–119. https://doi.org/10.1016/j.scitotenv.2005.01.010
- White, G. C., & Burnham, K. P. (1999). Program MARK: Survival estimation from populations of marked animals. *Bird Study*, 46, 120–139. https://doi.org/10.1080/00063659909477239

- White, G. C., Franklin, A. B., & Shenk, T. M. (2002). Estimating parameters of PVA models from data on marked animals. In S. Beissinger, & D. R. McCullough (Eds.), *Population viability analysis* (pp. 169–190). Chicago, IL: University of Chicago Press.
- Williams, B. K., Nichols, J. D., & Conroy, M. J. (2002). Analysis and management of animal populations: Modeling, estimation, and decision making. San Diego, CA: Academic Press.
- Williams, R., Lusseau, D., & Hammond, P. S. (2006). Estimating relative energetic costs of human disturbance to killer whales (Orcinus orca). Biological Conservation, 133, 301–311. https://doi.org/10.1016/j. biocon.2006.06.010
- Wilson, B., Hammond, P. S., & Thompson, P. M. (1999). Estimating size and assessing trends in a coastal bottlenose dolphin population. *Ecological Applications*, 9, 288–300. https://doi.org/10.1890/ 1051-0761(1999)009[0288:ESAATI]2.0.CO;2
- Wilson, B., Reid, R. J., Grellier, K., Thompson, P. M., & Hammond, P. S. (2004). Considering the temporal when managing the spatial: A population range expansion impacts protected areas-based management for bottlenose dolphins. *Animal Conservation*, 7, 331–338. https://doi. org/10.1017/S1367943004001581

Würsig, B., & Jefferson, T. A. (1990). Methods for photoidentification for small cetaceans. Reports of the International Whaling Commission. (Special Issue, 12), 43–51.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

How to cite this article: Arso Civil M, Cheney B, Quick NJ, et al. Variations in age- and sex-specific survival rates help explain population trend in a discrete marine mammal population. *Ecol Evol*. 2019;9:533–544. https://doi.org/10.1002/ece3.4772