

1 Aquafaba from commercially canned chickpeas as potential egg replacer for the
2 development of vegan mayonnaise: recipe optimization and storage stability

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4 Running title: Aquafaba as egg replacer in mayonnaise

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29 **Summary** Aquafaba, the viscous liquid recovered from canned chickpeas, was used
30 as egg replacer for the development of vegan mayonnaise. The textural, microstructural
31 and physicochemical properties of mayonnaise were determined during cold storage to
32 optimize the aquafaba to oil ratio (A/O) of the formulation (15-25/80-70%). Aquafaba
33 was capable to form a stable emulsion with an average value of droplet size distribution
34 below 4 μ m. The physical stability determined by the Turbiscan Stability Index (TSI)
35 was unaffected by the A/O ratio during 21 days of storage at 4 °C. The lowest droplet
36 size distribution was obtained for samples with a low A/O ratio (15/80%). Firmness,
37 adhesive force and adhesiveness decreased ($p<0.05$) with increasing the A/O ratio,
38 whereas consistency remained unaffected. The oxidative stability of the oil phase was
39 similar for all formulations and remained unaffected during storage. Aquafaba can be
40 used effectively to replace egg in mayonnaise formulations containing oil at standard
41 levels.

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43 **Keywords:** emulsion, reformulation, food waste, texture, oxidation

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45 **Introduction**

46 Mayonnaise is a semi-solid, acidic condiment which is widely appreciated for adding
47 texture and flavor to other foods such as salads and sandwiches. In its traditional recipe,
48 it contains 70-80% vegetable oil, egg yolk, salt, sugar, vinegar and spices, typically
49 mustard (Depre and Savage, 2001). Mayonnaise is a colloidal system from a structural
50 perspective, formed by emulsified oil droplets of spherical shape in a homogeneous
51 aqueous phase. The oil droplet stability is mediated primarily by the emulsifying action
52 of granular micro-particles formed from the phosphoprotein and low-density
53 lipoprotein constituents of egg yolk (Laca *et al.*, 2010).

54 Numerous attempts have been made in recent years to reformulate mayonnaise in order
55 to meet consumer demands for a low-fat product with improved lipid profile. The
56 primary objective of re-designing the recipe of mayonnaise focused on reducing the
57 amount or altering the type of fat which, if overly consumed may be harmful for the

58 onset and development of chronic diseases (Worrasinchai *et al.*, 2006; Liu *et al.*, 2007;
59 Di Mattia *et al.*, 2015). Other potentially harmful ingredients for human health have
60 also been targeted for reduction in the recipe, such as yolk cholesterol and salt. Selective
61 egg yolk proteins are also considered responsible for triggering adverse immunological
62 reactions in infants, young children and to a lesser extent the adult population (Caubet
63 and Wang, 2011).

64 During the last years there has been considerable effort from the food industry to
65 remove egg yolk from the formula of mayonnaise. This stems from health-related as
66 well as sustainability issues associated with the consumption of animal products and is
67 manifested as a market-driven preference towards the development of healthier, “free-
68 from” and “natural” products. Furthermore, an egg-free mayonnaise may also be more
69 cost effective from a manufacturer’s perspective since pasteurization will no longer be
70 a requirement during production. One of the main challenges encountered in the process
71 of developing an egg-free mayonnaise is to identify suitable ingredients to replace egg
72 yolk from the traditional recipe, without impairing stability, taste and color. Emulsifiers
73 of animal or plant origin have been employed for this purpose (Riscardo *et al.*, 2003;
74 Herald *et al.*, 2009; Nikzade *et al.*, 2012). White lupin protein, soy milk, wheat germ
75 protein isolate, chia mucilage, Durian seed gum and modified potato starch have been
76 tested so far for their ability to replace egg yolk and the main challenge was to generate
77 a stable emulsion structure of fine oil droplets capable to prevent coalescence and
78 flocculation for prolonged periods of storage (Cornelia *et al.*, 2015; Fernandes &
79 Mellado, 2018; Ghazaei *et al.*, 2015; Rahbari *et al.*, 2015; Rahmati *et al.*, 2014;
80 Raymundo *et al.*, 2002).

81 “Aquafaba” is the term used to describe the viscous liquid formed during cooking of
82 legume seeds (typically chickpeas) or the one encountered in canned products of the
83 same origin. The exact composition of aquafaba depends on the legume and is a mixture
84 of carbohydrates, proteins and water (Shim *et al.*, 2018). This liquid, which is usually
85 discarded as food waste, is also a source of phenolic compounds and saponins (Damian
86 *et al.*, 2018). Research has indicated aquafaba as a valuable ingredient with desirable

87 functional properties (i.e. foaming, emulsifying and gelling) which can be used in
88 various formulations to replace eggs and milk in vegan products (Serventi *et al.*, 2018;
89 Shim *et al.*, 2018; Stantiall *et al.*, 2018; Mustafa *et al.*, 2018).

90 To the best of our knowledge, there are no studies to investigate the potential of
91 aquafaba as an egg yolk replacer for the development of vegan mayonnaise. The aim
92 of the present study is to develop and optimize the recipe of mayonnaise using aquafaba
93 from chickpeas. Formulation effects on texture and physicochemical properties of
94 mayonnaise are determined during cold storage to assess the applicability of this
95 secondary food ingredient as an emulsifier and stabilizer of high-fat colloidal systems.

96

97 **Materials and methods**

98 **Materials**

99 Canned chickpeas (Lot FDI CC26 6 LJ085), rapeseed oil, table salt, sugar and white
100 wine vinegar were purchased from the local supermarket (Tesco, UK). The average
101 chickpea to aquafaba weight ratio was 1.68. Folin-Denis' reagent and Nile red were
102 supplied by Sigma Aldrich (St Louis, MO, USA). All standards and reagents used were
103 of analytical grade.

104

105 **Preparation and storage of mayonnaise**

106 The formulation contained the following ingredients on % weight basis: 80% oil, 15%,
107 aquafaba, 4% vinegar, 0.5% sugar, 0.5% salt. For reduced-fat mayonnaise (70%-75%),
108 oil was replaced by an equal amount of aquafaba (20%-25%). A coarse emulsion was
109 formed by adding the oil gradually to the aqueous mixture (aquafaba, vinegar, sugar
110 and salt) and mixing for 10 min with a Russell Hobbs hand blender (Argos, UK).
111 Mayonnaise was then homogenized with a T25 digital Ultra-turrax® homogenizer at
112 13500 rpm for 2 min (IKA® England Ltd, Oxford, UK). 500 gr of mayonnaise were
113 prepared for each batch and for each formulation three batches were prepared.
114 Mayonnaise was aliquoted and stored at 4 °C until further analysis at weekly intervals.

115

116 Proximate composition and determination of total phenols, tocopherols and carotenoids
117 in aquafaba
118 Energy, moisture, ash, fat, carbohydrates, total sugars, and dietary fiber in the samples
119 were determined according to the standard AOAC (1990) official methods. Protein
120 content was determined by combustion according to the Dumas principle.
121 Carbohydrates were determined by subtracting the sum of moisture, protein, fat, and
122 ash percentages from 100%. Total phenols were determined using the Folin-Ciocalteu
123 (F-C) colorimetric method according to Raikos *et al.* (2014) and results are expressed
124 as mg GAE/g of dried aquafaba. A reverse-phase HPLC method was employed to
125 quantify carotenoids and tocopherols and samples were analyzed in duplicates (Hess *et*
126 *al.*, 1991).

127

128 pH and color determination

129 pH was recorded using a portable food and dairy pH meter (Hanna Instruments Ltd.,
130 Leighton Buzzard, UK) and color was determined by a Konica Minolta CR1 10
131 colorimeter (Konica Minolta Solutions. Ltd., Basildon, UK). ΔE^* (total colour change)
132 of mayonnaise samples during cold storage was calculated from the following equation:

$$133 \Delta E^* = \sqrt{(\Delta L^*)^2 + (\Delta a^*)^2 + (\Delta b^*)^2}$$

134

135 Lipid extraction from mayonnaise

136 Twenty-gram portions of mayonnaise were poured into 50 ml polypropylene centrifuge
137 tubes and were frozen at -20 °C for 24 h according to the procedure of Lagunes-Galvez
138 *et al.* (2002). After storage samples were thawed at room temperature until the emulsion
139 was broken and oil phase was collected with centrifugation at 2,400 x g for 5 min with
140 an Eppendorf 8810R apparatus (Eppendorf UK Ltd, Stevenage, UK).

141

142 Oxidation stability testing

143 The oxidative stability of mayonnaise was determined by a 743 Rancimat device
144 (Metrohm Ltd., Herisau, Switzerland) according as described by Raikos *et al.* (2016)

145 with slight modifications. Three grams of extracted oil was poured into the reaction
146 tubes, samples were exposed to an air flow of 20L/h at 120 °C and the induction time
147 (IP) was calculated by the 743 Rancimat software 1.1.

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149 Emulsion stability

150 The physical stability of mayonnaise samples was monitored using a Turbiscan
151 MA2000 apparatus (Formulation, RamonvilleSt. Agne, France) as described by
152 Raikos *et al.* (2017) with slight modifications. The sample in the cell was scanned every
153 5 min for 30 min at 40 °C and the changes in the intensity of the backscattered light
154 (Δ BS) in unit time was taken as a measure of the stability of the emulsions. The particle
155 size (mean spherical equivalent diameter) was computed from the Δ BS values based on
156 the refractive index of particles and continuous phase, and the volume fraction (ϕ) of
157 particles in the sample (Mie theory). The volume fraction was adjusted at 80%, 75% or
158 70% depending on the oil added for each mayonnaise formulation and the refractive
159 indices for particle size calculation were 1.47 for the dispersed phase (rapeseed oil) and
160 1.33 for the continuous phase. The Turbiscan stability index (TSI) was calculated from
161 Δ BS that indicate the particles aggregation and migration by Turbisoft 2.0.

162

163 Texture analysis

164 Texture measurements were performed using a CT3 Texture Analyzer (Brookfield
165 Engineering Laboratories Inc., Middleboro, MA) **attached with a 10 kg load cell and**
166 **equipped with** a special mayonnaise mesh probe (TA-MP, length:3.8cm, width:3.4cm,
167 mesh size:0.4mm) **A two cycle Texture Profile Analysis compression test (TPA) was**
168 **performed with the following TPA settings: pre-test speed: 2 mm/s; test speed: 1 mm/s;**
169 **return speed: 1 mm/s; trigger load: 10 g; target mode distance: 20 mm; data acquisition**
170 **rate: 10 points/s.** Mayonnaise samples (200 g) were **carefully scooped** into 250-mL
171 Corning® polypropylene cone beakers (Sigma-Aldrich, St. Louis, MO) and the mesh
172 probe compressed the sample **at a constant crosshead speed of 1 mm/s twice** to a depth
173 of 20mm **of the initial height at room temperature.** Data were recorded using Texture

174 Proc CT V1.3 Build 15 software (Brookfield Engineering Laboratories Inc.) and the
175 parameters determined were hardness (firmness), adhesive force - adhesiveness
176 (stickiness) and cohesiveness (consistency). Hardness was calculated from the load
177 detected at highest peak during compression, adhesive force from the peak negative
178 value, adhesiveness from the area under the negative peak and cohesiveness from the
179 ratio of the areas under the compression stroke of the second and first cycles.

180

181 Microstructure analysis

182 Mayonnaise microstructure was analyzed with a confocal laser scanning microscope
183 (CLSM) (Carl Zeiss Ltd, Cambridge, UK) according to the method of Raikos et al.
184 (2019). Nile red dye was used to stain the fat globules and observations were performed
185 at 543 nm using a 63x oil immersion objective. Images were captured at a resolution of
186 1024 x 1024 pixels.

187

188 Statistical analysis

189 The data are reported as means±standard error (SE) for duplicate measurements **from 3**
190 **batches (n=6) unless otherwise stated**. Analysis of variance (SPSS for Windows 22,
191 SPSS Inc., Chicago, IL) were conducted to identify differences among the means by
192 the *Tukey's* post hoc test. Statistical significance was set at $p < 0.05$.

193

194 **Results and discussion**

195 Proximate and chemical composition of aquafaba

196 The proximal composition of aquafaba from commercial canned chickpeas is presented
197 in Table 1. The values obtained for protein, moisture, ash and simple and complex
198 carbohydrates agree with previously published data (Mustafa et al., 2018). Fat was not
199 detected in measurable levels which agrees with published literature (Mustafa et al.,
200 2018; Stantiall et al., 2018). The total phenolic content was lower, and the sodium
201 content was higher compared to the data from Damian et al. (2018). The observed
202 differences can be due to pulse compositional differences or to degradation effects from

203 different processing methods (Shim *et al.*, 2018). The vitamin C and tocopherol content
204 of raw and processed chickpea seeds from previously published research indicates that
205 this legume is a fair source of tocopherols and the main tocopherol detected was the γ -
206 isomer (7.7 mg/100 g of dry matter) (Fernandez-Orozco *et al.*, 2009). To the best of our
207 knowledge, the vitamin and carotenoid content of aquafaba remains largely unknown.
208 Our data indicated that although tocopherols (mainly γ -tocopherol) were detected in
209 aquafaba, these were at very low levels. Non-surprisingly, only traces of carotenoids
210 were detected which is due to the absence of fat from the proximal composition of the
211 viscous liquid. It is therefore assumed that their nutritional and functional contribution
212 to the properties of mayonnaise is negligible.

213

214 Microstructure and textural properties of mayonnaise

215 The main objective of this study was to determine the efficiency of aquafaba to form a
216 stable colloidal dispersion of oil droplets in mayonnaise structure. As there is no
217 existing literature to provide preliminary data, different formulations with respect to
218 aquafaba to oil ratios were tested. Mayonnaise is a semisolid food which exhibits
219 pseudoplastic and time-dependent behavior (Olsson *et al.*, 2018). The microstructure
220 and texture of mayonnaise relate to its viscoelastic properties, which in turn determine
221 product quality and acceptability. The microstructure of mayonnaise is determined by
222 different factors including the type and concentration of emulsifiers used to form the
223 emulsion, the viscosity of the aqueous phase, the oil content and the droplet size (Laca
224 *et al.*, 2010). The microstructure of the freshly formed mayonnaise with different
225 aquafaba to oil ratios were analyzed by CLSM (Fig. 1). Microstructural analysis
226 revealed that all mayonnaises consisted of finely dispersed, spherical oil droplets in the
227 aqueous medium. Due to the high oil content (>60%), the droplets are densely packed
228 and show a certain degree of polydispersity (different size). The close packing of the
229 droplets, also denoted as droplet density, favors inter-droplet interactions and is at least
230 partially responsible for the stability of the mayonnaise structure (Depree & Savage,
231 2001). The appearance and mean spherical equivalent diameter (3.4-4.1 μm) of the

232 droplets is comparable to the microstructure of traditional mayonnaise made from egg
233 yolk (Patil & Benjakul, 2019).

234 The textural parameters of the freshly prepared mayonnaise samples are presented in
235 Table 2. Hardness, adhesive force and adhesiveness are significantly affected by the
236 aquafaba to oil ratio. Hardness (or firmness) indicates the force required to compress
237 food between molar teeth, adhesive force represents the force required to overcome the
238 attractive forces between mayonnaise and the surface of other materials and
239 adhesiveness is the energy required to separate mayonnaise from the spoon or knife
240 (Chandra & Shamasundar, 2015; Raikos *et al.*, 2016). Cohesiveness (or consistency),
241 which indicates the strength of internal bonds within mayonnaise and the degree to
242 which it can be deformed before rupture, was not affected by the aquafaba to oil ratio.
243 Increasing the oil to aquafaba ratio resulted in significant increases in hardness,
244 adhesiveness and adhesive force. The observed increase in textural parameters is
245 reflective of the increase in viscosity due to higher oil content. The larger the amount
246 of oil added, the greater the number of droplets formed and thus emulsion viscosity
247 increases (Fernandes & Mellado, 2017). According to Liu *et al.* (2007), viscosity can
248 at least partially determine the textural profile of mayonnaise. Data from previous
249 research also indicates that firmness and adhesiveness of mayonnaise are affected by
250 changes in viscosity when the protein, hydrocolloid or oil content of the formulation is
251 modified (Nikzade *et al.*, 2012; Raymundo *et al.*, 2002)

252

253 Physicochemical stability of mayonnaise during cold storage

254 Mayonnaise in its traditional recipe is an acidic emulsion with a long shelf-life (up to 6
255 months) under refrigerated temperatures (Herald *et al.*, 2009). The long-term stability
256 of mayonnaise is attributed to the high fat content which results in a highly dense
257 packing of droplets with limited space to move and to the presence of highly efficient
258 emulsifiers (i.e. egg lecithin) capable to reduce the interfacial tension between the
259 dispersed and continuous phases and form a strong viscoelastic film around the oil
260 droplets which is electrically charged (McClements, 2009; Mun *et al.*, 2009). Although

261 there have been considerable efforts to (at least) partially replace egg yolk from the
262 traditional formulation of mayonnaise, long-term stability of the reformulated product
263 has been compromised in most attempts. The long-term stability of mayonnaise is
264 associated, among other factors, with the mean particle size and particle-size
265 distribution of the oil droplets (Yildirim *et al.*, 2016). All mayonnaise samples from
266 three independent batches were acidic (pH ranging from 3.45 to 3.67) and pH values of
267 three aquafaba to oil formulations did not differ significantly ($p>0.05$). The change of
268 particle size (mean spherical equivalent diameter) and Turbiscan Stability Index (TSI)
269 were used to evaluate the emulsion stability of mayonnaise during refrigerated storage
270 (Fig. 2). TSI is a stability specific parameter that can be used for the determination of
271 emulsion stability and is obtained as the sum of all destabilization phenomena taking
272 place during the monitoring process. High TSI values indicate decreased stability of the
273 system (Sun *et al.*, 2015). As shown in Fig. 2, the mean spherical equivalent diameter
274 remained unaffected during 21 days of refrigerated storage for all 3 formulations. At
275 the end of the storage period, the formulation with 15% aquafaba had the smallest
276 particle size (3.22 μm), which suggests that the amount of emulsifier present in the
277 lowest aquafaba to oil ratio (15:80) is adequate to form a stable emulsion. The
278 emulsifying ability of aquafaba is attributed to the protein and carbohydrate content
279 (Table 1). Both macromolecular components should contribute to the formation and
280 stabilisation of the emulsion structure. Previous research has shown that proteins and
281 polysaccharides can interact to form a thick interfacial film surrounding the oil droplet,
282 which results in a stable emulsion (Ghoush *et al.*, 2008). Although the chemical
283 composition of aquafaba is not standardised or known in detail so far, there is evidence
284 to suggest that it is a good source of storage proteins (albumins and globulins) and
285 complex, digestible and non-digestible carbohydrates (Shim *et al.*, 2018). Aquafaba is
286 known to contain saponins, amphiphilic glycosides which can act as surfactants and
287 thus may also contribute to the high emulsifying ability of the viscous liquid from
288 canned chickpeas (Damian *et al.*, 2018).

289 Mayonnaise is a high-fat food (70%-80% vegetable oil) and therefore is susceptible to

290 oxidative deterioration through auto-oxidation of the unsaturated and polyunsaturated
291 fats in the oil, which depending on the extent is likely to have a negative impact on
292 flavor, aroma, colour and nutritional value of food (Depree & Savage, 2001). Several
293 strategies can be effective against lipid oxidation of mayonnaise such as the addition of
294 antioxidants or the use of a lipid source that is naturally rich in compounds with
295 powerful antioxidant activity (Di Mattia *et al.*, 2015; Li *et al.*, 2015). The oxidative
296 stability of the extracted lipid phase of mayonnaise samples during storage at 4 °C was
297 determined by Rancimat analysis and is presented in Figure 3. The induction time,
298 which is an indication of the ability of oil to resist oxidation under accelerated oxidation
299 conditions, of all three formulations was similar and was not affected significantly after
300 21 days of storage. The formulation effect (oil to aquafaba ratio) was non-significant
301 with respect to the oxidative stability of mayonnaise. The induction time of all samples
302 ranged between 3.33 h to 3.55 h, which is comparable to previously published data for
303 rapeseed oil (Maszewska *et al.* 2018). Data from colour properties of the mayonnaise
304 samples also suggests that samples remained stable during storage as indicated by the
305 total differences in colour (ΔE^*) (Table 3). There was a non-significant ($p < 0.05$)
306 incremental effect for all formulations at the end of the study period which was evident
307 for all colour coordinates (L^* , a^* , b^*). The formulation effect (oil to aquafaba ratio)
308 was observed only for b^* (yellowness), which non-surprisingly increased significantly
309 for higher oil to aquafaba ratios.

310

311 **Conclusions**

312 This study aimed to evaluate the potential of aquafaba as an egg replacer for the
313 development of a vegan mayonnaise formulation. Microstructural and light scattering
314 data indicated that aquafaba can be effectively used to form a fine emulsion, which
315 remained stable during cold storage for 21 days. The A/O ratio used for product
316 development had a minor impact on colour properties and no significant effect on the
317 physicochemical stability of the mayonnaise during cold storage. The A/O ratio
318 significantly affected the textural properties and this effect was dependent on the oil

319 contribution to the formulation. Increasing the aquafaba at levels above 15% had no
320 beneficial effect on the long-term stability or the antioxidant properties of the
321 formulation. Sensory evaluation of the mayonnaise made from aquafaba should be
322 carried out to determine consumer acceptability. This study suggests that aquafaba is
323 currently an underutilized secondary (waste) product which can have several
324 applications in the food industry thanks to its nutritional, functional and health-related
325 properties.

326

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329 Scottish Governments Rural and Environment Science and Analytical Services
330 Division (RESAS). Microscopy was performed in the Microscopy and Histology Core
331 Facility at the University of Aberdeen.

332

333 **Data Availability**

334 Research data are not shared

335

336 **Ethical Guidelines**

337 Ethics approval was not required for this research

338

339 **Conflicts of interest**

340 The authors declare no conflicts of interest.

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343

344 **Figure captions**

345 **Graphical abstract** The development of vegan mayonnaise from secondary products
346 of food processing and its benefits

347 **Figure 1** Confocal Laser Scanning Microscopic (CLSM) images of mayonnaise
348 prepared with A: 15%, B: 20% and C: 25% aquafaba. Lipid droplets are stained with
349 Nile red and scale bar equals to 20 μm .

350 **Figure 2** Effect of aquafaba to oil ratio on A: the mean spherical equivalent diameter
351 (d) and B: the physical stability of mayonnaise (TSI) during 21 days of storage period
352 at 4 °C. Results are presented as means \pm SE. Different letters denote significant
353 differences ($p<0.05$).

354 **Figure 3** Effect of aquafaba to oil ratio on the oxidative stability (IP) of mayonnaise
355 during 21 days of storage period at 4 °C. Results are presented as means \pm SE.

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373 **References**

- 374 AOAC. 1990. Official Methods of Analysis. 15th ed. Association of Official Analytical
375 Chemists, Washington, DC.
- 376 Caubet, J.C. & Wang, J. (2011). Current understanding of egg allergy. *Pediatric Clinics*
377 *of North America*, **58(2)**, 427-443.
- 378 Cornelia, M., Siratantri, T. & Prawita, R. (2015). The utilization of extract Durian
379 (Durio zibethinus L.) seed gum as an emulsifier in vegan mayonnaise. *Procedia Food*
380 *Science*, **3**, 1-18.
- 381 Damian, J.J., Huo, S. & Serventi, L. (2018). Phytochemical content and emulsifying
382 ability of pulses cooking water. *European Food Research and Technology*, **244**, 1647-
383 1655.
- 384 Depree, J.A. & Savage G.P. (2001). Physical and flavor stability of mayonnaise. *Trends*
385 *in Food Science and Technology*, **12**, 157–163.
- 386 Di Mattia, C., Balestra, F., Sachetti, G., Neri, L., Mastrocola, D. & Pitia P. (2015).
387 Physical and structural properties of extra-virgin olive oil based mayonnaise. *LWT-*
388 *Food Science and Technology*, **62**, 764-770.
- 389 Fernandes, SS. & Mellado, M.D.L.M.S. (2018). Development of mayonnaise with
390 substitution of oil or egg yolk by the addition of chia (*Salvia Hispânica* L.) mucilage.
391 *Journal of Food Science*, **83(1)**, 74-83.
- 392 Ghazaei, S., Mizani, M., Piravi-Vanak & Alimi, M. (2015). Particle size and cholesterol
393 content of a mayonnaise formulated by OSA-modified potato starch. *Food Science and*
394 *Technology*, **35(1)**, 150-156.
- 395 Ghoush, M.A., Samhour, M, Al-Holy, M. & Herald, T. (2008). Formulation and fuzzy
396 modeling of emulsion stability and viscosity of a gum–protein emulsifier in a model
397 mayonnaise system. *Journal of Food Engineering*, **84**, 348–57.
- 398 Fernandez-Orozco, R., Frias, J., Zielinski, H., Muñoz, R., Piskula, M.K., Kozłowska,
399 H. & Vidal-Valverde, C. (2009). Evaluation of bioprocesses to improve the antioxidant
400 properties of peas. *LWT-Food Science and Technology*, **42**, 885-892.
- 401 Herald, T.J., Abugoush, M. & Aramouni, F. (2009). Physical and sensory properties of

402 egg yolk and egg yolk substitutes in a model mayonnaise system. *Journal of Texture*
403 *Studies*, **40**, 692-709.

404 Hess, D., Keller, H.E., Oberlin, B., Bonfanti, R. & Schüep, W. (1991). Simultaneous
405 determination of retinol, tocopherols, carotenes and lycopene in plasma by means of
406 high-performance liquid chromatography on reverse phase. *International Journal of*
407 *Vitamin and Nutrition Research*, **61**, 232–238.

408 influence of emulsifier concentration and storage time. *International Journal of Food*
409 *Science and Technology*, **52**, 348-358.

410 Laca, A., Sàenz, M. C., Paredes, B. & Diaz, M. (2010). Rheological properties, stability
411 and sensory evaluation of low cholesterol mayonnaises prepared using egg yolk
412 granules as emulsifying agent. *Journal of Food Engineering*, **97**, 243-252.

413 Lagunes-Galvez, L., Cuvelier, M.E., Ordonnaud, C., Berset, C. (2002). Oxidative
414 stability of some mayonnaise formulations during storage and daylight irradiation,
415 *Journal of Food Lipids*, **9**, 211–224.

416 Li, C.-Y., Kim, H.-W., Li, H., Lee, D.-C. & Rhee, H.-I. (2014). Antioxidative effect of
417 purple corn extracts during storage of mayonnaise. *Food Chemistry*, **152**, 592-596.

418 Liu, H., Xu, X. M. & Guo, S. D. (2007). Rheological, texture and sensory properties of
419 low-fat mayonnaise with different fat mimetics. *LWT- Food Science and Technology*,
420 **40**, 946-954.

421 Maszewska, M., Florowska, A., Dłużewska, E., Wroniak, M., Marciniak-Lukasiak, K.,
422 & Żbikowska, A. (2018). Oxidative Stability of Selected Edible Oils. *Molecules*, **23(7)**,
423 1746.

424 Di Mattia, C., Balestra, F., Sacchetti, G., Neri, L., Mastrocola, D. & Pittia, P. (2015).
425 Physical and structural properties of extra-virgin olive oil based mayonnaise. *LWT-*
426 *Food Science and Technology*, **62**, 764-770.

427 McClements DJ (2009) Biopolymers in food emulsions. In: Kasapis S, Norton IT,
428 Ubbink JB (ed) *Modern Biopolymer Science*, Springer. pp 129–166.

429 Mun, S., Kim, Y.L., Kang, C.G. & Park, K.H. (2009). Development of reduced-fat
430 mayonnaise using 4 α GTASE modified rice starch and xanthan gum. *International*

431 *Journal of Biological Macromolecules*, **44**, 400–407.

432 Mustafa, R., He, Y., Shim, Y.Y. & Reaney M.J.T. (2018). Aquafaba, wastewater from
433 chickpea canning, functions as an egg replacer in sponge cake. *International Journal*
434 *of Food Science and Technology*, **53**, 2247-2255.

435 Nikzade, V., Tehrani, M.M., Saadatmand-Tarzjan, M. (2012). Optimization of low-
436 cholesterol-low-fat mayonnaise formulation: Effect of using soy milk and some
437 stabilizer by a mixture design approach. *Food Hydrocolloids*, **28**, 344-352.

438 Olsson, V., Håkansson, A., Purhagen, J. & Wendin, K. (2018). The effect of emulsion
439 intensity on selected sensory and instrumental texture properties of full-fat mayonnaise.
440 *Foods*, **7**, 1-9.

441 Patil, U. & Benjakul, S. (2019). Physical and textural properties of mayonnaise
442 prepared using virgin coconut oil/fish oil blend. *Food Biophysics*, **14(3)**, 260-268.

443 Rahbari, M., Aalami, M., Kashaninejad, M., Maghsoudlou, Y. & Aghdaei, S.S.A.
444 (2015). A mixture design approach to optimizing low cholesterol mayonnaise
445 formulation prepared with wheat germ protein isolate. *Journal of Food Science and*
446 *Technology*, **52(6)**, 3383-3393.

447 Rahmati, K., Tehrani, M.M. & Daneshvar, K. (2014). Soy milk as an emulsifier in
448 mayonnaise: physico-chemical, stability and sensory evaluation. *Journal of Food*
449 *Science and Technology*, **51(11)**, 3341-3347.

450 Raikos, V., Duthie, G. & Ranawana, V. (2017). Comparing the efficiency of different
451 food-grade emulsifiers to form and stabilise orange oil-in-water beverage emulsions:
452 Raikos, V., McDonagh, A., Ranawana, V. & Duthie G. (2016). Processed beetroot (Beta
453 vulgaris L.) as a natural antioxidant in mayonnaise: Effects on physical stability, texture
454 and sensory attributes. *Food Science and Human Wellness*, **5**, 191-198.

455 Raikos, V., Neacsu M., Morrice, P. & Duthie G. (2014). Physicochemical stability of
456 egg protein-stabilised oil-in-water emulsions supplemented with vegetable powders.
457 *International Journal of Food Science and Technology*, **49**, 2433-2440.

458 Raymundo, A., Franco, J.M. Empis, J. & Sousa, I. (2002). Optimization of the
459 composition of low-fat oil-in-water emulsions stabilized by white lupin protein.

460 *Journal of the American Oil Chemists' Society*, **79(8)**, 783-790.

461 Riscardo, M. A., Franco, J. M. & Gallegos, C. (2003). Influence of composition of
462 emulsifier blends on the rheological properties of salad dressing-type emulsion. *Food*
463 *Science and Technology International*, **9**, 53-63.

464 Serventi, L., Wang, S., Zhu, J., Liu, S. & Fei, F. (2018). Cooking water of yellow
465 soybeans as emulsifier in gluten-free crackers. *European Food Research and*
466 *Technology*, **244**, 2141-2148.

467 Shim, Y.Y., Mustafa, R., Shen, J., Ratanapariyanuch, K. & Reaney, M.J.T. (2018).
468 Composition and properties of aquafaba: water recovered from commercially canned
469 chickpeas. *Journal of Visualized Experiments*, **132**(e56305), 1–14.

470 Stantiall, S.E., Dale, K.J., Calizo, F.S. & Serventi, L. (2018). Application of pulses
471 cooking water as functional ingredients: the foaming and gelling abilities. *European*
472 *Food Research and Technology*, **244**, 97–104.

473 Sun, C., Wu, T., Liu, R., Liang, B., Tian, Z., Zhang, E. & Zhang, M. (2015). Effects of
474 superfine grinding and microparticulation on the surface hydrophobicity of whey
475 protein concentrate and its relation to emulsions stability. *Food Hydrocolloids*, **51**, 512-
476 518.

477 Worrasinchai, S., Suphantharika, M., Pinjai, S. & Jamnong, P. (2006). β -Glucan
478 prepared from spent brewers yeast as a fat replacer in mayonnaise. *Food Hydrocolloids*,
479 **20**, 60-78.

480 Yildirim, M., Sumnu, G. & Sahin, S. (2016). Rheology, particle-size distribution, and
481 stability of low-fat mayonnaise produced via double emulsions. *Food Science and*
482 *Biotechnology*, **25**, 1613-1618.

483