

1 **An agent-based model that simulates the spatio-temporal dynamics of sources and**
2 **transfer mechanisms contributing faecal indicator organisms to streams. Part 1:**
3 **Background and model description**

4

5 *Aaron J. Neill^{a,b}, Doerthe Tetzlaff^{e,d,a}, Norval J.C. Strachan^e, Rupert L. Hough^b, Lisa M. Avery^b, Sylvain*
6 *Kuppel^{f,g,a}, Marco P. Maneta^h, Chris Soulsby^{a,c}*

7

8 ^a Northern Rivers Institute, University of Aberdeen, Aberdeen, AB24 3UF, Scotland, United Kingdom.

9

10 ^b The James Hutton Institute, Craigiebuckler, Aberdeen, AB15 8QH, Scotland, United Kingdom.

11

12 ^c IGB Leibniz Institute of Freshwater Ecology and Inland Fisheries, 12587 Berlin, Germany.

13

14 ^d Department of Geography, Humboldt University Berlin, 10099 Berlin, Germany.

15

16 ^e School of Biological Sciences, University of Aberdeen, Cruickshank Building, St Machar Drive, Aberdeen,
17 AB24 3UU, Scotland, United Kingdom.

18

19 ^f Institut de Physique du Globe de Paris, CNRS UMR 7154 - University of Paris, 75231 Paris, France.

20

21 ^g INRAE, RiverLy, 69625 Villeurbanne, France.

22

23 ^h Geosciences Department, University of Montana, Missoula, MT 59812-1296, USA.

24

25 Corresponding Author

26 Aaron J. Neill

27 aaron.neill@abdn.ac.uk

28 Northern Rivers Institute, University of Aberdeen, Aberdeen, AB24 3UF, Scotland, United Kingdom.

29 **Highlights**

- 30 • Present a new agent-based model for simulating faecal indicator organisms (FIOs).
- 31 • Simulates fate and transport of agents representing a sufficient sample of FIOs.
- 32 • Models loading, die-off, detachment, surface routing, seepage and channel routing.
- 33 • Hydrological environment is generated by a robustly-constrained hydrological model.
- 34 • Present EcH₂O-iso as an appropriate hydrological environment generator.

35

36 **Abstract**

37 A new Model for the Agent-based simulation of Faecal Indicator Organisms (MAFIO) is developed
38 that attempts to overcome limitations in existing faecal indicator organism (FIO) models arising from
39 coarse spatial discretisations and poorly-constrained hydrological processes. MAFIO is a spatially-
40 distributed, process-based model presently designed to simulate the fate and transport of agents
41 representing FIOs shed by livestock at the sub-field scale in small (<10 km²) agricultural catchments.
42 Specifically, FIO loading, die-off, detachment, surface routing, seepage and channel routing are
43 modelled on a regular spatial grid. Central to MAFIO is that hydrological transfer mechanisms are
44 simulated based on a hydrological environment generated by an external model for which it is possible
45 to robustly determine the accuracy of simulated catchment hydrological functioning. The spatially-
46 distributed, tracer-aided ecohydrological model EcH₂O-iso is highlighted as a possible hydrological
47 environment generator. The present paper provides a rationale for and description of MAFIO, whilst a
48 companion paper applies the model in a small agricultural catchment in Scotland to provide a proof-of-
49 concept.

50

51

52

53 *Key words:*

54 Diffuse pollution; *E. coli*; EcH₂O-iso; Microbial water quality; Tracer-aided modelling; Water quality
55 modelling

56 **1 Introduction**

57 Transfer of faecal pathogens (e.g. *E. coli* O157) to water bodies represents a significant risk to public
58 health, with the ingestion of contaminated water through drinking or recreational use having the
59 potential to cause severe gastrointestinal illness in humans (Fewtrell and Kay, 2015; Oliver et al.,
60 2005a). Faecal indicator organisms (FIOs), such as generic *E. coli*, are commonly used to monitor
61 microbial water quality and indicate the potential presence of pathogens which are more difficult to
62 quantify directly (Geldreich, 1996). Risk of impaired microbial water quality is often elevated in
63 agricultural catchments due to the large range of FIO sources that are present (e.g. different livestock)
64 and the multiple transport mechanisms (e.g. overland flow, seepage from areas of degraded soil) that
65 can transfer FIOs to streams (Chadwick et al., 2008). Successful implementation of mitigation measures
66 in such environments requires the spatio-temporal dynamics of sources and transfer mechanisms
67 contributing FIOs to streams at the sub-field scale to be understood (Oliver et al., 2007, 2016; also c.f.
68 Greene et al., 2015; Vinten et al., 2017). This likely requires the integration of empirical data with
69 process-based FIO models (de Brauwere et al., 2014); however, uncertainties over the robustness of
70 hydrological processes simulated by such models and the coarse spatial discretisation they often employ
71 may hinder their use in understanding the sub-field-scale drivers of in-stream FIO dynamics that emerge
72 at the catchment scale (c.f. Rode et al., 2010; Wellen et al., 2015).

73

74 The overall aim of this paper is to outline a new Model for the Agent-based simulation of Faecal
75 Indicator Organisms (MAFIO); it is structured as follows. Section 2 reviews the pertinent limitations
76 of existing process-based FIO models before exploring how agent-based and tracer-aided modelling
77 approaches have potential in helping these to be addressed. This provides the rationale for the
78 development of MAFIO, which simulates the sub-field-scale fate and transport of agents representing
79 FIOs in a spatially-distributed and process-based manner, underpinned by a hydrological environment
80 generated by a robust hydrological model. A detailed model description is presented in Section 3.
81 Finally, Section 4 introduces the tracer-aided ecohydrological model EcH₂O-iso (Kuppel et al., 2018a)
82 as a potential example of a robust hydrological environment generator and details its coupling to
83 MAFIO. A companion paper (Neill et al., *in review*) provides a proof-of-concept application of MAFIO
84 focusing on process representation and the model's potential for use in a management context.

85

86 **2 Rationale for model development**

87 *2.1 Current limitations to simulating sources and transfer mechanisms contributing FIOs to streams at*
88 *the sub-field scale*

89 A prerequisite to using models to understand the drivers of dynamics in water quality parameters is
90 proper constraint of processes to which the parameters may be sensitive (Sokolova et al., 2018; Vaché
91 and McDonnell, 2006). For microbial water quality, hydrological processes are often hypothesised as a
92 major control on observed FIO dynamics (Kay et al., 2008; Oliver et al., 2005a; Tetzlaff et al., 2012).
93 However, catchment hydrological functioning in process-based FIO models can often only be
94 constrained through calibration to discharge data from a catchment outlet (Cho et al., 2016). Such data
95 primarily capture the speed at which perturbations (e.g. rainfall) to the system are transmitted through
96 the catchment, with limited insight into internal catchment states and fluxes (Birkel et al., 2014a;
97 McDonnell and Beven, 2014). Consequently, flow path dynamics and resulting hydrological
98 connectivity are likely to be poorly constrained in FIO models, potentially undermining their ability to
99 accurately represent hydrological mechanisms that transfer FIOs to streams (c.f. Sokolova et al., 2018;
100 Vaché and McDonnell, 2006; Wellen et al., 2015).

101

102 A further issue is coarse spatial discretisation in many process-based FIO models, with lumped (e.g.
103 Haydon and Deletic, 2006; Neill et al., 2019) to semi-distributed (e.g. Sadeghi and Arnold, 2002;
104 Whitehead et al., 2016) model structures generally being the norm. Whilst limited examples of fully-
105 distributed FIO models do exist, they have often been implemented at coarse resolutions (e.g. the 1 km²-
106 resolution WATFLOOD model of Dorner et al., 2006). Consequently, the spatial discretisation adopted
107 by many FIO models is often inconsistent with the scales at which heterogeneity in hydrological
108 processes and FIO fate and transport is expressed, and at which mitigation measures can be
109 implemented (c.f. Fatichi et al., 2016; Rode et al., 2010; Wellen et al., 2015).

110

111 Recent progress in spatially-distributed tracer-aided modelling has potential in simultaneously
112 overcoming issues of scale and hydrological process realism in FIO models (see Section 2.3). However,
113 spatially-distributed simulation of FIO dynamics alone is unlikely to be sufficient for fully
114 understanding sources and transfer mechanisms contributing FIOs to streams; this requires explicit
115 tracking of FIOs as they move through a catchment. As summarised by Reaney (2008), spatially-
116 distributed models conventionally simulate fluxes into and out of individual grid cells, but no
117 information is available on the spatial origin of the constituents (water/contaminants) comprising those
118 fluxes; consequently, it is often only possible to simulate the locations of active fluxes and what their
119 magnitude is, but not actually whether those fluxes contribute water/contaminants to the stream. A
120 possible solution, which could also have value in simulating the heterogenous behaviour of FIOs of
121 different types or from different host animals (e.g. die-off kinetics: Avery et al., 2004), is the
122 incorporation of agent-based methods into spatially-distributed modelling frameworks (Reaney, 2008).

123

124 *2.2 The potential of agent-based modelling approaches*

125 Agent-based models (ABMs) comprise three major elements: 1) agents representing autonomous
126 individuals, each associated with a set of attributes and the ability to sense and process information
127 regarding their surroundings; 2) a simulation environment that can be distributed in space in which the
128 agents operate; 3) a set of rules, conditional or stochastic in nature, defining how agents interact with
129 the environment and with each other based on their attributes (Abdou et al., 2012; Crooks and
130 Heppenstall, 2012). Agent attributes can relate to any characteristic necessary to simulate the
131 behaviours of the phenomena being studied or that is of interest for the problem being addressed (Macal
132 and North, 2010). Whilst often more demanding than aggregative modelling approaches in terms of
133 computation, data requirements and evaluation, the bottom-up approach of ABMs allows greater scope
134 for representing heterogeneity amongst individual agents and their interactions with the simulation
135 environment, permitting richer simulation of small-scale behaviours and how these cause higher-level
136 system dynamics to emerge (O’Sullivan et al., 2012).

137

138 Use of ABMs for the process-based simulation of water quantity/quality dynamics has been relatively
139 limited. However, there are promising examples of the development of such models to elucidate spatio-
140 temporal patterns of surface hydrological connectivity through tracking locational attributes of agents
141 representing water particles as they move through catchments (Reaney, 2008), and to simulate the
142 hydrologically-induced erosion and transport of markers (similar to agents) representing sediment
143 particles based on their unique characteristics (Cooper et al., 2012). Furthermore, several studies have
144 successfully implemented closely-related yet simpler (Crooks and Heppenstall, 2012) cellular automata
145 models to simulate hydrological processes (e.g. Cirbus et al., 2013; Hodge and Hoey, 2012; Ravazzani
146 et al., 2011; Shao et al., 2015). Consequently, representing FIOs as agents within an ABM could have
147 significant potential for simulating their fate and transport, with agent attributes governing how they
148 interact with different transfer mechanisms operating in a simulated environment and permitting their
149 pathways through catchments to be tracked.

150

151 *2.3 Progress in spatially-distributed, tracer-aided (eco)hydrological modelling*

152 The dynamics of stable isotopes (^2H and ^{18}O) in streamwater contain information with respect to the
153 velocities of water passing through a catchment, as influenced by factors such as hydrological
154 connectivity, storage and mixing (Birkel and Soulsby, 2015; McDonnell and Beven, 2014).
155 Consequently, incorporation of isotopes as tracers into hydrological models allows performance
156 assessments to advance from how well a model fits an observed hydrograph, to also how consistent the
157 model is with internal catchment processes giving rise to the velocity response (Birkel et al., 2014a;

158 McDonnell and Beven, 2014). Improved confidence in the realistic simulation of catchment functioning
159 afforded by such tracer-aided hydrological models has facilitated their use in providing valuable
160 insights into how catchments store and release water (Birkel and Soulsby, 2015), and how these factors
161 drive various water quality parameters including dissolved organic carbon (Birkel et al., 2014b; Dick
162 et al., 2015) and FIOs (Neill et al., 2019). Recently, tracer-aided models have evolved from their early
163 lumped conceptual structures towards ones that are fully-distributed and increasingly physically-based
164 (e.g. Kuppel et al., 2018a; Remondi et al., 2018; van Huijgevoort et al., 2016), reflecting cheaper isotope
165 analysis (e.g. Berman et al., 2013) and recognition of the need for catchment systems to be modelled in
166 this way to tackle issues of water quality and environmental change (Fatichi et al., 2016). Such models
167 have permitted more nuanced simulation of catchment-scale discharge and tracer dynamics, and also
168 facilitate specific interrogation of intra-catchment water and isotope dynamics to further increase
169 confidence that internal catchment processes are being correctly represented (Ala-aho et al., 2017;
170 Knighton et al., 2020; Kuppel et al., 2018a; Piovano et al., 2018, 2019; Remondi et al., 2018; Smith et
171 al., 2019).

172

173 In addition to the value of tracers, the importance of resolving how water is partitioned between “green”
174 (evaporation and transpiration) and “blue” (discharge and groundwater recharge) hydrological fluxes
175 when representing the storage and release of water by catchments is increasingly recognised (Brooks et
176 al., 2015; Falkenmark and Rockström, 2006). Most tracer-aided models only explicitly simulate “blue”
177 fluxes, with “green” fluxes specified as a combined variable usually partitioned based on model
178 parameters (Fatichi et al., 2016; Vivoni, 2012). However, tracer-aided ecohydrological models are
179 emerging that explicitly resolve both “blue” and “green” fluxes whilst also simulating stable isotope
180 dynamics to enable further evaluation of process consistency (Kuppel et al., 2018a; Maneta and
181 Silverman, 2013). Such a model, EcH₂O-iso (Kuppel et al., 2018a), is introduced in Section 4 as an
182 example of a hydrological environment generator for MAFIO. However, it is stressed that any model
183 can be used, providing it can be robustly determined as to whether catchment hydrological functioning
184 is being successfully captured.

185

186 **3 Presentation of MAFIO**

187 This presentation follows the ODD (Overview, Design Concepts, Details) protocol for describing agent-
188 based models (Grimm et al., 2006, 2010). MAFIO is written in the Python 3.6 programming language
189 and makes use of the PCRaster module (Karssenberget al., 2010) for handling spatial inputs and
190 outputs. Given that it can be coupled with any robust hydrological model that provides the necessary

191 outputs, the following presentation of MAFIO is provided in a generic form, with Section 4 outlining
192 the specific interfacing with EcH₂O-iso.

193

194 *3.1 Purpose*

195 The purpose of this initial version of MAFIO is to reveal the spatio-temporal dynamics of sources and
196 transfer mechanisms contributing FIOs to streams at the sub-field scale in small (< 10 km²) agricultural
197 catchments. This is achieved by simulating and tracking, in a spatially-distributed, process-based
198 manner, the fate and transport of agents representing FIOs shed by different types of livestock (Figure
199 1).

200

201 *3.2 State variables and scales*

202 MAFIO consists of four entities. Two environments drive the simulation: a catchment environment and
203 a hydrological environment. These set the attributes of a spatial grid on which simulations take place
204 over discrete timesteps. The behaviour of agents representing FIOs (FIO-agents) is then simulated based
205 on their individual attributes and those of the spatial grid for each timestep.

206

207 *3.2.1 Catchment environment*

208 The catchment environment defines, on a regular grid, the following fixed spatial characteristics of the
209 catchment being modelled:

- 210 1) Spatial extent;
- 211 2) Distribution of land parcels, that may be delineated based on land uses (e.g. pastoral field, arable
212 field) or covers (e.g. forest, moorland);
- 213 3) Local flow direction based on the D8 algorithm (O'Callaghan and Mark, 1984);
- 214 4) Location and width of the stream channel;
- 215 5) Locations where soil is degraded adjacent to the channel;
- 216 6) Grid cells immediately upslope of those containing a channel;
- 217 7) Grid cells immediately upslope of those containing degraded soil;
- 218 8) Vertical discretisation of the landscape, currently limited to the land surface and the first few
219 centimetres of the upper soil profile (exact depth unspecified) where it is assumed most
220 interactions of FIOs with the soil will occur (Stocker et al., 2015).

221 Additionally, counts of each livestock type (e.g. sheep, cattle) considered in the simulation for each
222 land parcel, and which areas of the stream are accessible to livestock, are specified, both of which vary
223 in time.

224

225 3.2.2 Hydrological environment

226 The hydrological environment (Figure 2) is provided by a spatially-distributed hydrological model that
227 can simulate the following on a regular spatial grid: effective precipitation (P_{Eff}), effective solar
228 radiation (SR_{Eff}), ponded surface water prior to infiltration ($Pond$), infiltration flux ($Infil$), soil water
229 content prior to exfiltration (SWC_{Exf}), exfiltration flux ($Exfil$), soil saturation deficit (scaled between 0
230 for fully saturated and 1 for only residual soil moisture present; Sat_{Def}) and soil skin temperature (T_{Skin}).
231 Here, effective precipitation is total precipitation minus interception, whilst effective solar radiation is
232 that absorbed by the ground after accounting for transmission and reflective losses. Whilst not explicitly
233 part of the hydrological environment, run-on and run-off are accounted for implicitly by assuming that
234 any water remaining on the surface following infiltration/exfiltration runs off to contribute to $Pond$ of
235 the next downslope grid cell (consistent with EcH₂O-iso). The hydrological environment gives the
236 following abstract conceptualisation of catchment hydrological functioning from the perspective of
237 MAFIO (Figure 2). For a given grid cell, there are two hydrological stores: Ponded Surface Water and
238 Soil Water. First, P_{Eff} and run-on from upslope cells enter Ponded Surface Water to define $Pond$. Water
239 can then infiltrate ($Infil$) into Soil Water to increase SWC_{Exf} and reduce Sat_{Def} , after which water can
240 exfiltrate ($Exfil$) back to Ponded Surface Water if the soil is now saturated (i.e. $Sat_{Def} = 0$). Finally, any
241 remaining water in Ponded Surface Water is routed to the next downslope cell unless a channel is
242 present, in which case remaining water is routed to the stream and out of the catchment. It should be
243 noted that this conceptualisation only includes stores and fluxes of water relevant to the transport of
244 FIOs in naturally-drained environments; however, future iterations of MAFIO may look to allow for
245 artificial drainage as a potentially-efficient FIO transport pathway (Oliver et al., 2005b).

246

247 3.2.3 Timestep

248 MAFIO operates on discrete timesteps consistent with the fixed temporal resolution of variables defined
249 by the aforementioned environments. Here, a daily timestep is used. However, if needed to account for
250 non-linearities in observed FIO dynamics (McKergow and Davies-Colley, 2010), sub-daily timesteps
251 could be used with appropriate changes to parameter values and sufficient computing power.

252

253 3.2.4 Spatial grid

254 Simulations take place on a regular spatial grid (Figure 1), with each grid cell having fixed and dynamic
 255 attributes (Table 1). The catchment environment determines values of fixed attributes. The position of
 256 a cell within the environment (*X-Y location* [Loc_{Att}]) determines values for *Channel cell* ($chan_{Att}$), *Flow*
 257 *direction* ($fDir_{Att}$), and *Land parcel(s)* (LP_{Att}). Multiple land parcels can be specified for a cell containing
 258 a channel to indicate those on either side of the channel. The attribute *Cell size* ($size_{Att}$) is determined
 259 by the spatial resolution of the catchment and hydrological environments. However, to allow for spatial
 260 resolutions too coarse to resolve small channels (Zhang and Montgomery, 1994) and processes
 261 occurring at the land-water interface that are possibly crucial for water quality (e.g. Karr and Schlosser,
 262 1978), sub-grid heterogeneity can be represented in cells containing a channel. These have the
 263 additional fixed attributes *Proportion of cell area occupied by channel* (P_Chan_{Att}) and *Degraded soil*
 264 *in cell* ($degSoil_{Att}$), to allow for the channel not occupying the whole cell and the presence of degraded
 265 soil in the vicinity of the channel from livestock compaction, respectively. The latter may result in the
 266 chronic seepage of water and FIOs to the stream (Bilotta et al., 2007). P_Chan_{Att} is calculated by
 267 multiplying the width and straight-line length ($streamDist$, equal to either $size_{Att}$ or $[2 \times size_{Att}^2]^{0.5}$)
 268 depending on $fDir_{Att}$ of the channel occupying a cell, then dividing by cell area.

269

270 The dynamic attributes of the spatial grid are calculated for each timestep. They are predominantly
 271 derived from the hydrological environment (Figure 3) and are largely responsible for the behaviour of
 272 FIO-agents before entering the channel. *Effective precipitation* (P_{Att}), *Effective solar radiation* (SR_{Att})
 273 and *Soil skin temperature* (TS_{Att}) are equal to P_{Eff} , SR_{Eff} and T_{Skin} , respectively. The attributes *Probability*
 274 *of exfiltration* (P_Exfil_{Att}) and *Probability of infiltration* (P_Infil_{Att}) are defined as:

275

$$276 \quad P_Exfil_{Att} = \frac{Exfil}{SWC_{Exf}} \quad Eq. 1$$

$$277 \quad P_Infil_{Att} = \frac{Infil}{Pond} \quad Eq. 2$$

278

279 Figure 3a gives a visual example of how hydrological environment variables translate to attributes of
 280 the spatial grid. Since sub-grid heterogeneity is represented in MAFIO but not necessarily in the
 281 hydrological environment simulator, dynamic attributes of cells containing a channel may need
 282 derivation from variables simulated for neighbouring cells. This could involve taking average values
 283 from upslope cells which drain into the cell in question (Figure 3a); conceptually, this implies that the
 284 land surface adjacent to the channel implicitly represented in MAFIO is an aggregated extension of the
 285 upslope cells.

286

287 The attribute *Probability of seepage* ($P_{SeepAtt}$) uses variables of the catchment and hydrological
288 environments to encapsulate the combined effects of livestock grazing pressure and soil moisture on
289 soil degradation (Drewry, 2006) and the associated potential for FIO seepage into streams (Bilotta et
290 al., 2007; Tetzlaff et al., 2012). To indicate livestock grazing pressure, livestock count(s) for the present
291 timestep for land parcels(s) associated with cells containing degraded soil are converted into livestock
292 units per hectare ($LU\ ha^{-1}$; Natural England, 2013). This variable is then classified into a band (“low”,
293 “medium” or “high” - see Section 3.7.2.4) tied to a certain fraction of soil that could be degraded on the
294 current timestep (D_{frac_new} ; c.f. Sheath and Carlson, 1998); if no livestock are present, D_{frac_new} is 0.
295 However, since soil takes time to recover once degraded, the final fraction of degraded soil (D_{frac}) used
296 for the timestep is the maximum of D_{frac_new} and the damage fraction of the previous timestep (D_{frac_old})
297 subjected to exponential decay (Figure 3b; c.f. Elliot et al., 2002); the latter occurs at a rate specified
298 by the parameter k_d in units of T^{-1} , where T is the length of a timestep:

299

$$300 \quad D_{frac} = \max\{D_{frac_new}, D_{frac_old} \cdot e^{-k_d T}\} \quad Eq. 3$$

301

302 The value of D_{frac} is then scaled by the average Sat_{Def} of cells immediately upslope (as cells with
303 degraded soil also contain a channel) to account for the impact of moisture content in the extent of soil
304 degradation (Drewry, 2006), and to give the final probability of seepage (Figure 3b):

305

$$306 \quad P_{SeepAtt} = D_{frac} \cdot (1 - Sat_{Def}) \quad Eq. 4$$

307

308 3.2.5 FIO-agents

309 FIOs in MAFIO are represented by FIO-agents. Given likely large numbers of FIOs present within
310 agricultural catchments, it is impossible for each FIO to be represented by a single FIO-agent.
311 Consequently, MAFIO initially adopts the simple approach of assuming that the overall behaviour of a
312 population can be inferred from simulating a suitably-sized sample (Parry and Bithell, 2012; Reaney,
313 2008). FIO-agents are characterised by dynamic, fixed and memory attributes (Table 2). Dynamic
314 attributes can be updated multiple times within a timestep due to die-off and transport processes.
315 *Attached-stream sediment* and *Detached-faeces* indicate whether an FIO-agent is attached to stream
316 sediment or detached from a faecal deposit, respectively. The former determines how the FIO-agent is
317 routed through the stream, whilst the latter specifies whether an FIO-agent is available for transport.

318 *Domain type* accounts for the position of the FIO-agent at the sub-grid scale (either in a cell without a
319 channel or otherwise its location within a cell containing a channel). *Infiltration stage* denotes where in
320 the vertical discretisation of the catchment the FIO-agent is, which will affect the die-off and transport
321 processes it can experience. *Life state* and *Movement tracker* follow whether the FIO-agent is alive or
322 dead (to determine whether it is still tracked) and if the FIO-agent moved in the current timestep,
323 respectively. Finally, *Timestep*, *X co-ordinate* and *Y co-ordinate* specify the current timestep, and X
324 and Y co-ordinates of the FIO-agent; these allow recording of when changes to certain dynamic
325 attributes occur in the simulation and where in the spatial grid the FIO-agent is situated. Memory
326 attributes keep track of how dynamic attributes of the FIO-agents change over the simulation; this
327 permits tracking of, for example, the origin of an FIO-agent that reaches the stream or the path that it
328 took to the stream. Fixed attributes are assigned when the FIO-agent spawns and record its original host
329 animal (*Livestock type*) and the land parcel into which it spawned (*Land parcel*). At present, MAFIO is
330 configured to consider only a single type of FIO (*E. coli*); however, an additional fixed attribute for
331 FIO-agents specifying their FIO type could easily allow representation of multiple FIO types with
332 specific parameterisations.

333

334 *3.3 Process overview and scheduling*

335 Process scheduling during each timestep is shown in Figure 4a. First, environmental variables define
336 dynamic attributes of the spatial grid, which is loaded in. Next, the Defecation Sub-model (Section
337 3.7.1) simulates FIO loading by spawning FIO-agents in each land parcel in a set order, based on their
338 associated livestock counts. When new FIO-agents are spawned, their fixed attributes and initial values
339 for dynamic attributes are set. Next, for each FIO-agent in turn (Figure 4b), behaviours resulting from
340 fate and transport processes operating at the sub-field scale during the current timestep are simulated
341 via the small-scale behaviour sub-models (Section 3.7.2). Once the behaviour of all FIO-agents has
342 been simulated, those with *Life state* = 0 are removed during a “clean-up” operation, before the spatio-
343 temporal outputs for the current timestep are finally saved (Figure 4a).

344

345 For each FIO-agent, the sequence of small-scale behaviour sub-models that runs depends on certain
346 attribute values (Figure 4b), and may include Die-off (Section 3.7.2.1; Figure 5), Detachment (Section
347 3.7.2.2; Figure 6), Surface Routing (Section 3.7.2.3; Figure 7), Seepage (Section 3.7.2.4; Figure 8) and
348 Channel Routing (Section 3.7.2.5; Figure 9). As indicated in Figures 5-9, different sub-models utilise
349 various attributes of the spatial grid as input and have the potential to update the dynamic and associated
350 memory attributes of the FIO-agent. Memory attributes are also updated for the start of a new timestep
351 if the FIO-agent was already present in the model domain (Figure 4b). There are six conditions that will
352 cause MAFIO to move on to simulate the next FIO-agent: 1) the Die-off Sub-model results in the FIO-

353 agent dying; 2) the FIO-agent is still attached to a “faecal deposit” after running the Detachment Sub-
354 model and hence cannot move during the current timestep; 3) the FIO-agent is in the soil but does not
355 exfiltrate when the Surface Routing Sub-model is run on the current timestep; 4) the FIO-agent remains
356 in an area of degraded soil following the Seepage Sub-model being run on the current timestep; 5) the
357 agent is deposited on the stream bed as it is routed through the channel by the Channel Routing Sub-
358 model; or 6) the FIO-agent is successfully routed to the catchment outlet by the Channel Routing Sub-
359 model (Figure 4b).

360

361 *3.4 Design concepts*

362 Of the 11 design concepts outlined in the ODD protocol, only basic principles, emergence, sensing,
363 interaction, stochasticity and observation are relevant to MAFIO.

364

365 *3.4.1 Basic principles*

366 The process-based, spatially-distributed nature of MAFIO reflects the need for such models to further
367 quantitative process understanding of water quality dynamics (de Brauwere et al., 2014) and to
368 overcome limitations associated with lumped/semi-distributed model structures (Section 2.1). The
369 principles underlying the process-based simulation of FIO loading, die-off and transport within MAFIO
370 are as follows (Figure 1).

371

372 The Defecation Sub-model (Section 3.7.1) conceptualises FIO loading by livestock, assumed to be the
373 main source of FIOs in agricultural areas (Chadwick et al., 2008). Loading by different livestock types
374 can be simulated to account for differences in defecation characteristics (e.g. concentration of FIOs in
375 faeces; Oliver et al., 2018) and FIO behaviour (e.g. die-off kinetics; Avery et al., 2004). FIO loading
376 can be onto the land or directly into the channel via direct deposition (Chadwick et al., 2008). The
377 potential for the latter depends on farm management, such as whether streams are fenced/gated off from
378 the surrounding fields (Kay et al., 2007; Vinten et al., 2008). A sample of the FIOs loaded into the
379 environment by individual animals is simulated in MAFIO by introducing one FIO-agent into the
380 simulation for every specified number of FIOs loaded in reality (c.f. Parry and Bithell, 2012). This
381 ensures that FIO-agents are introduced in correct proportions to account for different loadings by
382 individual animals and livestock types. The accuracy with which the behaviour of the true population
383 of FIOs can be inferred and resolved will vary with the number of FIOs for which an FIO-agent is
384 introduced and the consequent total number of FIO-agents in the simulation (c.f. Parry and Bithell,
385 2012; Reaney, 2008).

386

387 The Die-off Sub-model (Section 3.7.2.1) conceptualises the die-off of FIOs in response to temperature
388 and solar radiation once in the environment; these factors are often considered the most important in
389 determining FIO mortality (Blaustein et al., 2013; Hipsey et al., 2008). Both are used to determine the
390 die-off of FIO-agents on the land surface, whilst only temperature is used for die-off of FIO-agents in
391 the soil (Whitehead et al., 2016). Given the likely short (<1 day) travel-times of water in small
392 agricultural streams, it is assumed that die-off of FIOs once in the stream is negligible. It may also be
393 possible for FIOs to grow in the environment (e.g. Muirhead, 2009; Soupir et al., 2008). However, given
394 the limited extent to which this process is understood and the consequent uncertainty (e.g. Hipsey et al.,
395 2008; Oliver et al., 2010), it is not considered in this initial development of MAFIO.

396

397 The Detachment Sub-model (Section 3.7.2.2) simulates the release of FIO-agents from faecal deposits
398 using a model based on effective precipitation depth and the assumption that this metric is sufficient to
399 allow observed release kinetics to be captured (Blaustein et al., 2015b). Once released, lateral transport
400 of FIOs can occur via overland flow, whilst infiltration may cause FIOs to move vertically into the soil
401 where soil retention effects including straining and filtering may limit their further transport (Oliver et
402 al., 2005a). Consequently, lateral movement of FIO-agents in MAFIO is only possible via overland
403 flow; infiltrating FIO-agents are retained in the soil. However, FIOs are generally retained in the upper
404 few centimetres of the soil, which may be within the “effective depth of interaction” of the soil with
405 surface flows that could facilitate lateral movement of the FIOs (Stocker et al., 2015). To account for
406 this, infiltrated FIO-agents can be exfiltrated back to the surface for transport in overland flow. These
407 processes of lateral movement, infiltration and exfiltration are conceptualised within the Surface
408 Routing Sub-model (Section 3.7.2.3).

409

410 In agricultural settings, compaction in areas of animal congregation or locations where there is a high
411 frequency of livestock traffic (e.g. gateways between fields) can cause soil degradation (Bilotta et al.,
412 2007). If proximal to the channel, this can increase the likelihood that these low permeability areas will
413 seep water and contaminants to the stream, potentially causing them to act as chronic sources of FIOs
414 (Davies-Colley et al., 2004). For cells containing a channel where degraded soil is present, the Seepage
415 Sub-model (Section 3.7.2.4) simulates this transfer to the stream.

416

417 Once in the stream, FIOs have an increased likelihood of settling to the channel bed if attached to
418 sediment (Pachepsky and Shelton, 2011). Several attempts have been made to model the settling of
419 sediment-associated FIOs (e.g. Collins and Rutherford, 2004; Hipsey et al., 2008; Jamieson et al., 2005;

420 Kim et al., 2010). However, substantial uncertainty still surrounds exactly how the process of settling
421 should be represented (Pachepsky and Shelton, 2011; Pandey et al., 2012). Consequently, a simple
422 distance-decay approach is initially adopted in MAFIO, with the likelihood of an FIO-agent settling to
423 the streambed increasing with distance travelled within the stream. The exact form of the relationship
424 can be assumed to implicitly account for different factors (e.g. settling velocity) affecting the likelihood
425 of settling (Kay and McDonald, 1980). The association of FIO-agents with stream sediment and their
426 resulting transport in the stream is simulated by the Channel Routing Sub-model (Section 3.7.2.5).
427 Whilst resuspension of FIOs has been found to contribute to stream FIO dynamics (Pachepsky and
428 Shelton, 2011), this is not currently included in MAFIO as in the relatively low-energy streams
429 characteristic of small agricultural catchments, settling will be the dominant process except in the
430 largest flow events (c.f. Nagels et al., 2002). Therefore, only settling was considered to assess whether
431 this is sufficient to capture observed FIO dynamics (c.f. Clark et al., 2011).

432

433 3.4.2 Emergence

434 The model environments impose certain constraints on how simulations can unfold. For example,
435 livestock counts, stream accessibility to livestock and locations of degraded soil determine into which
436 land parcels FIO-agents can be spawned, where direct deposition may occur and where FIO-agents may
437 chronically seep to the stream, respectively; thus, the field-scale distribution of *possible* FIO source
438 areas is relatively imposed. Furthermore, the spatial distribution and magnitude of hydrological fluxes
439 dictate where there are active flow paths capable of transporting FIO-agents. However, it is from
440 simulating the small-scale behaviour of FIO-agents and their interaction with the variables of the model
441 environments that the following simulated characteristics emerge: 1) The spatial distribution of FIO-
442 agents stored on the surface and in the soil at the sub-field scale; 2) Fluxes of FIO-agents in the stream;
443 3) The pathways taken by FIO-agents from their source areas to the stream; 4) Time-varying
444 contributions of FIO-agents to the stream made by different transfer mechanisms (overland flow,
445 seepage and direct deposition) and host livestock; 5) The total number of FIO-agents active in the model
446 domain at a given time.

447

448 3.4.3 Sensing and interaction

449 The behaviour of an FIO-agent is simulated based on it knowing its own attributes and those of the grid
450 cell it currently occupies. Consequently, there are no interactions between FIO-agents, with all
451 behaviours happening independently. A one-way interaction occurs between the FIO-agents and the
452 spatial grid, with the FIO-agents responding to the conditions in the grid cell they occupy.

453

454 3.4.4 Stochasticity

455 MAFIO employs the following stochastic elements. In the Defecation Sub-model, uniform random
456 sampling (c.f. Dorner et al., 2006; Haydon and Deletic, 2006) determines the spatial grid cells into
457 which new FIO-agents are spawned to reflect natural variability (Benhamou, 2006, 2013) and
458 uncertainty in livestock movement and consequent defecation locations at the sub-field scale. If an FIO-
459 agent spawns in a cell containing a channel to which livestock have access, stochasticity is used to
460 ensure that direct deposition occurs with a frequency that reflects the size of the channel relative to the
461 total cell area. Whilst the number of FIOs shed by individual animals of different livestock types is
462 currently deterministic (i.e. relies on fixed parameters to specify factors such as FIO concentrations in
463 faeces, allowing MAFIO to be used in data-limited situations), variability and uncertainty in this
464 variable could be accounted for stochastically where a basis exists for defining reasonable sampling
465 distributions for relevant parameters (e.g. Schijven et al., 2015).

466

467 The Die-off, Detachment and Channel Routing Sub-models employ stochasticity to define the
468 behaviour of individual FIO-agents based on governing equations that describe population-level
469 behaviour. In the Surface Routing Sub-model, stochasticity conserves “concentrations” of FIO-agents
470 in hydrological stores (i.e. Poned Surface Water and Soil Water) and fluxes (e.g. infiltration,
471 exfiltration). With the assumption that FIO-agents are fully and uniformly mixed within a hydrological
472 store, probabilities define the behaviour of individual FIO-agents such that when a hydrological flux is
473 generated, the proportion of water from the store that generates the flux is equal to the proportion of
474 FIO-agents in the store that it transports. Finally, the Seepage Sub-model employs stochasticity to
475 represent the general tendency for increased seepage potential as soils degrade in response to greater
476 livestock compaction and wetness, without explicitly representing poorly-defined seepage processes
477 (c.f. Collins and Rutherford, 2004).

478

479 3.4.5 Observation

480 For each timestep, the simulated flux of FIO-agents in the stream is recorded for each cell containing a
481 channel. This facilitates comparison with spatially-distributed observations of stream FIO loads. For
482 FIO-agents ultimately reaching the catchment outlet, the following attributes are observed: *Domain*
483 *type*, *Livestock type*, *Location memory* (Table 2). These observations are fundamental to MAFIO
484 fulfilling its stated purpose (Section 3.1) as they allow derivation of the transfer mechanisms, host
485 animals and pathways contributing FIOs to the stream at each timestep. The number and spatial
486 distribution of active FIO-agents in the model domain is also observed at each timestep so FIO-agent
487 fluxes can be linked to storage dynamics. Simulated outputs should be observed for an ensemble of

488 model runs using the same inputs and parameterisation to account for the effects of stochasticity in
489 MAFIO (Abdou et al., 2012).

490

491 *3.5 Initialisation*

492 For the first timestep, an initial value of D_{frac} must be specified for each cell containing degraded soil
493 to account for previous actions of livestock. Ideally, values should be based on livestock counts prior
494 to the simulation start date; otherwise representative average counts (e.g. from the simulation period)
495 may be used. In addition, each land parcel may be initialised with FIO-agents in the soil to allow for
496 the possibility that FIOs from soil reservoirs reflecting past grazing patterns (which may persist for
497 several months) contribute to FIO dynamics of the simulation period (Muirhead, 2009). Specifically, a
498 number of FIO-agents can be randomly seeded in the soil of each land parcel at the start of the
499 simulation based on estimates of FIOs in the soil derived from concentrations measured in the upper
500 soil profiles of each land parcel, past livestock counts or literature values. Whilst initial values of D_{frac}
501 and the number of FIO-agents to be seeded in the soil are consistent between simulations, the spatial
502 locations of the latter are determined stochastically and therefore vary.

503

504 *3.6 Inputs*

505 MAFIO requires input data to characterise the catchment and hydrological environments. Table 3
506 documents those required for the catchment environment. The hydrological environment is
507 characterised using inputs from an appropriate hydrological model, as detailed in Section 3.2.2.
508 Spatially distributed inputs are provided as map files (.map) compatible with PCRaster (Karssenberget
509 al., 2010).

510

511 *3.7 Sub-models*

512 MAFIO consists of the Defecation Sub-model and several small-scale behaviour sub-models (Figure
513 4). The required parameters and their initial values based on daily simulation of *E. coli* from sheep and
514 cattle are given in Table 4. In the following equations, a sub-script $[LS]$ denotes that the value of the
515 variable or parameter varies based on the livestock type. Meanwhile, a sub-script $[xy]$ indicates an
516 attribute or derived variable of the spatial grid whose value varies depending on the cell.

517

518 *3.7.1 Defecation Sub-model*

519 The Defecation Sub-model runs once every timestep (Figure 4a). The parameter *lsType* indicates which
 520 livestock are being considered in the simulation. For each livestock type, the number of FIO-agents
 521 introduced into the simulation for each defecation of an individual animal is calculated. First, the
 522 concentration of FIOs per gram of faeces is specified by the parameter *faecesConc*. Total FIO loading
 523 per defecation (*FIO_Load*) is then calculated as:

524

$$525 \quad FIO_Load_{[LS]} = faecesConc_{[LS]} * FWeight_{[LS]} \quad Eq. 5$$

526

527 where *FWeight* is the weight of a single defecation in grams. This value is then translated into a number
 528 of FIO-agents introduced into the simulation per defecation (*LS_Agents*), based on a specified number
 529 of FIOs assumed to be represented by an FIO-agent (*agentsRepresent*):

530

$$531 \quad LS_Agents_{[LS]} = \left\lceil \frac{FIO_Load_{[LS]}}{agentsRepresent} \right\rceil \quad Eq. 6$$

532

533 For each livestock type, the Defecation Sub-model then introduces FIO-agents to land parcels based on
 534 livestock counts for the current timestep. For each individual of a given livestock type, the number of
 535 defecations per timestep is specified by *defecationsPerDay*. For each defecation, a cell of the current
 536 land parcel is chosen using uniform random sampling. At this location, the number of FIO-agents
 537 specified by *LS_Agents* are spawned. If the cell contains a channel (specified by *ChanAtt* of the cell)
 538 accessible to livestock, the probability that the FIO-agents spawn in the channel to simulate direct
 539 deposition is:

540

$$541 \quad P(Channel) = P_ChanAtt_{[xy]} \quad Eq. 7$$

542

543 If they do not spawn in the channel or livestock do not have channel access, the FIO-agents either spawn
 544 in an area of degraded soil if present (specified by *degSoilAtt*), or on the land adjacent to the channel.
 545 Whilst the fixed nature of *faecesConc*, *FWeight*, and *defecationsPerDay* dictates that the number of
 546 FIO-agents introduced by individual animals of a given livestock type is deterministic, as noted in
 547 Section 3.4.4, stochastic treatment of this variable is possible if data exist to constrain appropriate
 548 sampling distributions for the aforementioned parameters.

549

550 3.7.2 Small-scale behaviour sub-models

551 The small-scale behaviour sub-models simulating FIO-agent behaviour resulting from fate and transport
552 processes operating at the sub-field scale are applied every timestep to each FIO-agent in the model
553 domain in turn (Figure 4a). The exact sequence of sub-models that runs depends on certain attributes
554 of the FIO-agent in question (Figure 4b). MAFIO will move on to simulate the behaviour of the next
555 FIO-agent only once one of the six conditions in Section 3.3 are met.

556

557 3.7.2.1 Die-off Sub-model

558 The Die-off Sub-model (Figure 5) only runs for FIO-agents that spawned before the current timestep
559 (Figure 4b), with die-off modelled based on first-order kinetics following Chick's Law (Chick, 1908):

560

$$561 \frac{N_{Alive}}{N_0} = e^{-k_{dead} \cdot T} \quad Eq. 8$$

562

563 where N_{Alive}/N_0 is the fraction of the original population of FIOs (N_0) that is still alive (N_{Alive}) after being
564 subject to exponential decay over a timestep, T , at a rate specified by the inactivation rate constant
565 parameter k_{dead} with units of T^{-1} . To translate this population-level behaviour to individual FIO-agents,
566 a probability that an FIO-agent dies is derived from Eq. 8 that is a function of the attributes of the grid
567 cell it occupies and its host animal:

568

$$569 P(Dies) = 1 - e^{-k_{dead}[LS][xy] \cdot T} \quad Eq. 9$$

570

571 Eq. 9 assumes all FIO-agents from a given livestock type within a cell can be treated as a single
572 population experiencing the same conditions and probability of die-off. The parameter k_{dead} is an overall
573 inactivation rate constant obtained by summing the individual inactivation rate constants arising from
574 factors (temperature and solar radiation in MAFIO) assumed to cause die-off (Chapra, 1997). The
575 inactivation rate constant due to temperature (k_{temp}) is based on the equation proposed by Mancini
576 (1978):

577

578 $k_{temp_{[LS][xy]}} = k_{0_{[LS]}} \cdot \theta_{[LS]}^{T_{SAtt[xy]} - 20}$ Eq. 10

579

580 where k_0 is the inactivation rate constant (units of T^{-1}) at a reference temperature of 20 °C, and θ is a
 581 temperature sensitivity parameter (dimensionless). Meanwhile, the inactivation rate constant due to
 582 solar radiation (k_{solar}) is calculated as (Thomann and Mueller, 1987):

583

584 $k_{solar_{[LS][xy]}} = \alpha_{SR} \cdot SR_{Att[xy]}$ Eq. 11

585

586 where α_{SR} is a proportionality constant. When solar radiation is expressed in units of $ly\ hr^{-1}$, α_{SR} is
 587 approximately unity (Thomann and Mueller, 1987). The final value of k_{dead} depends on whether the
 588 FIO-agent is in the soil (Eq. 12a) or on the surface (Eq. 12b):

589

590 $k_{dead_{[LS][xy]}} = k_{temp_{[LS][xy]}}$ Eq. 12a

591 $k_{dead_{[LS][xy]}} = k_{temp_{[LS][xy]}} + k_{solar_{[LS][xy]}}$ Eq. 12b

592

593 If an FIO-agent dies, its attribute *Life state* is set to “0” so that it is no longer moved or tracked for the
 594 rest of the simulation.

595

596 3.7.2.2 Detachment Sub-model

597 The Detachment Sub-model (Figure 6) only runs for FIO-agents with the attribute *Detached-faeces* =
 598 “0” and *Life state* = “1” (Figure 4b). In MAFIO, the exponential model of Bicknell et al. (1997)
 599 simulates the rainfall-induced release of FIOs from faeces:

600

601 $\frac{N_{Release}}{N_{Faeces}} = 1 - e^{-k_{release} \cdot P}$ Eq. 13

602

603 where $N_{Release}/N_{Faeces}$ is the fraction of FIOs in the faeces (N_{Faeces}) released ($N_{Release}$), $k_{release}$ is a rate
 604 constant with units of cm^{-1} and P is precipitation (centimetres). This is used because it is a single-

605 parameter model that has not been shown to be substantially worse than alternative multi-parameter
606 models (Blaustein et al., 2015a, 2016). From Eq. 13, the probability that an FIO-agent is detached from
607 its faecal deposit is derived as:

608

$$609 \quad P(\text{Detached}_{\text{Faeces}}) = 1 - e^{-k_{\text{release}} \cdot P_{\text{Att}}[xy]} \quad \text{Eq. 14}$$

610

611 Despite the value of k_{release} likely varying with livestock type (e.g. reflecting differently-composed
612 faeces; c.f. Hodgson et al., 2009), a current lack of information (c.f. Sokolova et al., 2018) dictates this
613 parameter be specified for the type of FIO being modelled. If the FIO-agent is detached, then *Detached-*
614 *faeces* is set to “1” so that the Surface Routing Sub-model can run (Figures 4b and 6). Otherwise, the
615 FIO-agent cannot move for the current timestep.

616

617 3.7.2.3 Surface Routing Sub-model

618 The Surface Routing Sub-model (Figure 7) only runs for FIO-agents with the attributes *Life state* = “1”,
619 *Detached-faeces* = “1” and *Domain type* = “Land” or “Land_Channel” (Figure 4b). The sub-model
620 assumes full and immediate mixing of FIO-agents within hydrological stores and fluxes (c.f. Dorner et
621 al., 2006); consequently, as soon as an FIO-agent enters a new hydrological store (i.e. Pondered Surface
622 Water or Soil Water), it is immediately available for transport from that store in a hydrological flux
623 (e.g. overland flow, exfiltration). The validity of this assumption varies with cell size, being more valid
624 for smaller grid cells (Reaney, 2008).

625

626 For each timestep, the Surface Routing Sub-model proceeds as follows for each applicable FIO-agent
627 (Figure 7). If the attribute *Infiltration stage* of the FIO-agent is not equal to “Soil”, its infiltration into
628 the soil is determined based on a probability of infiltration:

629

$$630 \quad P(\text{Infiltrate}) = P_{\text{Infil}}[Att[xy]] \quad \text{Eq. 15}$$

631

632 If the FIO-agent infiltrates, its attribute *Infiltration stage* is set to “Soil”. Whether it exfiltrates back to
633 the surface then depends on a probability of exfiltration:

634

$$635 \quad P(\text{Exfiltrate}) = P_{\text{ExfilAtt}}[xy] \quad \text{Eq. 16}$$

636

637 If the *Infiltration stage* attribute of the FIO-agent was already “Soil” to begin with, only the potential
 638 for exfiltration is evaluated. If the FIO-agent exfiltrates, its *Infiltration stage* attribute is set to “Surface”.
 639 If after infiltration and exfiltration the FIO-agent has *Infiltration stage* equal to “Surface”, then it is
 640 assumed to be transported laterally in overland flow, and its attribute *Movement tracker* is set to “1”. If
 641 the FIO-agent is already in a cell containing a channel (cell with $\text{ChanAtt} = 1$), then its attribute *Domain*
 642 *type* is set to “Channel” to denote that overland flow has transported the FIO-agent from the land
 643 adjacent to the channel into the channel itself; the sub-model then exits and the Channel Routing Sub-
 644 model runs (Figure 4b). If not already in a cell with $\text{ChanAtt} = 1$, the FIO-agent moves to the downslope
 645 cell based on $f\text{DirAtt}$ of the cell it currently occupies. If the FIO-agent moves into a cell with $\text{ChanAtt} = 1$,
 646 what happens next depends on the presence of degraded soil (specified by degSoilAtt). If present, the
 647 *Domain type* and *Infiltration stage* attributes of the FIO-agent are changed to “Seepage” and “Soil”,
 648 respectively, and the sub-model exits so the Seepage Sub-model can run (Figure 4b). If not present,
 649 then *Domain type* is set to “Land_Channel” and the Surface Routing Sub-model runs again to determine
 650 whether the FIO-agent infiltrates into the soil next to the channel or is routed into the channel. If the
 651 latter, *Domain type* is set to “Channel” and the sub-model exits so the Channel Routing Sub-model can
 652 run; otherwise MAFIO moves on to simulate the next FIO-agent (Figure 4b). If the FIO-agent did not
 653 move into a cell with $\text{ChanAtt} = 1$, the Surface Routing Sub-model continues until the FIO-agent
 654 infiltrates into the soil but does not exfiltrate, reaches an area of degraded soil or enters the channel; an
 655 FIO-agent cannot, therefore, remain on the surface after the Surface Routing Sub-model has run.

656

657 3.7.2.4 Seepage Sub-model

658 The Seepage Sub-model (Figure 8) runs for FIO-agents with *Life state* and *Domain type* = “1” and
 659 “Seepage”, respectively (Figure 4b). It determines whether an FIO-agent in an area of degraded soil
 660 adjacent to the channel seeps to the stream based on a probability of seepage:

661

$$662 \quad P(\text{Seep}) = P_{\text{SeepAtt}}[xy] \quad \text{Eq. 17}$$

663

664 If the FIO-agent seeps to the stream, its attribute *Domain type* is set to “Channel” so the Channel
 665 Routing Sub-model can run (Figure 4b). Otherwise, the FIO-agent remains in the degraded soil for the
 666 current timestep. Whilst the Seepage Sub-model does not utilise any parameters, it is necessary to

667 specify the number of livestock units each animal of a given livestock type represents (*LUs*), the values
668 of livestock units per hectare that constitute the “low”, “medium” and “high” risk bands (*riskBands*),
669 the damage fraction associated with each risk band (*dFrac_Bands*) and the rate constant for the
670 exponential decline in damage fraction over time (k_d) used in the calculation of the spatial grid attribute
671 $P_{SeepAtt}$, (Section 3.2.4).

672

673 3.7.2.5 Channel Routing Sub-model

674 The Channel Routing Sub-model (Figure 9) runs for FIO-agents with *Domain type* = “Channel” (Figure
675 4b). First, the sub-model determines whether the FIO-agent attaches to stream sediment when entering
676 the channel based on the parameter *fracAttached*:

677

$$678 \quad P(Attached_{StreamSed}) = fracAttached \quad Eq. 18$$

679

680 If the FIO-agent does not attach, it is assumed it cannot settle to the streambed and the FIO-agent is
681 routed through the channel network to the catchment outlet. If the FIO-agent attaches, its attribute
682 *Attached-stream sediment* is set to “1”, and routing occurs as follows. The probability that the FIO-
683 agent settles to the streambed in its current cell is determined based on length of the channel in the cell
684 (*streamDist*) and the distance-decay model of Kay and McDonald (1980):

685

$$686 \quad P(Settle) = 1 - 10^{-\lambda_{sed} \cdot streamDist_{[xy]}} \quad Eq. 19$$

687

688 where λ_{sed} is the rate constant parameter in units of m^{-1} . If the FIO-agent does not settle, it is routed to
689 the next downstream cell, and the probability is calculated again. This continues until the FIO-agent
690 settles to the streambed or is routed out of the catchment. Once the FIO-agent reaches the cell
691 representing the catchment outlet, it may still settle to account for the length of the stream in that cell.
692 As with the Surface Routing Sub-model, the Channel Routing Sub-model assumes full and immediate
693 mixing of FIO-agents within each cell. In addition, it assumes that all FIO-agents passing through a cell
694 can be considered as a single population experiencing the same decay with distance, similar to the Die-
695 off Sub-model.

696

697 4 A hydrological environment generator: *EcH₂O-iso*

698 4.1 Overview of *EcH₂O-iso*

699 *EcH₂O-iso* is a process-based, spatially-distributed ecohydrological model that simulates coupled
700 energy, hydrological and vegetation dynamics (Kuppel et al., 2018b; Maneta and Silverman, 2013) with
701 the capacity to track the stable isotope (²H and ¹⁸O) compositions and water ages of simulated
702 hydrological stores and fluxes (Kuppel et al., 2018a). *EcH₂O-iso* has successfully been applied to
703 temperate (Kuppel et al., 2018a) and cold (Smith et al., 2019) regions, and at the plot- (Douinot et al.,
704 2019) to catchment-scale (Knighton et al., 2020; Kuppel et al., 2018a; Smith et al., 2019), demonstrating
705 the versatility of the model in different environments and process domains.

706

707 *EcH₂O-iso* has assumed full and immediate mixing for tracer and age tracking within each liquid
708 hydrological store to minimize parameterization and computing time (Kuppel et al., 2018a). However,
709 limitations to this approach have been highlighted, even in wet, energy-limited catchments where
710 heterogeneous mixing and flow is reduced compared to other ecoclimatic settings (Geris et al., 2015;
711 Kuppel et al., 2018a). More generally, there is a need to consider partial mixing in tracer-aided models
712 (Cain et al., 2019; Knighton et al., 2017) to accommodate possible ecohydrological separation of water
713 in the subsurface – that is its partitioning between water that is tightly-bound to the soil matrix in small
714 pores and mobile water which may predominantly contribute to groundwater recharge and streamflow
715 (Brooks et al., 2010; Goldsmith et al., 2012; Sprenger et al., 2018). To the authors' knowledge, two-
716 pore conceptualisations of the subsurface in physically-based models are only just emerging in
717 catchment studies (Hopp et al., 2020), with efforts largely having been limited to the plot scale (Jackish
718 and Zehe, 2018; Sprenger et al., 2018; Stumpp and Maloszewski, 2010; Vogel et al., 2010).

719

720 To incorporate heterogeneous subsurface mixing, *EcH₂O-iso* has been updated so that the tracked
721 isotopic composition and age of water considers a two-pore domain in the upper two layers of the three-
722 layer soil profile (Figure 10). The two domains (tightly-bound [TB] and mobile [MW]) are
723 distinguished using a tension threshold for mobile water (Ψ_{MW}), a parameter shared by both layers, with
724 a corresponding water content threshold (SWC_{MW}) determined by the Brooks-Corey pedo-transfer
725 functions used in *EcH₂O-iso* (Maneta and Silverman, 2013). In each layer, the MW domain only exists
726 if total water storage in the layer (SWC_t) exceeds the storage capacity of TB ($SWC_t > SWC_{MW}$). Liquid
727 water fluxes downwards (infiltration and percolation) and upwards (return flow) draw from MW and
728 primarily refill TB, except for surface water considered as MW (Figure 10). Secondary flow paths (MW
729 to MW) occur if the recipient TB reaches its maximum storage capacity. Soil evaporation and plant
730 transpiration draw from both domains in proportion to their respective water contents. After evaporative
731 losses, dispersion replenishes TB from MW (potentially emptying MW), assuming continuity in water
732 contents between the two domains.

733

734 The two-pore domain conceptualisation only affects tracer and age tracking; computation of water
735 fluxes between layers (infiltration/percolation, return flow) and from the subsurface system (soil
736 evaporation, transpiration) is unchanged from previous versions of EcH₂O-iso. The groundwater store
737 is also still defined using a field capacity tension threshold (Ψ_{FC} , fixed to 3.36 m), and tracking assumes
738 full mixing within the 3rd soil layer even if unsaturated (Figure 10). The remaining workings of EcH₂O-
739 iso are as described in Maneta and Silverman (2013) and Kuppel et al. (2018a, b), with the further
740 addition of species-dependent root profiles as implemented in Douinot et al. (2019) and Knighton et al.
741 (2020).

742

743 *4.2 Interfacing EcH₂O-iso with MAFIO*

744 EcH₂O-iso provides all outputs required to characterise the hydrological environment of MAFIO.
745 However, EcH₂O-iso simulates infiltration twice within a grid cell, first following the addition of
746 effective precipitation to ponded surface water and then again after the addition of snowmelt and run-
747 on from upslope cells (see Figure 1 in Kuppel et al., 2018a). For processes in MAFIO to be consistent
748 with this, two probabilities of infiltration need to be assigned to each cell of the spatial grid ($P_{Infil1Att}$
749 and $P_{Infil2Att}$). Then, FIO-agents which have been detached by rainfall on a given timestep and are
750 still in their initial cell may first infiltrate with probability $P_{Infil1Att}$ and then, if still on the surface,
751 with probability $P_{Infil2Att}$, whilst FIO-agents which move into a cell in run-on from upslope can only
752 infiltrate with probability $P_{Infil2Att}$. This aside, the process conceptualisation of EcH₂O-iso is
753 compatible with the catchment conceptualisation represented by the variables of the hydrological
754 environment of MAFIO (Section 3.2.2). As EcH₂O-iso conceptualises three hydrological layers in the
755 soil, $Exfil$, SWC_{Exf} and Sat_{Def} are taken from the upper layer to be consistent with the representation of
756 the upper soil profile in MAFIO.

757

758 **5 Conclusions**

759 This paper has reported the rationale for and development of MAFIO, a new agent-based model for
760 simulating the behaviour and transport of agents representing FIOs. The model is intended to elucidate
761 the sources and transfer mechanisms contributing FIOs to streams at the sub-field scale in small
762 agricultural catchments, with improved confidence in hydrological process representation afforded by
763 the use of a robust external model for simulating the hydrological environment. Here, the spatially-
764 distributed, tracer-aided ecohydrological model EcH₂O-iso was introduced as an exemplar hydrological
765 environment generator. A companion paper (Neill et al., *in review*) provides a proof-of-concept

766 application of MAFIO focusing on process representation and the potential the model has for use in a
767 management context.

768

769 **Software availability**

770 The source code for MAFIO as outlined in this work is available via the University of Aberdeen PURE
771 repository: <https://doi.org/10.20392/66f74663-ece3-4a52-8bed-f0cf52d0831a>

772 The source code for EcH₂O-iso is available at: https://bitbucket.org/sylka/ech2o_iso/src/master_2.0/

773

774 **Acknowledgements**

775 Funding for this work from the Scottish Government's Hydro Nation Scholars Programme is gratefully
776 acknowledged. Many thanks go to Sim Reaney and John Wainwright for initial discussions regarding
777 environmental agent-based modelling approaches and methods for their implementation.

778

779 **References**

- 780 Abdou, M., Hamill, L., Gilbert, N., 2012. Designing and Building an Agent-Based Model: 141-166 in Heppenstall,
781 A.J., Crooks, A.T., See, L.M., Batty, M. (eds). Agent-Based Models of Geographical Systems. Springer:
782 Dordrecht, Heidelberg, London, New York.
- 783 Ala-aho, P., Tetzlaff, D., McNamara, J.P., Laudon, H., Soulsby, C., 2017. Using isotopes to constrain water flux
784 and age estimates in snow-influenced catchments using the STARR (Spatially distributed Tracer-Aided
785 Rainfall-Runoff) model. Hydrological Earth Systems Science 21: 5089-5110.
- 786 Avery, S.M., Moore, A., Hutchison, M.L., 2004. Fate of *Escherichia coli* originating from livestock faeces
787 deposited directly onto pasture. Letters in Applied Microbiology 38: 355-359.
- 788 Benhamou, S., 2006. Detecting an orientation component in animal paths when the preferred direction is
789 individual-dependent. Ecology 87: 518-528.
- 790 Benhamou, S., 2013. Of scales and stationarity in animal movements. Ecology Letters 17: 261-272.
- 791 Berman, E.S.F., Levin, N.E., Landais, A., Li, S., Owano, T., 2013. Measurement of $\delta^{18}\text{O}$, $\delta^{17}\text{O}$, and ^{17}O -excess in
792 Water by Off-Axis Integrated Cavity Output Spectroscopy and Isotope Ratio Mass Spectrometry.
793 Analytical Chemistry 85: 10392-10398.
- 794 Bicknell, B.R., Imhoff, J.C., Kittle Jr., J.L., Donigan Jr., A.S., Johanson, R.C., 1997. Hydrological simulation
795 program – FORTRAN user's manual for version 11. Environmental Protection Agency Report No:
796 EPA/600/R-97/080. U.S. Environmental Protection Agency: Athens, Ga.
- 797 Bilotta, G.S., Brazier, R.E., Haygarth, P.M., 2007. The impacts of grazing animals on the quality of soils,
798 vegetation and surface waters in intensively managed grasslands. Advances in Agronomy 94: 237-280.
- 799 Birkel, C., Soulsby, C., 2015. Advancing tracer-aided rainfall-runoff modelling: a review of progress, problems
800 and unrealised potential. Hydrological Processes 29: 5227-5240.
- 801 Birkel, C., Soulsby, C., Tetzlaff, D., 2014a. Developing a consistent process-based conceptualization of catchment
802 functioning using measurements of internal state variables. Water Resources Research 50: 3481-3501.
- 803 Birkel, C., Soulsby, C., Tetzlaff, D., 2014b. Integrating parsimonious models of hydrological connectivity and
804 soil biogeochemistry to simulate stream DOC dynamics. Journal of Geophysical Research 119: 1030-
805 1047.
- 806 Blaustein, R.A., Hill, R.L., Micallef, S.A., Shelton, D.R., Pachepsky, Y.A., 2016. Rainfall intensity effects on
807 removal of fecal indicator bacteria from solid dairy manure applied over grass-covered soil. Science of
808 the Total Environment 539: 583-591.
- 809 Blaustein, R.A., Pachepsky, Y., Hill, R.L., Shelton, D.R., 2015a. Solid manure as a source of fecal indicator
810 microorganisms: Release under simulated rainfall. Environmental Science and Technology 49: 7860-
811 7869.

812 Blaustein, R.A., Pachepsky, Y., Shelton, D.R., Hill, R.L., 2015b. Release and removal of microorganisms from
813 land-deposited animal waste and animal manures: A review of data and models. *Journal of Environmental*
814 *Quality* 44: 1338-1354.

815 Blaustein, R.A., Pachepsky, Y., Hill, R.L., Shelton, D.R., Whelan, G., 2013. *Escherichia coli* survival in waters:
816 Temperature dependence. *Water Research* 47(2): 569-578

817 Brooks, J.R., Barnard, H.R., Coulombe, R., McDonnell, J.J., 2010. Ecohydrologic separation of water between
818 trees and streams in a Mediterranean climate. *Nature Geoscience* 3: 100-104.

819 Brooks, P.D., Chorover, J., Fan, Y., Godsey, S.E., Maxwell, R.M., McNamara, J.P., Tague, C., 2015. Hydrological
820 partitioning in the critical zone: Recent advances and opportunities for developing transferable
821 understanding of water cycle dynamics. *Water Resources Research* 51: 6973-6987.

822 Cain, M.R., Ward, A.S., Hrachowitz, M., 2019. Ecohydrologic separation alters interpreted hydrologic stores and
823 fluxes in a headwater mountain catchment. *Hydrological Processes* 33: 2658-2675.

824 Chadwick D., Fish, R., Oliver, D.M., Heathwaite, L., Hodgson, C., Winter, M., 2008. Management of livestock
825 and their manure to reduce the risk of microbial transfers to water – the case for an interdisciplinary
826 approach. *Trends in Food Science and Technology* 19: 240-247.

827 Chapra, S.C., 1997. *Surface water-quality modelling*. McGraw-Hill: New York.

828 Chick, H., 1908. Investigation of the laws of disinfection. *Journal of Hygiene* 8: 655.

829 Cho, K.H., Pachepsky, Y.A., Oliver, D.M., Muirhead, R.W., Park, Y., Quilliam, R.S., Shelton, D.R., 2016.
830 Modeling fate and transport of fecally-derived microorganisms at the watershed scale: State of the
831 science and future opportunities. *Water Research* 100: 38-56.

832 Cirbus, J., Podhoranyi, M., 2013. Cellular automata for the flow simulations on the Earth surface, optimisation
833 computation process. *Applied Mathematics and Information Sciences* 7: 2149-2158.

834 Clark, M.P., Kavetski, D., Fenicia, F., 2011. Pursuing the method of multiple working hypotheses for hydrological
835 modeling. *Water Resources Research* 47: W09301.

836 Collins, R., Rutherford, K., 2004. Modelling bacterial water quality in streams draining pastoral land. *Water*
837 *Research* 38: 700-712.

838 Cooper, J.R., Wainwright, J., Parsons, A.J., Onda, Y., Fukuwara, T., Obana, E., Kitchener, B., Long, E.J.,
839 Hargrave, G.H., 2012. A new approach for simulating the redistribution of soil particles by water erosion:
840 A marker-in-cell model. *Journal of Geophysical Research* 117: F04027.

841 Crooks, A.T., Heppenstall, A.J., Introduction to Agent-Based Modelling: 85-108 in Heppenstall, A.J., Crooks,
842 A.T., See, L.M., Batty, M. (eds). *Agent-Based Models of Geographical Systems*. Springer: Dordrecht,
843 Heidelberg, London, New York.

844 Davies-Colley, R.J., Nagels, J.W., Smith, R.A., Young, R.G., Phillips, C.J., 2004. Water quality impact of a dairy
845 cowherd crossing a stream. *New Zealand Journal of Marine and Freshwater Research* 38(4): 569–576.

846 de Brauwere, A., Ouattara, N.K., Servais, P., 2014. Modeling fecal indicator bacteria concentrations in natural
847 surface waters: A review. *Critical Reviews in Environmental Science and Technology* 44: 2380-2453.

848 Dick, J.J., Tetzlaff, D., Birkel, C., Soulsby, C., 2015. Modelling landscape controls on dissolved organic carbon
849 sources and fluxes to streams. *Biogeochemistry* 122: 361-374.

850 Dorner, S.M., Anderson, W.B., Slawson, R.M., Kouwen, N., Huck, P.M., 2006. Hydrologic modeling of pathogen
851 fate and transport. *Environmental Science and Technology* 40: 4746-4753.

852 Drewry, J.J., 2006. Natural recovery of soil physical properties from treading damage of pastoral soils in New
853 Zealand and Australia: A review. *Agriculture, Ecosystems and Environment* 144: 159-169.

854 Douinot, A., Tetzlaff, D., Maneta, M., Kuppel, S., Schulte-Bisping, H., Soulsby, C., 2019. Ecohydrological
855 modelling with ECH2O-iso to quantify forest and grassland effects of water partitioning and flux ages.
856 *Hydrological Processes* 33: 2174-2191.

857 Elliot, A.H., Tian, Y.Q., Rutherford, J.C., Carlson, W.T., 2002. Effect of cattle treading on interrill erosion from
858 hill pasture: modelling concepts and analysis of rainfall simulator data. *Australian Journal of Soil
859 Research* 40: 963-976.

860 Falkenmark, M., Rockström, J., 2006. The new blue and green water paradigm: Breaking new ground for water
861 resources planning and management. *Journal of Water Resources Planning and Management* 132: 129-
862 132.

863 Fatichi, S., Vivoni, E.R., Ogden, F.L., Ivanov, V.Y., Mirus, B., Gochis, D., Downer, C.W., Camporese, M.,
864 Davison, J.H., Ebel, B., Jones, N., Kim, J., Mascaro, G., Niswonger, R., Restrepo, P., Rigon, R., Shen,
865 C., Sulis, M., Tarboton, D., 2016. An overview of current applications, challenges, and future trends in
866 distributed process-based models in hydrology. *Journal of Hydrology* 537: 45-60.

867 Fewtrell, L., Kay, D. 2015. Recreational water and infection: a review of recent findings. *Current Environmental
868 Health Reports* 2(1): 85-94.

869 Geldreich, E.E., 1996. Pathogenic agents in freshwater resources. *Hydrological Processes* 10: 315-333.

870 Geris, J., Tetzlaff, D., McDonnell, J., Anderson, J., Paton, G., and Soulsby, C., 2015. Ecohydrological separation
871 in wet, low energy northern environments? A preliminary assessment using different soil water extraction
872 techniques. *Hydrological Processes* 29: 5139-5152.

873 Goldsmith, G.R., Munoz-Villers, L.E., Holwerda, F., McDonnell, J.J., Asbjornsen, H., Dawson, T.E., 2012. Stable
874 isotopes reveal linkages among ecohydrological processes in a seasonally dry tropical montane cloud
875 forest. *Ecohydrology* 5: 779-790.

876 Greene, S., Johnes, P.J., Bloomfield, J.P., Reaney, S.M., Lawley, R., Elkhatib, Y., Freer, J., Odoni, N., Macleod,
877 C.J.A., Percy, B., 2015. A geospatial framework to support integrated biogeochemical modelling in the
878 United Kingdom. *Environmental Modelling and Software* 68: 219-232.

879 Grimm, V., Berger, U., DeAngelis, D.L., Polhill, J.G., Giske, J., Railsback, S.F., 2010. The ODD protocol: A
880 review and first update. *Ecological Modelling* 221: 2760-2768.

881 Grimm, V., Berger, U., Bastiansen, F., Eliassen, S., Ginot, V., Giske, J., Goss-Custard, J., Grand, T., Heinz, S.K.,
882 Huse, G., Huth, A., Jepsen, J.U., Jørgensen, C., Mooij, W.M., Müller, B., Pe'er, G., Piou, C., Railsback,
883 S.F., Robbins, A.M., Robbins, M.M., Rossmanith, E., Rürger, N., Strand, E., Souissi, S., Stillman, R.A.,
884 Vabø, R., Visser, U., DeAngelis, D.L., 2006. A standard protocol for describing individual and agent-
885 based models. *Ecological Modelling* 198: 115-126.

886 Haydon, S., Deletic, A., 2006. Development of a coupled pathogen-hydrologic catchment model. *Journal of*
887 *Hydrology* 328: 467-480.

888 Himathongkham, S., Bahari, S., Riemann, H., Cliver, D., 1999. Survival of *Escherichia coli* O157:H7 and
889 *Salmonella typhimurium* in cow manure and cow manure slurry. *FEMS Microbiology Letters* 178: 251-
890 257.

891 Hipsey, M.R., Antenucci, J.P., Brookes, J.D., 2008. A generic, process-based model of microbial pollution in
892 aquatic systems. *Water Resources Research* 44: W07408.

893 Hodge, R.A., Hoey, T.B., 2012. Upscaling from grain-scale processes to alluviation in bedrock channels using a
894 cellular automaton model. *Journal of Geophysical Research* 117: F01017.

895 Hodgson, C.J., Bulmer, N., Chadwick, D.R., Oliver, D.M., Heathwaite, A.L., Fish, R.D., Winter, M., 2009.
896 Establishing relative release kinetics of faecal indicator organisms from different faecal matrices. *Letters*
897 *in Applied Microbiology* 49: 124-130.

898 Hopp, L., Glaser, B., Klaus, J., & Schramm, T., 2020. The relevance of preferential flow in catchment scale
899 simulations: Calibrating a 3D dual-permeability model using DREAM. *Hydrological Processes* 34(5):
900 1237-1254.

901 Jackisch, C., Zehe, E., 2018. Ecohydrological particle model based on representative domains. *Hydrology and*
902 *Earth Systems Science* 22: 3639-3662.

903 Jamieson, R., Joy, D.M., Lee, H., Kostaschuk, R., Gordon, R., 2005. Transport and deposition of sediment-
904 associated *Escherichia coli* in natural streams. *Water Research* 39: 2665-2675.

905 Karr, J.R., Schlosser, I.J., 1978. Water resources and the land-surface interface. *Science* 201: 229-234.

906 Karssenber, D., Schmitz, O., Salamon, P., de Jong, K., Bierkens, M.F.P., 2010. A software framework for
907 construction of process-based stochastic spatio-temporal models and data assimilation. *Environmental*
908 *Modelling and Software* 25: 489-502.

909 Kay, D., McDonald, A., 1980. Reduction of coliform bacteria in two upland reservoirs: The significance of
910 distance decay relationships. *Water Research* 14: 305-318.

911 Kay, D., Crowther, J., Stapleton, C.M., Wyer, M.D., Fewtrell, L., Anthony, S., Bradford, M., Edwards, A., Francis,
912 C.A., Hopkins, M., Kay, C., McDonald, A.T., Watkins, J., Wilkinson, J., 2008. Faecal indicator organism
913 concentrations and catchment export coefficients in the UK. *Water Research* 42: 2649-2661.

914 Kay, D., Aitken, M., Crowther, J., Dickson, I., Edwards, A.C., Francis, C., Hopkins, M., Jeffrey, W., Kay, C.,
915 McDonald, A.T., McDonald, D., Stapleton, C.M., Watkins, J., Wilkinson, J., Wyer, M.D., 2007.
916 Reducing fluxes of faecal indicator compliance parameters to bathing waters from diffuse agricultural
917 sources: The Brighthouse Bay study, Scotland. *Environmental Pollution* 147: 138-149.

918 Kim, J., Pachepsky, Y.A., Shelton, D.R., Coppock, C., 2010. Effect of streambed bacteria release on *E. coli*
919 concentrations: Monitoring and modeling with the modified SWAT. *Ecological Modelling* 221: 1592-
920 1604.

921 Knighton, J., Saia, S.M., Morris, C.K., Archibald, J.A., Todd Walter, M., 2017. Ecohydrological considerations
922 for modeling of stable water isotopes in a small intermittent watershed. *Hydrological Processes* 31: 2438-
923 2452.

924 Knighton, J., Kuppel, S., Smith, A., Spenger, M., Soulsby, C., Tetzlaff, D., 2020. Using isotopes to incorporate
925 tree water storage and mixing dynamics into a distributed hydrologic modeling framework.
926 *Ecohydrology* 13: e2201.

927 Kuppel, S., Tetzlaff, D., Maneta, M.P., Soulsby, C., 2018a. EcH2O-iso 1.0: water isotopes and age tracking in a
928 process-based, distributed ecohydrological model. *Geoscientific Model Development* 11: 3045-3069.

929 Kuppel, S., Tetzlaff, D., Maneta, M.P., Soulsby, C., 2018b. What can we learn from multi-criteria calibration of
930 a process-based ecohydrological model? *Environmental Modelling and Software* 101: 301-316.

931 Macal, C.M., North, M.J., 2010. Tutorial on agent-based modelling and simulation. *Journal of Simulation* 4(3):
932 151-162.

933 Mancini, J.L., 1978. Numerical estimates of coliform mortality rates under various conditions. *Journal of Water*
934 *Pollution Control Federation* 50: 2477-2484.

935 Maneta, M.P., Silverman, N.L., 2013 A spatially distributed model to simulate water, energy, and vegetation
936 dynamics using information from regional climate models. *Earth Interactions* 17: 1-44.

937 McDonnell, J.J., Beven, K., 2014. Debates – The future of hydrological sciences: A (common) path forward? A
938 call to action aimed at understanding velocities, celerities and residence time distributions of the
939 headwater hydrograph. *Water Resources Research* 50: 5342-5350.

940 McKergow, L.A., Davies-Colley, R.J., 2010. Stormflow dynamics and loads of *Escherichia coli* in a large mixed
941 land use catchment. *Hydrological Processes* 24: 276-289.

942 Moriarty, E.M., Mackenzie, M.L., Karki, N., Sinton, L.W., 2011. Survival of *Escherichia coli*, enterococci and
943 *Campylobacter* spp. in sheep feces on pastures. *Applied and Environmental Microbiology* 77: 1797-
944 1803.

945 Muirhead, R., 2009. Soil and faecal material reservoirs of *Escherichia coli* in a grazed pasture. New Zealand
946 Journal of Agricultural Research 52: 1-8.

947 Nagels, J.W., Davies-Colley, R.J., Donnison, A.M., Muirhead, R.W., 2002. Faecal contamination over flood
948 events in a pastoral agricultural stream in New Zealand. Water Science and Technology 45: 45-52.

949 Natural England, 2013. Higher level stewardship: Environmental stewardship handbook (Fourth Edition).
950 Available at: <http://publications.naturalengland.org.uk/file/2819648>

951 Neill, A.J., Tetzlaff, D., Strachan, N.J.C., Soulsby, C., 2019. To what extent does hydrological connectivity control
952 dynamics of faecal indicator organisms in streams? Initial hypothesis testing using a tracer-aided model.
953 Journal of Hydrology 570: 423-435.

954 Neill, A.J., Tetzlaff, D., Strachan, N.J.C., Hough, R.L., Avery, L.M., Maneta, M.P., Soulsby, C., *in review*. An
955 agent-based model that simulates the spatio-temporal dynamics of sources and transfer mechanisms
956 contributing faecal indicator organisms to streams. Part 2: Application to a small agricultural catchment.
957 Journal of Environmental Management.

958 O’Callaghan, J.F., Mark, D.A., 1984. The extraction of drainage networks from digital elevation data. Computer
959 Vision, Graphics and Image Processing 28: 323-344.

960 O’Sullivan, D., Millington, J., Perry, G. & Wainwright, J., 2012. Agent-based models – because they’re worth it?:
961 109-123 in Heppenstall, A.J., Crooks, A.T., See, L.M., Batty, M. (eds). Agent-Based Models of
962 Geographical Systems. Springer: Dordrecht, Heidelberg, London, New York.

963 Oliver D.M., Clegg C.D., Haygarth P.M., Heathwaite A.L., 2005a. Assessing the potential for pathogen transfer
964 from grassland soils to surface waters. Advances in Agronomy 85: 125–180.

965 Oliver, D.M., Heathwaite, L., Haygarth, P.M., Clegg, C.D., 2005b. Transfer of *Escherichia coli* to Water from
966 Drained and Undrained Grassland after Grazing. Journal of Environmental Quality 34: 918-925.

967 Oliver, D.M., Heathwaite, A.L., Hodgson, C.J., Chadwick, D.R., 2007. Mitigation and current management
968 attempts to limit pathogen survival and movement within farmed grassland. Advances in Agronomy 93:
969 95-152.

970 Oliver, D.M., Page, T., Heathwaite, A.L., Haygarth, P.M., 2010. Re-shaping models of *E. coli* population
971 dynamics in livestock faeces: Increased bacterial risk to humans? Environmental International 36: 1-7.

972 Oliver, D.M., Bartie, P.J., Heathwaite, A.L., Reaney, S.M., Parnell, J.A.P., Quilliam, R.S., 2018. A catchment-
973 scale model to predict spatial and temporal burden of *E. coli* on pasture from grazing livestock. Science
974 of the Total Environment 616-617: 678-687.

975 Oliver, D.M., Fish, R.D., Hodgson, C.J., Heathwaite, A.L., Chadwick, D.R., Winter, M., 2009. A cross-
976 disciplinary toolkit to assess the risk of faecal indicator loss from grassland farm systems to surface
977 waters. Agriculture, Ecosystems and Environment 129: 401-412.

978 Oliver, D.M., Porter, K.D.H., Pachepsky, Y.A., Muirhead, R.W., Reaney, S.M., Coffey, R., Kay, D., Milledge,
979 D.G., Hong, E., Anthony, S.G., Page, T., Bloodworth, J.W., Mellander, P., Carbonneau, P., McGrane,
980 S.J., Quilliam, R.S., 2016. Predicting microbial water quality with models: Over-arching questions for
981 managing risk in agricultural catchments. *Science of the Total Environment* 544: 39-47.

982 Pachepsky, Y.A., Shelton, D.R., 2011. *Escherichia coli* and fecal coliforms in freshwater and estuarine sediments.
983 *Critical Reviews in Environmental Science and Technology* 41(12): 1067-1110.

984 Pandey, P.K., Soupir, M.L., Rehmann, C.R., 2012. A model for predicting resuspension of *Escherichia coli* from
985 streambed sediments. *Water Research* 46: 115-126.

986 Parry, H.R., Bithell, M., 2012. Large scale agent-based modelling: A review and guidelines for model scaling:
987 271-308 in Heppenstall, A.J., Crooks, A.T., See, L.M., Batty, M. (eds). *Agent-Based Models of*
988 *Geographical Systems*. Springer: Dordrecht, Heidelberg, London, New York.

989 Piovano, T.I., Tetzlaff, D., Ala-aho, P., Buttle, J., Mitchell, C.P.J., Soulsby, C., 2018. Testing a spatially
990 distributed tracer-aided runoff model in a snow-influenced catchment: Effects of multicriteria calibration
991 on streamwater ages. *Hydrological Processes* 30: 3089-3107.

992 Piovano, T.I., Tetzlaff, D., Carey, S.K., Shatilla, N.J., Smith, A., Soulsby, C., 2019. Spatially distributed tracer-
993 aided runoff modelling dynamics of storage and water ages in a permafrost-influenced catchment.
994 *Hydrology and Earth Systems Science* 23: 2507-2523.

995 Ravazzani, G., Rametta, D., Mancini, M., 2011. Macroscopic cellular automata for groundwater modelling: A
996 first approach. *Environmental Modelling and Software* 26: 634-643.

997 Reaney, S.M., 2008. The use of agent based modelling techniques in hydrology: determining the spatial and
998 temporal origin of channel flow in semi-arid catchments. *Earth Surface Processes and Landforms* 33:
999 317-327.

1000 Remondi, F., Kirchner, J.W., Burlando, P., Fatichi, S., 2018. Water flux tracking with a distributed hydrological
1001 model to quantify controls on spatio-temporal variability of transit time distributions. *Water Resources*
1002 *Research* 54: 3081-3099.

1003 Rode, M., Arhonditsis, G., Balin, D., Kebede, T., Krysanova, V., van Griensven, A., van der Zee, S.E.A.T.M.,
1004 2010. New challenges in integrated water quality modelling. *Hydrological Processes* 24: 3447-3461.

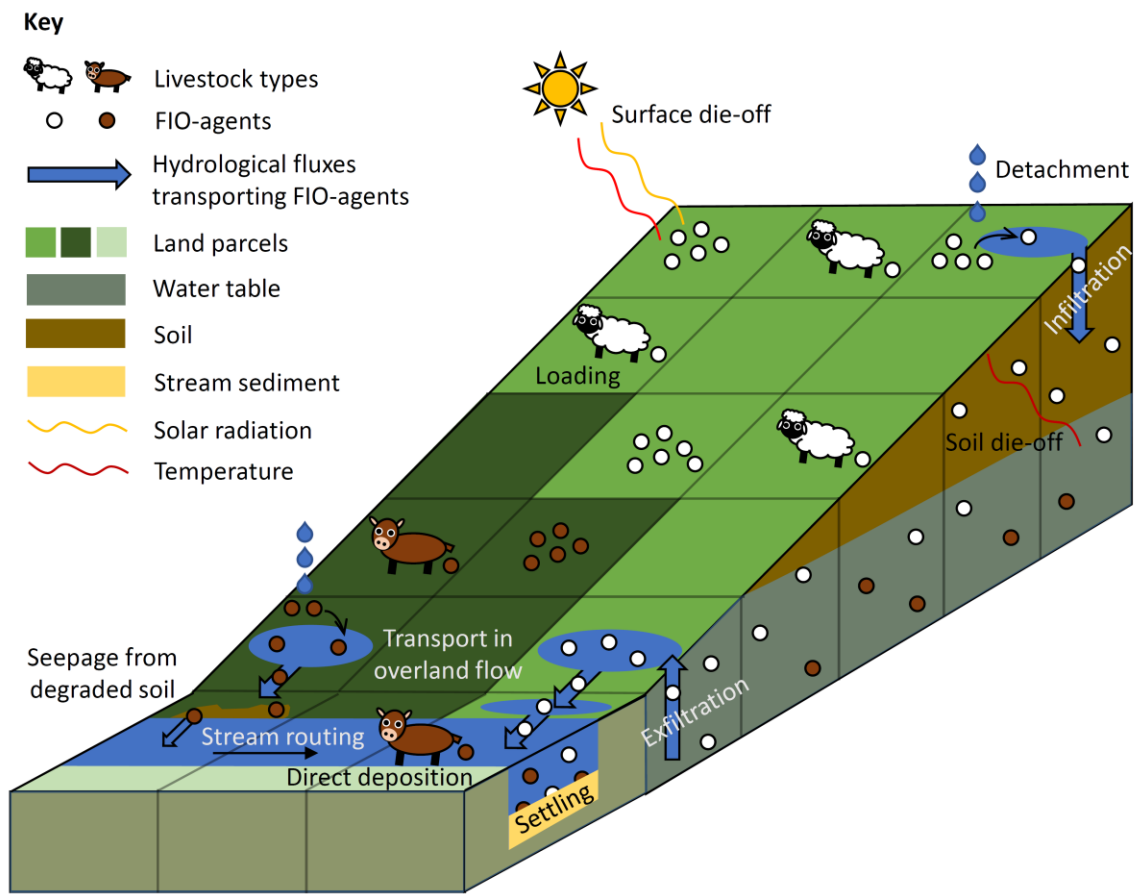
1005 Sadeghi, A., Arnold, J., 2002. A SWAT/Microbial Sub-Model for Predicting Pathogen Loadings in Surface and
1006 Groundwater at Watershed and Basin Scales. In *Total Maximum Daily Load (TMDL): Environmental*
1007 *Regulations, Proceedings of 2002 Conference* (p. 56). American Society of Agricultural and Biological
1008 Engineers.

1009 Schijven, J., Derx, J., de Roda Husman, A.M., Blaschke, A.P., Farnleitner, A.H., 2015. QMRACatch: Microbial
1010 Quality Simulation of Water Resources including Infection Risk Assessment. *Journal of Environmental*
1011 *Quality* 44: 1491-1502.

- 1012 Shao, Q., Weatherly, D., Huang, L., Baumgartl, T., 2015. RunCA: A cellular automata model for simulating
1013 surface runoff at different scales. *Journal of Hydrology* 529: 816-829.
- 1014 Sheath, G.W., Carlson, W.T., 1998. Impact of cattle treading on hill land: 1. Soil damage patterns and pasture
1015 status. *New Zealand Journal of Agricultural Research* 41(2): 271-278.
- 1016 Smith, A.A., Tetzlaff, D., Laudon, H., Maneta, M., and Soulsby, C., 2019. Assessing the influence of soil freeze-
1017 thaw cycles on catchment water storage-flux-age interactions using a tracer-aided ecohydrological
1018 model. *Hydrology and Earth Systems Science* 23: 3319-3334.
- 1019 Sokolova, E., Lindström, G., Pers, C., Strömqvist, J., Lewerin, S.S., Wahlström, H., Sörén, K., 2018. Water quality
1020 modelling: microbial risks associated with manure on pasture and arable land. *Journal of Water and*
1021 *Health* 16(4): 549-561.
- 1022 Soupir, M.L., Mostaghimi, S., Lou, J., 2008. Die-off of *E. coli* and Enterococci in dairy cowpats. *Transactions of*
1023 *the ASABE* 51(6): 1987-1996.
- 1024 Sprenger, M., Tetzlaff, D., Buttle, J., Laudon, H., Leistert, H., Mitchell, C.P.J., Snelgrove, J., Weiler, M., Soulsby,
1025 C., 2018. Measuring and modeling stable isotopes of mobile and bulk soil water. *Vadose Zone Journal*
1026 17: 170149.
- 1027 Stocker, M.D., Pachepsky, Y.A., Hill, R.L., Shelton, D.R., 2015. Depth-dependent survival of *Escherichia coli*
1028 and Enterococci in soil after manure application and simulated rainfall. *Applied and Environmental*
1029 *Microbiology* 81: 4801-4808.
- 1030 Stumpp, C., Maloszewski, P., 2010. Quantification of preferential flow and flow heterogeneities in an unsaturated
1031 soil planted with different crops using the environmental isotope $\delta^{18}\text{O}$. *Journal of Hydrology* 394: 407-
1032 415.
- 1033 Tetzlaff, D., Capell, R., and Soulsby, C., 2012. Land use and hydroclimatic influences on Faecal Indicator
1034 Organisms in two large Scottish catchments: Towards land use-based models as screening tools. *Science*
1035 *of the Total Environment* 434: 110-122.
- 1036 Thomann, R.V., Mueller, J.A., 1987. Principles of surface water quality modeling and control. Harper and Row:
1037 New York.
- 1038 Vaché, K.B., McDonnell, J.J., 2006. A process-based rejectionist framework for evaluating catchment runoff
1039 model structure. *Water Resources Research* 42: W02409.
- 1040 van Huijgevoort, M.H.J., Tetzlaff, D., Sutanudjaja, E.H., Soulsby, C., 2016. Using high resolution tracer data to
1041 constrain water storage, flux and age estimates in a spatially distributed rainfall-runoff model.
1042 *Hydrological Processes* 30: 4761-4778.
- 1043 Vinten, A., Sample, J., Ibiyemi, A., Abdul-Salam, Y., Stutter, M., 2017. A tool for cost-effectiveness analysis of
1044 field scale sediment-bound phosphorus mitigation measures and application to analysis of spatial and
1045 temporal targeting in the Lunan Water catchment, Scotland. *Science of the Total Environment* 586: 631-
1046 641.

- 1047 Vinten, A.J.A., Sym, G., Avdic, K., Crawford, C., Duncan, A., Merrilees, D.W., 2008. Faecal indicator pollution
1048 from a dairy farm in Ayrshire, Scotland: Source apportionment, risk assessment and potential of
1049 mitigation measures. *Water Research* 42: 997-1012.
- 1050 Vivoni, E. R., 2012. Diagnosing seasonal vegetation impacts on evapotranspiration and its partitioning at the
1051 catchment scale during SMEX04–NAME. *Journal of Hydrometeorology* 13(5): 1631–1638.
- 1052 Vogel, H.-J., Weller, U., Ippisch, O., 2010. Non-equilibrium in soil hydraulic modelling. *Journal of Hydrology*
1053 393: 20-28.
- 1054 Welch, D. 1982. Dung properties and defecation characteristics in some Scottish herbivores, with and evaluation
1055 of the dung-volume method of assess occupance. *Acta Theriologica* 27: 191-212.
- 1056 Wellen, C., Kamran-Disfani, A.R., Arhonditsis, G.B., 2015. Evaluation of the current state of distributed
1057 watershed nutrient water quality modeling. *Environmental Science and Technology* 49: 3278-3290.
- 1058 Whitehead, P.G., Leckie, H., Rankinen, K., Butterfield, D., Futter, M.N., Bussi, G., 2016. An INCA model for
1059 pathogens in rivers and catchments: Model structure, sensitivity analysis and application to the River
1060 Thames catchment, UK. *Science of the Total Environment* 572: 1601-1610.
- 1061 Zhang, W., Montgomery, D.R., 1994. Digital elevation model grid size, landscape representation, and hydrologic
1062 simulations. *Water Resources Research* 30: 1019-1028.
- 1063

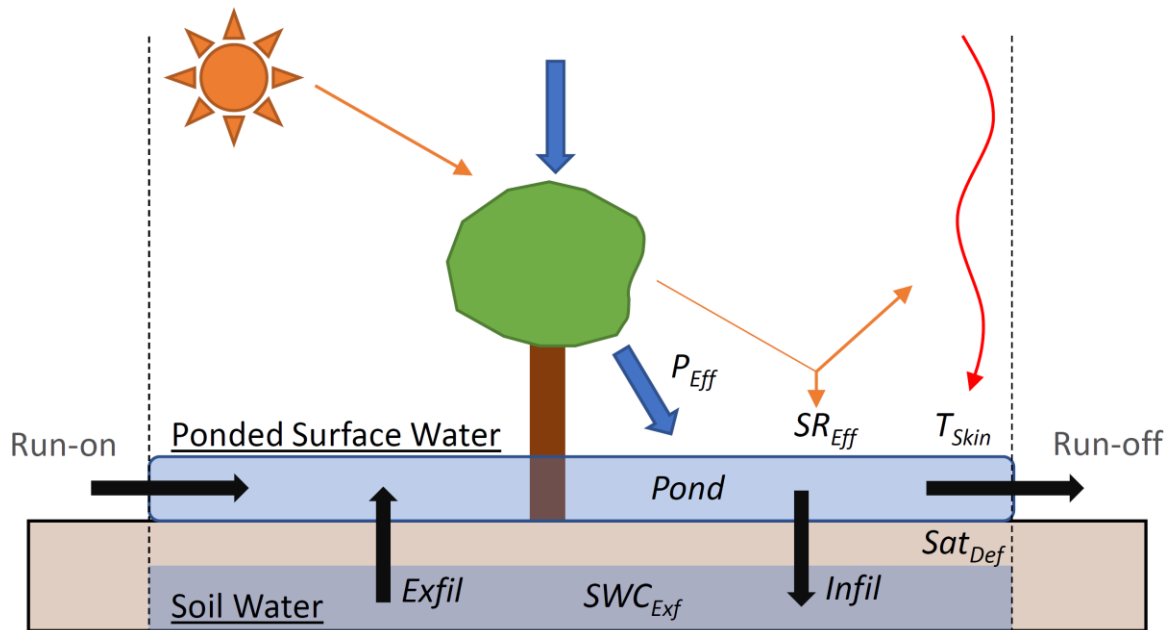
1064 **Figures**



1065

1066 *Figure 1: A conceptual diagram of MAFIO showing the faecal indicator organism (FIO) behaviour and*
 1067 *transport processes simulated by the model. Simulations take place on a regular spatial grid.*

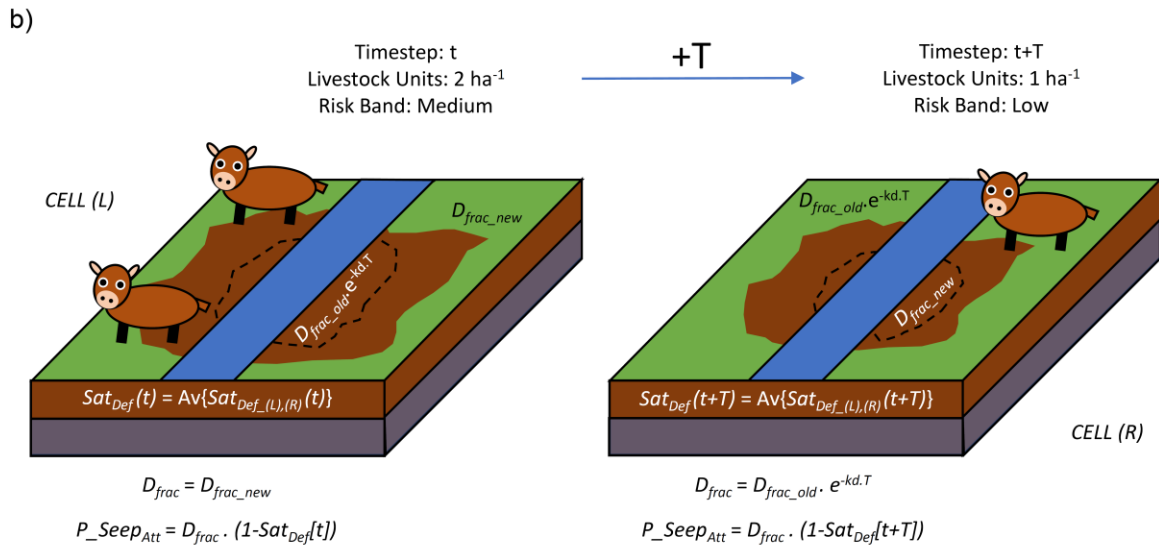
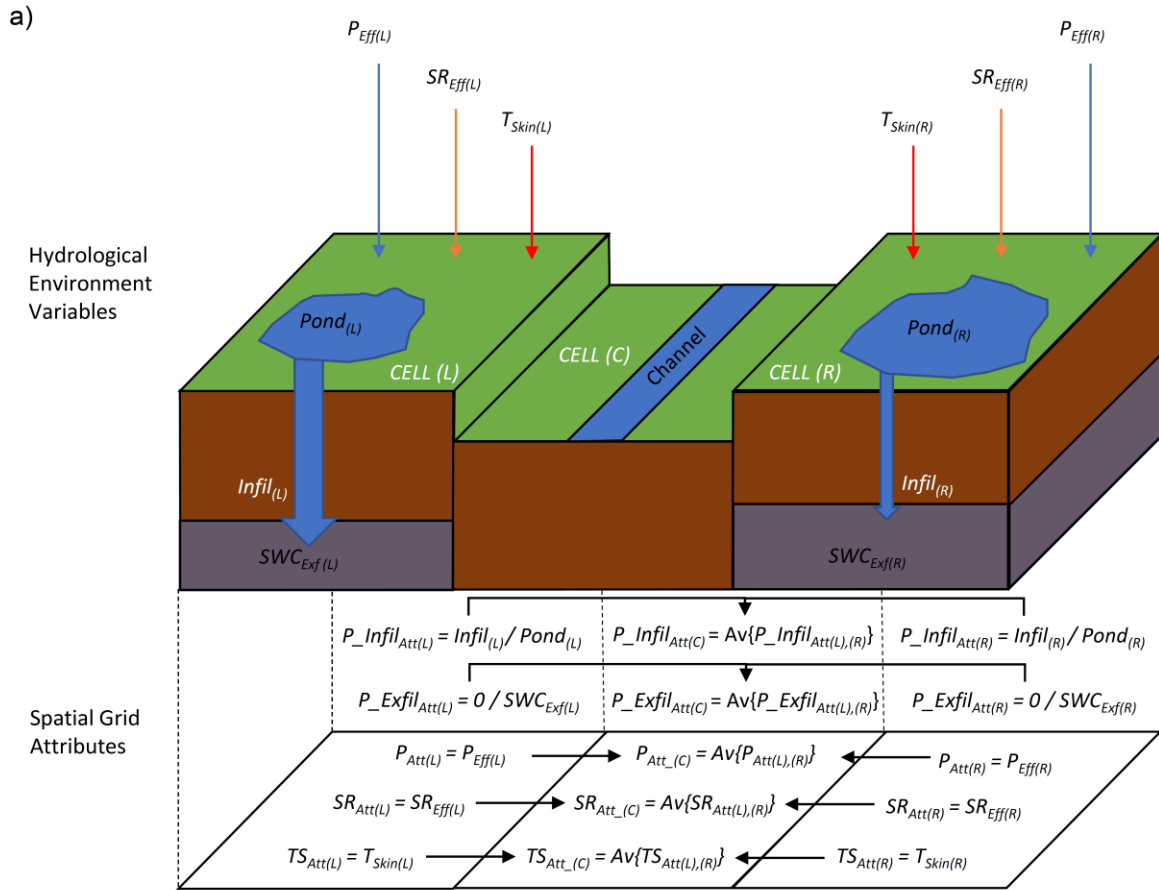
1068



1069

1070 *Figure 2: The abstract catchment system represented by the variables of the hydrological environment.*
 1071 Variables in black italic font are those provided explicitly by the hydrological environment: Effective
 1072 precipitation (P_{Eff}), effective solar radiation (SR_{Eff}), soil skin temperature (T_{Skin}), exfiltration ($Exfil$),
 1073 infiltration ($Infil$), ponded water prior to infiltration ($Pond$), soil water content prior to exfiltration
 1074 (SWC_{Exf}) and soil saturation deficit (Sat_{Def}). Grey, non-italicised font denotes fluxes capable of affecting
 1075 FIO-agent behaviour ($Run-on$ and $Run-off$) that are represented implicitly. Hydrological stores are
 1076 underlined. Note that $Exfil$ will be non-zero if Sat_{Def} is 0 – both are shown here for illustrative purposes
 1077 only.

1078

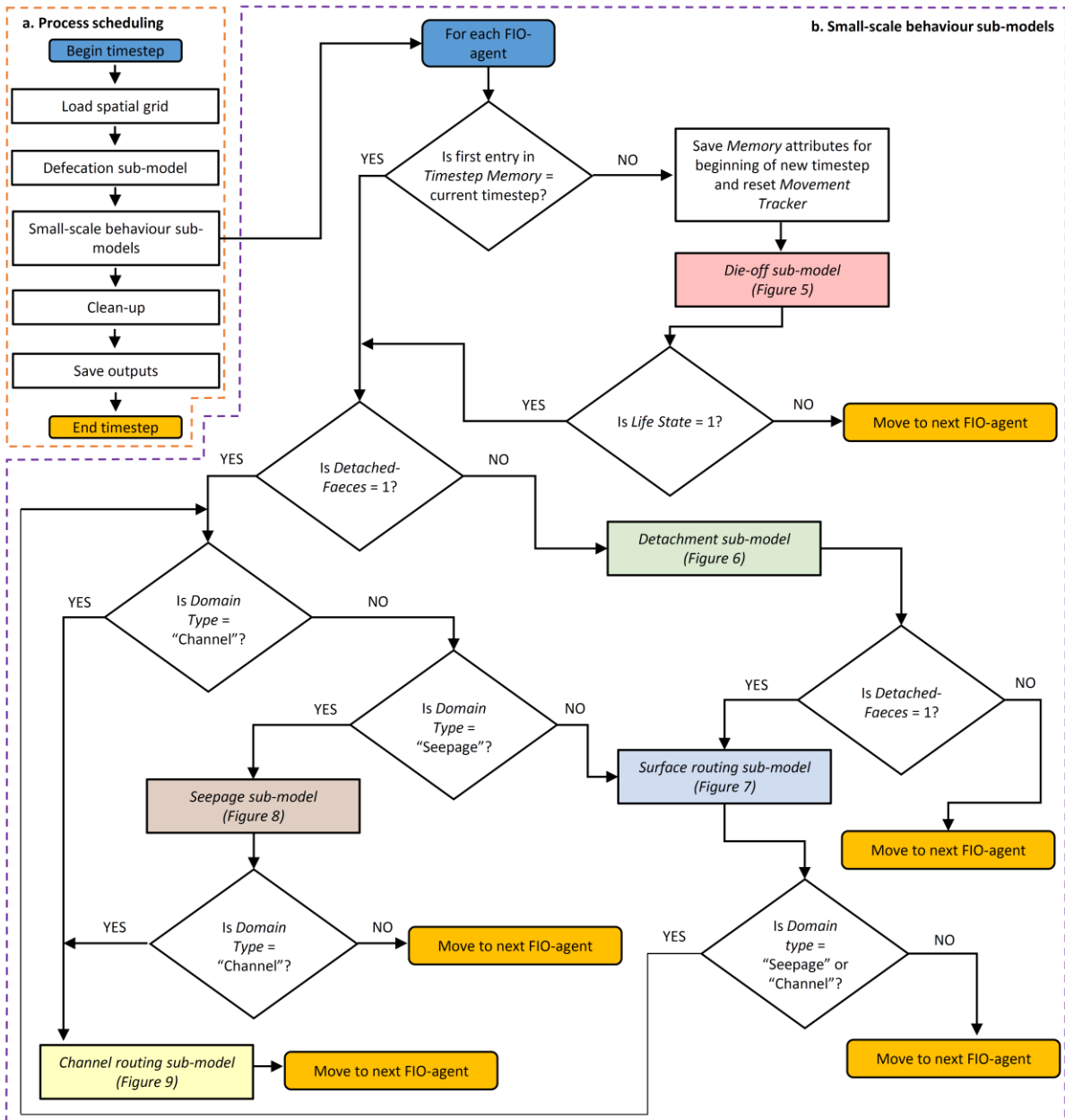


1079

1080 *Figure 3: Examples of how attributes of the spatial grid are calculated: a) Effective precipitation (P_{Eff}),*
 1081 *effective solar radiation (SR_{Eff}), soil skin temperature (T_{Skin}), ponded surface water prior to infiltration*
 1082 *($Pond$), infiltration flux ($Infil$) and soil water content prior to exfiltration (SWC_{Exf}) for two cells (L and*
 1083 *R) of the hydrological environment are used to calculate the attributes effective precipitation (P_{Att}),*
 1084 *effective solar radiation (SR_{Att}), soil skin temperature (TS_{Att}), probability of infiltration ($P_{_Infil_Att}$) and*

1085 probability of exfiltration ($P_{ExfilAtt}$) for the corresponding cells of the spatial grid, whilst the average
1086 of these are used as the attributes of a channel cell (C) into which the cells drain; b) $P_{SeepAtt}$ is
1087 calculated for two sequential timesteps – at timestep t , the new damage fraction (D_{frac_new}) is used in the
1088 calculation as the presence of a moderate number of livestock causes this to exceed the damage fraction
1089 from the previous timestep (D_{frac_old}) subject to exponential decay at rate k_d , whilst the reduction in
1090 livestock numbers at timestep $t+T$ causes the reverse to then be true. In a), black arrows denote how
1091 attributes/variables from cells (L) and (R) are used in determining average values for attributes/variables
1092 for cell (C).

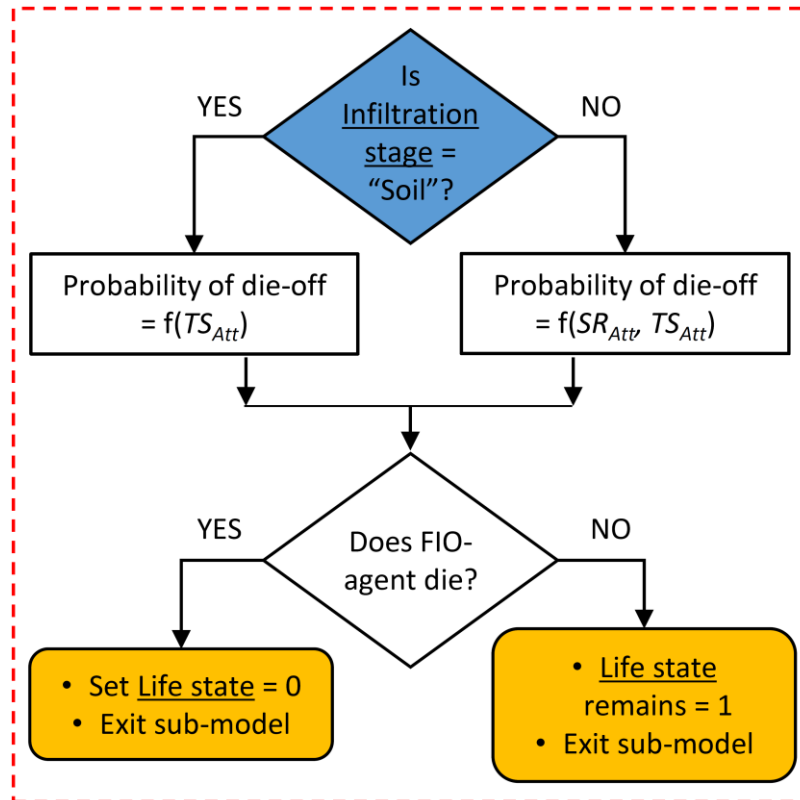
1093



1094

1095 *Figure 4:* Flowcharts showing a) The scheduling of processes during each timestep of MAFIO; b) The
 1096 sequence of small-scale behaviour sub-models that can be run for FIO-agents conditional on the values
 1097 of certain attributes (indicated by underlined font). Detailed flowcharts showing the functioning of
 1098 individual sub-models in b) are provided in Figures 5-9.

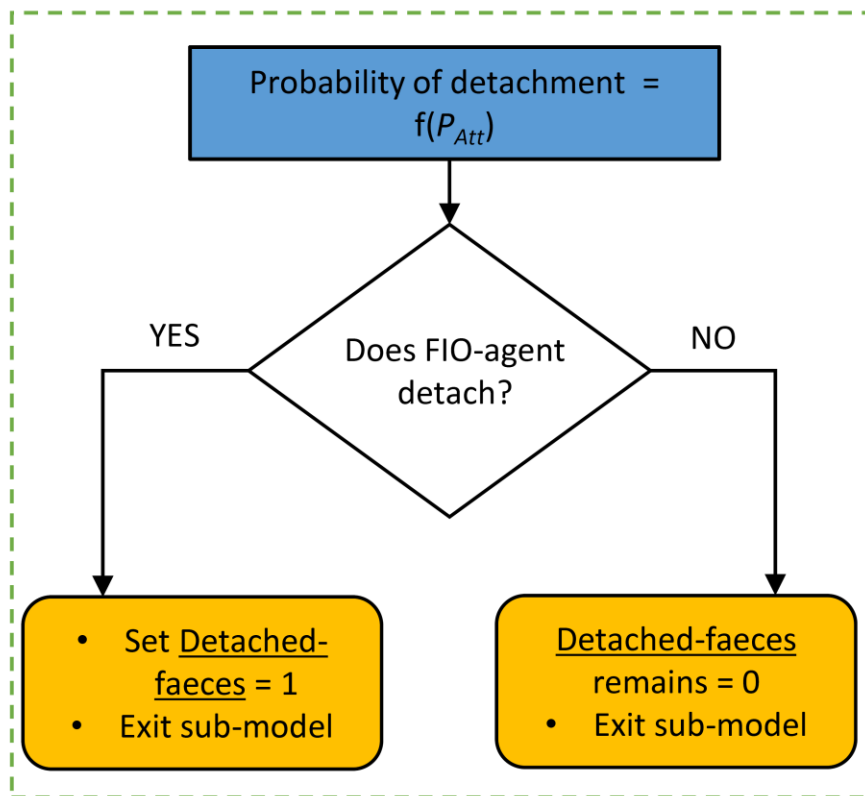
1099



1100

1101 *Figure 5:* Detailed flowchart for the Die-off Sub-model. Attributes of the spatial grid used by the sub-
 1102 model are denoted by italic font (*TS_{Att}* = soil skin temperature, *SR_{Att}* = effective solar radiation) whilst
 1103 attributes of FIO-agents that are used by the sub-model are underlined. Blue shapes denote the starting
 1104 operation of the sub-model whilst orange shapes denote where the sub-model can exit.

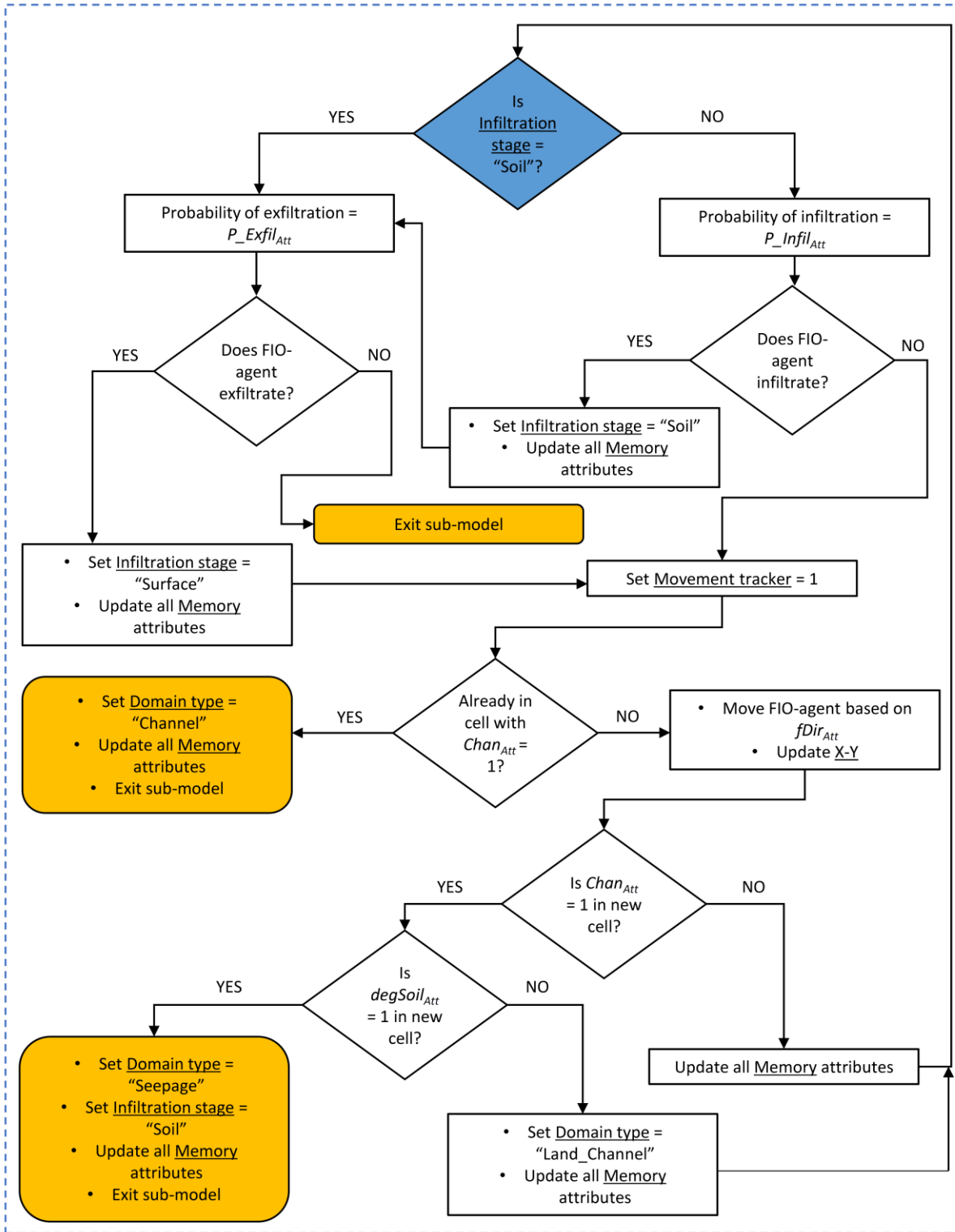
1105



1106

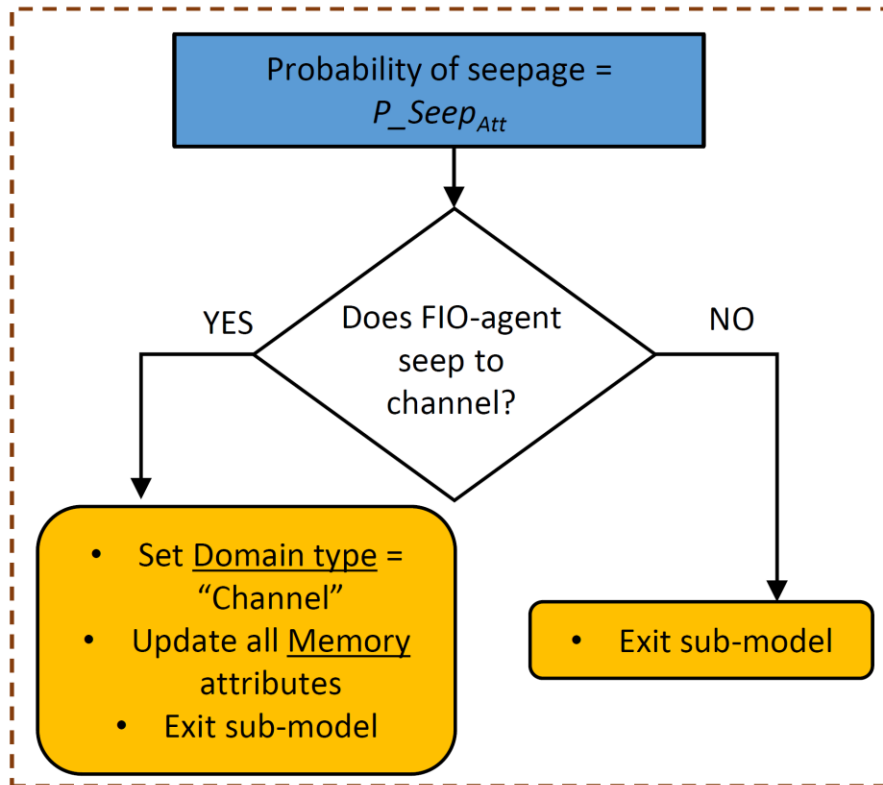
1107 *Figure 6:* Detailed flowchart for the Detachment Sub-model. Attributes of the spatial grid used by the
 1108 sub-model are denoted by italic font (P_{Att} = effective precipitation) whilst attributes of FIO-agents that
 1109 are used by the sub-model are underlined. Blue shapes denote the starting operation of the sub-model
 1110 whilst orange shapes denote where the sub-model can exit.

1111



1112

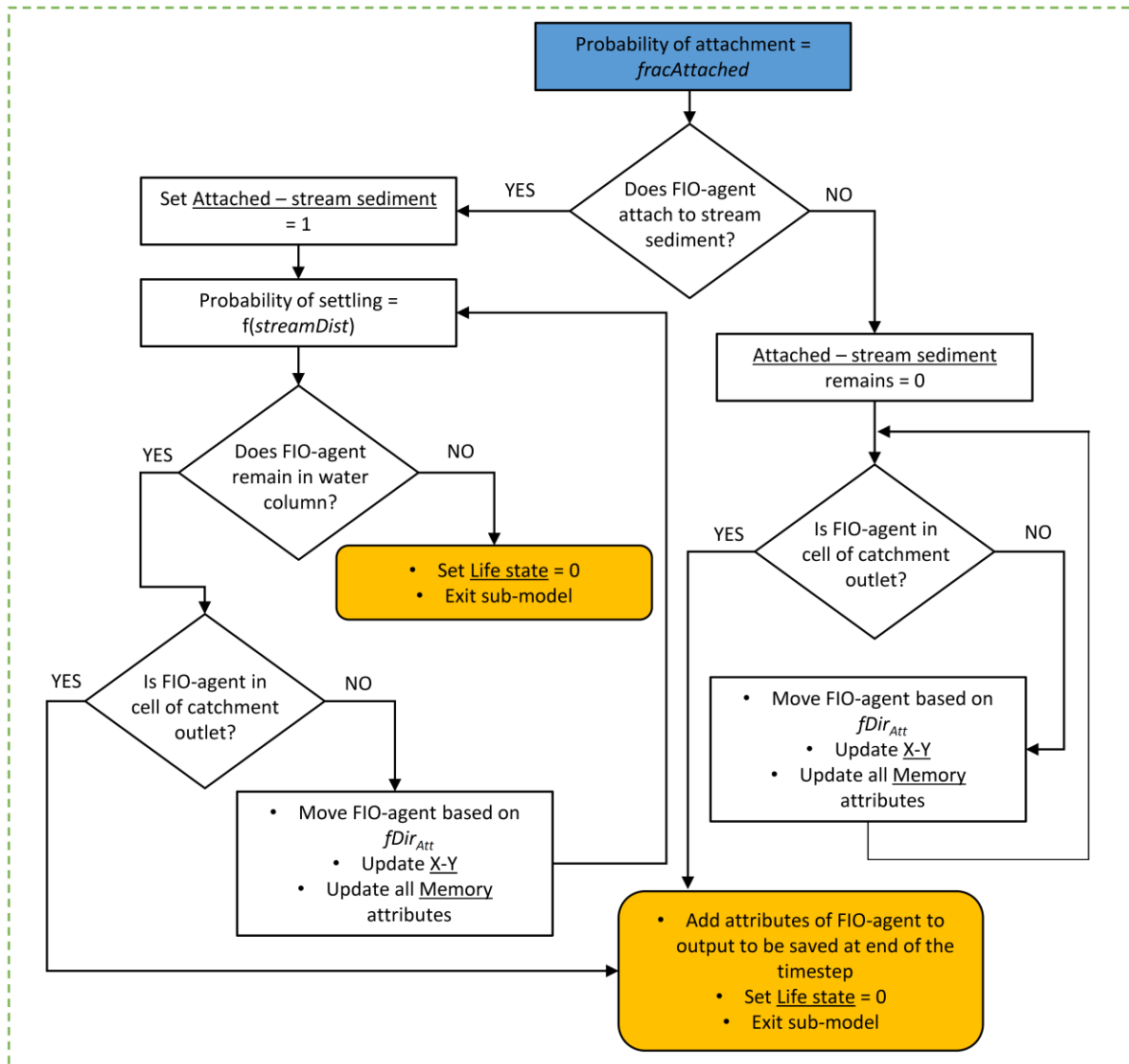
1113 *Figure 7: Detailed flowchart for the Surface Routing Sub-model. Attributes of the spatial grid used by*
 1114 *the sub-model are denoted by italic font (P_Infil_{Att} = probability of infiltration, P_Exfil_{Att} = probability*
 1115 *of exfiltration, $Chan_{Att}$ = channel cell, $fDir_{Att}$ = flow direction, $degSoil_{Att}$ = degraded soil in cell) whilst*
 1116 *attributes of FIO-agents that are used by the sub-model are underlined. Blue shapes denote the starting*
 1117 *operation of the sub-model whilst orange shapes denote where the sub-model can exit.*



1118

1119 *Figure 8:* Detailed flowchart for the Seepage Sub-model. Attributes of the spatial grid used by the sub-
 1120 model are denoted by italic font ($P_{SeepAtt}$ = probability of seepage) whilst attributes of FIO-agents that
 1121 are used by the sub-model are underlined. Blue shapes denote the starting operation of the sub-model
 1122 whilst orange shapes denote where the sub-model can exit.

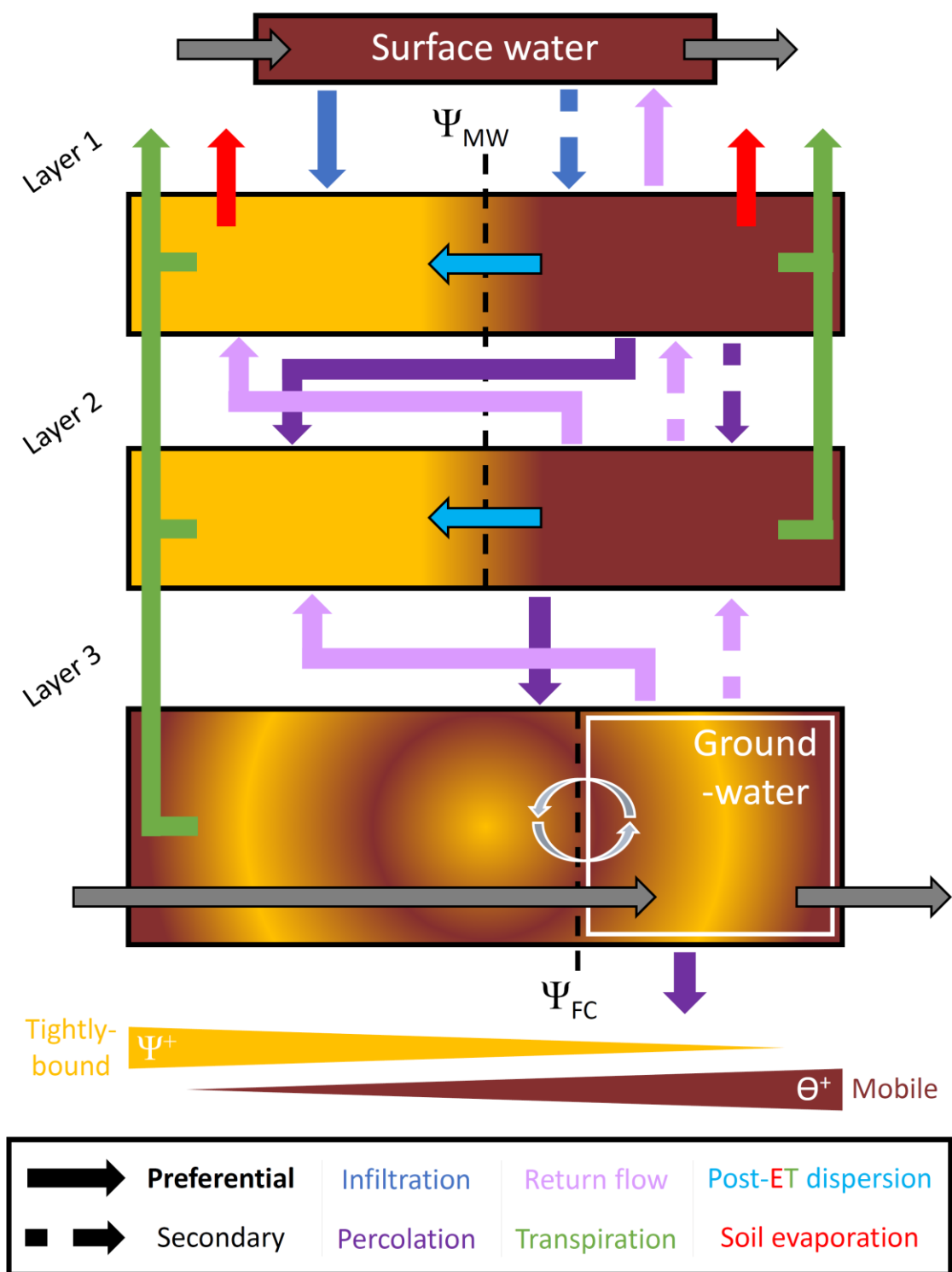
1123



1124

1125 *Figure 9: Detailed flowchart for the Channel Routing Sub-model. Attributes of the spatial grid used by*
 1126 *the sub-model are denoted by italic font ($fDir_{Att}$ = flow direction); an exception is $streamDist$ (straight-*
 1127 *line length of the channel), which is derived from the flow direction and cell size attributes of the spatial*
 1128 *grid. Attributes of FIO-agents that are used by the sub-model are underlined. $fracAttached$ is a global*
 1129 *model parameter denoting the fraction of FIOs assumed to be attached to sediment once in the stream.*
 1130 *Blue shapes denote the starting operation of the sub-model whilst orange shapes denote where the sub-*
 1131 *model can exit.*

1132



1133

1134 *Figure 10: A conceptual graphic of the below-canopy liquid stores and new water tracking scheme in*
 1135 *each pixel of ECH₂O-iso. The two-pore domain implementation (tightly-bound [TB] and mobile [MW])*
 1136 *in Layers 1 and 2 uses a tension threshold for mobile water (Ψ_{MW}). Liquid water exchanges between*
 1137 *layers (infiltration, percolation and return flow) follow the preferential pathways (solid arrows) unless*

1138 the recipient layer/domain is full, in which case secondary flow paths (dashed arrows) are triggered. In
1139 Layers 1 and 2, soil evaporation and plant transpiration draw from both domains in proportion to their
1140 respective water contents. Dispersion replenishes TB after evaporative losses, potentially emptying
1141 MW. The groundwater store is defined using a field capacity tension threshold (Ψ_{FC}), and tracking
1142 assumes full mixing with the non-saturated Layer 3.

1143

1144 **Tables**

1145 *Table 1: The dynamic and fixed attributes of the spatial grid. In the Description section, italics and underlining*
 1146 *are used to show where variables of the hydrological and catchment environments, respectively, are used in the*
 1147 *derivation of attribute values.*

Spatial grid attribute	Description	Units
<u><i>Dynamic</i></u>		
Effective precipitation (P_{Att})	<i>Effective precipitation (P_{Eff}) that reaches the surface after accounting for interception losses</i>	cm d ⁻¹
Effective solar radiation (SR_{Att})	<i>Effective solar radiation (SR_{Eff}) that is absorbed by the surface after accounting for transmission losses and reflection</i>	Ly hr ⁻¹
Probability of exfiltration ($P_{ExfilAtt}$)	The proportion of <i>soil water content prior to exfiltration (SWC_{Exf})</i> that constitutes the <i>exfiltration flux ($Exfil$)</i> to the surface	-
Probability of infiltration ($P_{InfilAtt}$)	The proportion of <i>ponded surface water prior to infiltration ($Pond$)</i> that constitutes the <i>infiltration flux ($Infil$)</i> into the soil	-
Probability of seepage ($P_{SeepAtt}$)	For cells with degraded soil adjacent to a channel, the probability of seepage to the channel as a function of <i>soil saturation deficit (Sat_{Def})</i> and <u>livestock counts</u> for the land parcel(s) associated with the cell	-
Soil skin temperature (TS_{Att})	<i>Soil skin temperature (T_{Skin})</i>	°C
<u><i>Fixed</i></u>		
Cell size ($size_{Att}$)	The length of a grid cell	Metres
Channel cell ($chan_{Att}$)	Whether (1) or not (0) a channel is present in the cell	Boolean
Degraded soil in cell ($degSoil_{Att}$)	If the cell contains a channel, whether (1) or not (0) there is an area of degraded soil surrounding the channel	Boolean
Flow direction ($fDir_{Att}$)	Identifier for the direction of steepest decent	-
Land parcel(s) (LP_{Att})	Identifier for the land parcel(s) to which the cell belongs based on its location within the catchment environment	-
Proportion of cell area occupied by channel ($P_{ChanAtt}$)	The channel area in the cell divided by the cell area	-
X-Y location (LOC_{Att})	The spatial co-ordinates of the cell	-

1148

1149

1150

1151

1152 *Table 2: The dynamic and fixed attributes of FIO-agents*

1153

FIO-agent attribute	Description	Possible values
<u>Dynamic</u>		
Attached-stream sediment	An abstract representation of whether the FIO-agent is attached to stream sediment once in the channel	<ul style="list-style-type: none"> • 0: Not attached (<i>Initial value</i>) • 1: Attached
Detached-faeces	An abstract representation of whether the FIO-agent is detached from a “faecal deposit”	<ul style="list-style-type: none"> • 0: Not detached (<i>Initial value for Domain type = Land or Land_Channel</i>) • 1: Detached (<i>Initial value for Domain type = Channel or Seepage</i>)
Domain type	The domain type that the FIO-agent currently occupies	<ul style="list-style-type: none"> • <i>Channel</i>: The FIO-agent is in a cell containing a channel, in the channel • <i>Land</i>: The FIO-agent is in a cell that does not contain a channel • <i>Land_Channel</i>: The FIO-agent is in a cell containing a channel, adjacent to the channel • <i>Seepage</i>: The FIO-agent is in a cell containing a channel, within an area of degraded soil
Infiltration stage	Denotes where the FIO-agent is within the vertical discretisation of the catchment environment	<ul style="list-style-type: none"> • <i>Surface</i>: The FIO-agent is on the land surface (<i>Initial value for Domain type = Land or Land_Channel</i>) • <i>Soil</i>: The FIO-agent is in the soil (<i>Initial value for Domain type = Seepage</i>)
Life state	Denotes if the FIO-agent is alive or dead	<ul style="list-style-type: none"> • 0: Dead • 1: Alive (<i>Initial value</i>)
Movement tracker	Tracker to record if the FIO-agent has moved on the current timestep	<ul style="list-style-type: none"> • 0: The FIO-agent has not moved (<i>Initial value</i>) • 1: The FIO-agent has moved

Timestep	The current timestep	<ul style="list-style-type: none"> The current timestep
X co-ordinate	The X co-ordinate of the FIO-agent	<ul style="list-style-type: none"> The current X co-ordinate of the FIO-agent
Y co-ordinate	The Y co-ordinate of the FIO-agent	<ul style="list-style-type: none"> The current Y co-ordinate of the FIO-agent
<i><u>Fixed</u></i>		
Land parcel	The ID of the land parcel into which the FIO-agent initially spawned	<ul style="list-style-type: none"> Land parcel ID
Livestock type	The ID of the host animal from which the FIO-agent came	<ul style="list-style-type: none"> Livestock ID
<i><u>Memory</u></i>		
Location memory	The X-Y co-ordinates previously occupied by the FIO-agent	NA
Timestep memory	The timesteps during which the FIO-agent has been in the model domain	NA
Infiltration state memory	The history of infiltration states the FIO-agent has experienced	NA
Domain type memory	The history of domain types that the FIO-agent has occupied	NA

1154
1155
1156
1157
1158
1159
1160
1161
1162
1163
1164

1165 *Table 3: The inputs required to characterise the catchment environment in MAFIO*

1166

Input	Description
<u>Maps</u>	
Catchment spatial extent	A binary map (1 and NaN) indicating the cells which comprise the catchment.
Cells upslope of channel	A map defining the cells which are immediately upslope of each cell containing a channel. The value of each cell is the unique channel ID of the cell containing a channel into which it drains.
Cells upslope of degraded soil	A map defining the cells which are immediately upslope of each cell containing an area of degraded soil. The value of each cell is the unique seepage ID of the cell containing degraded soil into which it drains.
Channel location	A map defining cells containing a channel. Each cell with a channel has a unique channel ID.
Channel width	A map of the channel widths for each cell containing a channel.
Degraded soil locations	A map indicating the locations of where soil adjacent to the channel is degraded. Each cell with degraded soil has a unique seepage ID.
Distribution of land parcels	A map of the land parcels in the catchment. Each land parcel has a unique ID.
Local drainage direction	A map of flow directions for each cell in the catchment based on the D8 algorithm.
<u>Timeseries</u>	
Livestock counts	A timeseries detailing the count(s) of each livestock type for each land parcel.
Stream access	A timeseries detailing for each unique channel ID, whether livestock from each land parcel in its list of associated land parcels have access to the stream.
<u>Lists</u>	
Land parcel(s) associated with degraded soil	For each unique seepage ID, a list of the land parcel(s) from which livestock can contribute to soil degradation.
Stream-associated land parcel(s)	For each unique channel ID, a list of the land parcel(s) on either side of the stream.

1167

1168

1169 Table 4: The parameters in the sub-models of MAFIO and their initial values based on simulating *E. coli* for
 1170 sheep and cattle with a daily timestep.

1171

Parameter	Description	Initial values*	Units	References*
<u>Defecation sub-model</u>				
<i>faecesConc</i> **	The concentration of FIOs per gram of faeces (wet weight)	[1.73×10^6 , 4.18×10^5]	MPN+ g ⁻¹	Avery et al. (unpublished data)
<i>FWeight</i>	The wet weight of a single defecation	[58.3, 2300]	grams	Welch (1982) [^]
<i>agentsRepresent</i>	The number of FIOs for which an FIO-agent is introduced	4.18×10^5	MPN	-
<i>defecationsPerDay</i>	The number of times an animal defecates per day	[16, 12]	day ⁻¹	Welch (1982)
<u>Die-off sub-model</u>				
<i>k_o</i> **	Inactivation rate constant at a reference temperature of 20 °C	[0.242, 0.090]	day ⁻¹	[Moriarty et al., (2011); Himathongkham et al., (1999)]
<i>θ</i> **	Temperature sensitivity parameter	[1.095, 1.069]	-	[Moriarty et al., (2011); Himathongkham et al., (1999)]
<i>α_{SR}</i>	Proportionality constant for die-off due to solar radiation	1	-	Thomann and Mueller (1987)
<u>Detachment sub-model</u>				
<i>k_{release}</i> **	Rate constant for rainfall-induced detachment of FIO-agents	0.153	cm ⁻¹	Blaustein et al. (2016) [^]
<u>Seepage sub-model</u>				

<i>LUs</i>	The number of livestock units represented by an individual animal	[0.12, 1]	LU ⁺⁺	Natural England (2013)
<i>riskBands</i>	The boundaries of each risk band	0 < Low ≤ 1 1 < Medium < 4 High ≥ 4	LU ha ⁻¹	Oliver et al. (2009)
<i>dFrac_Bands</i>	The damage fractions associated with each risk band	Low: 0.43 Medium: 0.53 High: 0.72	-	Sheath and Carlson (1998)
<i>k_d</i>	The rate constant for damage fraction decay	0.0063	day ⁻¹	Elliot et al. (2002) [^]
<u>Channel routing sub-model</u>				
<i>fracAttached</i>	Fraction of FIOs attached to stream sediment	0.8	-	Hipsey et al. (2008)
<i>λ_{sed}</i>	Distance decay rate constant	0.00037	m ⁻¹	Kay and McDonald (1980)

* Values in square brackets are for [sheep, cattle]; + MPN: most-probable number; ++ LU: livestock units; ** Parameter values are for *E. coli*; ^ Derived from data presented in references.

1172

1173