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## **eQTL set-based association analysis identifies novel susceptibility loci for Barrett's esophagus and esophageal adenocarcinoma**

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## **Abstract**

### **Background:**

Over 20 susceptibility single-nucleotide polymorphisms (SNPs) have been identified for esophageal adenocarcinoma (EAC) and its precursor, Barrett's esophagus (BE), explaining a small portion of heritability.

### **Methods:**

Using genetic data from 4,323 BE and 4,116 EAC patients aggregated by international consortia including the Barrett's and Esophageal Adenocarcinoma Consortium (BEACON), we conducted a comprehensive transcriptome-wide association study (TWAS) for BE/EAC, leveraging Genotype Tissue Expression (GTEx) gene expression data from six tissue types of plausible relevance to EAC etiology: mucosa and muscularis from the esophagus, gastroesophageal (GE) junction, stomach, whole blood, and visceral adipose. Two analytical approaches were taken: standard TWAS using the predicted gene expression from local expression quantitative trait loci (eQTLs), and set-based *SKAT* association using selected eQTLs that predict the gene expression.

### **Results:**

While the standard approach did not identify significant signals, the eQTL set-based approach identified eight novel associations, three of which were validated in independent external data (eQTL SNP sets for *EXOC3*, *ZNF641* and *HSP90AA1*).

### **Conclusions:**

This study identified novel genetic susceptibility loci for EAC and BE using an eQTL set based genetic association approach.

### **Impact:**

This study expanded the pool of genetic susceptibility loci for EAC and BE, suggesting the potential of the eQTL set based genetic association approach as an alternative method for TWAS analysis.

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## Introduction

Incidence of esophageal adenocarcinoma (EAC) has risen sharply over recent decades(1-4), and it is now the predominant subtype of esophageal cancer in the US and many other western countries. Patients diagnosed with advanced EAC have a 5-year survival rate below 20%(5-10). Progress has been made in identifying genetic and environmental risk factors for EAC and its epithelial precursor lesion, Barrett's esophagus (BE)(11). GE reflux(12,13), obesity(14,15), and tobacco smoking(16,17), collectively explain up to ~75% of cancer risk(18-20). While over 20 susceptibility single-nucleotide polymorphisms (SNPs) have been identified through genome-wide association studies (GWAS) in the Barrett's and Esophageal Adenocarcinoma Consortium (BEACON) and related efforts(21-29), these loci explain only a small portion of overall heritability ( $h^2_g$  estimated as 0.25 for EAC; 0.35 for BE)(30), and few have been linked specifically to progression to cancer(22,29).

One of the notable methodological advances in the post-GWAS era is integrating the transcriptome into genetic association analyses(31,32). Evidence is abundant that trait-associated SNPs are more likely to be expression quantitative trait loci (eQTLs)(33), which are pervasive in the human genome(34-38). Motivated by the premise that eQTL may influence disease phenotypes by altering gene expression levels, association approaches leveraging eQTL and transcriptome data in genotype-tissue expression (GTEx)(38), namely transcriptome-wide association studies (TWAS), have become a mainstream approach in post-GWAS analyses, leading to the discovery of multiple novel susceptibility genes(39-42), for prostate(43), ovarian(44), breast(45), and colorectal cancers(46,47). Notably, the initial TWAS method - *PrediXcan*(31) - first

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builds a genetic prediction model for gene expression, then assesses genetically predicted gene expression for its association with a trait of interest, much resembling the classical instrumental variable regression approach in econometrics. In the same vein, newer methods for TWAS(32,48) removed the requirement of individual-level genetic data by exploiting GWAS summary statistics and genetic correlation data from external data such as the 1000 Genomes Project. Recognizing a portion of eQTL regulation of gene expression can be conservative across tissues, new methodological development for TWAS has been focused on leveraging multiple tissues available in GTEx for improving power of genetic prediction and subsequent association(49,50). On the other hand, it has been noted with caution recently that TWAS can be prone to spurious results with expression data from non-trait-related tissues or cell types, and that the best practice may be choosing the most mechanistically related tissue(s) available(51).

In our view, the main challenge to apply TWAS to the EAC genetic research is that there is not yet a large set of BE samples, the mechanistically relevant tissue for EAC development, with both germline genotypes and transcriptome data available for eQTL mapping. Although the inherited genetic component of risk for BE largely coincides with that for EAC(30), the cellular origin of BE remains controversial, with hypotheses ranging from residual embryonic cells at the GE junction to undifferentiated gastric cells in the cardia(52-54). While the GTEx Project collected four upper gastrointestinal tract tissues, including mucosa and muscularis from the esophagus, GE junction, and stomach, a limitation of bulk RNA-sequencing data is that transcriptome profiles of rarer constituent cell types (such as progenitor cells) may not be well delineated.

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In this work, we conducted a comprehensive TWAS study for BE/EAC, leveraging six GTEx (V8) tissue types of plausible relevance to EAC etiology: mucosa and muscularis from the esophagus, GE junction, stomach, visceral adipose, and whole blood. Inclusion of the latter two tissues is in recognition that tissues beyond the esophago-gastric mucosa are likely to contribute biologically to the origins of BE/EAC. Abdominal obesity is a risk factor for these conditions, which not only affects reflux severity, but also increases levels of systemic inflammation through release of secreted mediators (55-57). Chronic inflammation is considered an important driver of BE/EAC pathogenesis, and the roles and contributions of circulating and infiltrating immune cells are under active investigation (58). We selected eQTLs collectively predicting RNA-sequencing based expression and built prediction models. The eQTLs predicting protein-coding genes were assessed for gene-level associations with BE/EAC risk using a discovery dataset (BEACON/Cambridge GWAS), and top signals identified were then advanced for evaluation using an independent GWAS dataset from Bonn, Germany. We used two methods to assess gene-set associations for selected eQTL: *i*) standard *PrediXcan* (31), computing a linear combination prediction of gene expression, and *ii*) the sequence kernel association test (*SKAT*)(59), testing gene-set association among selected eQTLs that predict gene expressions. Originally developed for rare-variant association tests, *SKAT* was used here to assemble genetic associations from eQTL without using the prediction weights derived from an extant gene-expression dataset, e.g., GTEx. An eQTL-based aggregate association strategy has been reported previously (60), though the previous method used the sum of 1-df chi-square values for the individual eQTLs. The following rationales motivate the eQTL aggregation strategy:

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i) the existing GTEx tissue types may not capture the cellular origin of BE; ii) even if the etiologically-relevant cell types were contained in one of GTEx tissues, genetically predicted gene expression derived from bulk tissue RNA-seq profiles may not adequately represent the genetic component of gene expression in rarer yet etiologically-relevant cell types within that tissue. Therefore, we hypothesize that the gene-expression prediction weights for eQTLs derived from GTEx may not be always appropriate for the targeted genetic risk prediction for BE and EAC, and we postulate that a more flexible set-based global test of selected eQTLs may improve the likelihood of capturing genetic associations with disease risk which are otherwise obscured when evaluating surrogate gene expression measures from bulk tissue.

## **Materials and Methods**

### **Individual-level data and summary statistics from existing GWAS**

Genome-wide association data from three genetic studies were obtained for this analysis. Given that our analytic plan encompassed a multitude of correlated analyses and included exploratory methodologic comparisons, e.g., two analysis strategies for six tissues and 3 trait comparisons of interest (BE vs control, EAC vs control and BE/EAC vs control), a discovery-validation approach was adopted to better control the potential false positive results. For the discovery set, individual-level genotype data were available from the BEACON consortium (dbGaP phs000869.v1.p1) (2,413 BE cases, 1,512 EAC cases and 6,718 control participants) and the Cambridge GWAS (873 BE cases, 995 EAC cases, and 3,408 control participants); for validation, SNP summary statistics were available from the Bonn GWAS (1,037 BE cases, 1,609 EAC cases, and 3,537 control participants). After quality control, the discovery set included 702,492

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SNPs on autosomal chromosomes. An additional 4,541 controls of European ancestry were obtained from the database of Genotypes and Phenotypes (dbGaP) (phs000187.v1.p1, phs000196.v2.p1, and phs000524.v1.p1) and merged with the BEACON discovery data to increase statistical power to detect risk loci. The Michigan imputation server (61) was used to impute genotype data on chromosomes 1-22, with the most accurate and largest panel - the Haplotype Reference Consortium (HRC) (Version r1.1 2016) for European (EUR) as the population reference. Imputed genotype data included 5,312,829 SNPs with imputation quality score  $> 0.4$ ,  $MAF > 0.05$ , call rate  $> 95\%$  and Hardy-Weinberg equilibrium P-value  $> 1e-5$ . For the Bonn dataset, imputation was previously carried out using the 1000 Genomes Phase1 EUR reference panel, and imputed genotype data included a total of  $\sim 9$  million SNPs with minor allele frequency  $> 0.001$ .

### **GTEx germline sequencing data and RNA-seq transcriptome data for eQTL prediction of gene expression**

GTEx data (V8) from subjects of European ancestry were used in this analysis. RNA-seq gene expression data were retrieved from 6 tissues of plausible biologic relevance to EAC development (esophagus GE junction:  $n=275$ , esophagus - mucosa:  $n=411$ , esophagus - muscularis:  $n=385$ , stomach:  $n=260$ , adipose - visceral:  $n=393$ , and whole blood:  $n=558$ ). Transcripts per million (TPM) data were downloaded, and the trimmed mean of M values (TMM) normalization method was implemented in *edgeR*(62). For each gene in a tissue, gene expression values were standardized across samples. SNP genotypes were obtained from whole genome sequencing data for  $\sim 46,569,000$  variants.



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The expression levels for a gene were modeled using an *ElasticNet* linear model with local SNPs in a 1Mb region flanking the TSS of the gene, and covariates including the top four genotype principal components, top 15 Probabilistic Estimation of Expression Residuals (PEER) factors, sex, age, sequencing platform indicator (Illumina HiSeq 2000 or HiSeq X), and sequencing protocol indicator (PCR based or PCR-free). The elastic net model was implemented using the R package *glmnet*(63). Highly correlated SNPs with Pearson correlation  $>0.9$  were removed before running the elastic net model. The penalty parameter was selected by the minimum ten-fold cross validation error. The ten-fold cross-validated  $R^2$  for genetically predicted gene expression was used to summarize the strength of genetic prediction. The distribution of  $R^2$  for predicting gene expressions in a tissue is displayed by violin plot. Genes with estimated  $R^2 > 0.01$  (correlation  $> 0.1$ ) for a tissue entered subsequent genetic association analysis, using the SNPs with non-zero estimated coefficients identified as eQTLs.

### **eQTL set-based association analysis in the discovery set (Beacon and Cambridge individual-level data)**

For each of six tissues and three trait comparisons (BE vs Control, EAC vs Control, BE/EAC combined vs Control), gene-set association analyses were conducted by the following two approaches. First, in the standard TWAS approach, predicted gene-expression from the GTEx-derived *ElasticNet* model was assessed for its association with the trait by a logistic model, adjusting for sex, age, and the top six genotype principal components. Second, the selected eQTLs from the GTEx-derived *ElasticNet* model were assessed for their collective association with the trait by *SKAT*(59),

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adjusting for the same set of covariates. Manhattan plots were drawn to show p-values for gene SNP sets by chromosome. False discovery rate (Benjamini-Hochberg FDR) was used for to account for multiple testing. For genes of interest for discovery, individual SNP-trait associations were also assessed and plotted using *LocusZoom* software. To determine whether an identified gene-set association is caused by a previously identified risk SNP in the neighborhood, a *SKAT* model was also fitted to include the known GWAS SNP in the region.

### **Validation of the discovered eQTL set based associations in the Bonn dataset**

Gene-level eQTL SNP sets putatively associated with a trait were next evaluated using Bonn GWAS summary data. The *SKAT* association statistics for the gene sets were approximated by a score-statistic method(64), using univariate summary statistics and the genetic correlation matrix computed from European ancestry participants of the 1000 Genomes Project. For a few SNPs in the discovery set but missing in the validation set due to different imputation panels, we used the closest SNPs within 50 bp and with correlation  $> 0.6$ , whenever available, as the proxy to minimize the impact of missing SNPs. To account for multiple testing in the validation stage, the Hochberg adjusted p-value for controlling family-wise error rate was used.

### **Data availability**

The BEACON data with supplemented controls were obtained from dbGaP (phs000869.v1.p1, phs000187.v1.p1, phs000196.v2.p1, and phs000524.v1.p1). The GTEx genotype and gene expression data were obtained from dbGaP (phs000424.v8.p2).

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## Results

### ***cis-eQTL predicted gene expressions in six etiologically relevant tissues***

Transcriptome data and germline whole-genome sequencing data from GTEx (V8) were analyzed for building genetic prediction models for gene expressions in each of six etiologically relevant tissues for BE/EAC: esophageal mucosa (n=411), esophageal muscularis (n=385), GE junction (n=275), stomach (n=260), adipose – visceral (n=393), and whole blood (n=558). Common SNPs (MAF>0.05) located within  $\pm 500$ kb of the transcription starting site (TSS) of a gene were identified from GTEx whole-genome sequencing data and selected to predict the transcript abundance by the *ElasticNet* method. Figure 1a shows the violin plots of  $R^2$  for genes with at least 1 SNP being selected and  $R^2 \geq 0.01$  (correlation of observed and predicted gene expression  $\geq 0.1$ ) in the six tissues. The four tissues in the upper GI tract (esophageal mucosa and muscularis, GE junction, and stomach) have a greater number of predictable genes and higher  $R^2$  in this subset: esophageal mucosa has the largest number of genes with  $R^2 \geq 0.01$  (n=7463); GE junction has the highest median (0.037) despite the smaller sample size for junction tissue. There is substantial variability among the numbers of “genetically predictable” genes across tissues (5160 in blood ~ 7463 in esophageal mucosa), and the genes shared between tissues. The latter is exemplified by a Venn diagram in Figure 1b, which shows the overlapping set between the three esophageal tissues. Between any two tissue types, 35~45% of genes are not shared, underscoring the significance of both cross-tissue and tissue-specific genetic regulation of gene expression.

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***eQTL set based association analysis identified susceptibility loci for BE and EAC in BEACON/Cambridge discovery set***

The selected eQTLs in the genetic prediction models were assessed for association with BE, EAC, or BE/EAC as a combined trait in BEACON/Cambridge discovery set, using two methods: 1) the standard TWAS approach, where the predicted gene expression from the *ElasticNet* model was assessed for its association with the trait in a logistic regression model; 2) the gene-set association method *SKAT*, using the selected eQTLs from the *ElasticNet* model. Across six tissues and for three trait association comparisons, there are a total of 116,853 gene set associations being tested. Because of high correlations between p-values from the same genes across different tissues and trait comparisons, the Bonferroni procedure can be overly conservative. Instead, false discovery rate (FDR) was used to adjust for multiple testing, in part because it can effectively account for correlation. Figure 2 shows the Manhattan plots of p-values for the three comparisons using the two methods (standard TWAS on the bottom of each panel and *SKAT*-eQTL on the top). No genes analyzed by the standard TWAS approach satisfied  $FDR < 0.05$  (minimum  $FDR = 0.187$ , e.g., *EXOC3*, *BARX1*, and *LDAH*). In contrast, the *SKAT*-eQTL method identified a total of twenty-one genes with significant associations at  $FDR < 0.05$  (red dotted line in Figure 2), representing a mix of novel and known loci. Table 1 shows eight novel eQTL set-based associations in six loci that either have not been reported previously or are independent of the known GWAS SNP in conditional analysis. Table 2 shows thirteen loci that have been previously linked to susceptibility, containing putative genes including *LDAH*, *BARX1*, *ALDH1A2*, and *CRTC1*.

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One consistent theme in Table 1 and 2 is that *SKAT*-eQTL produced uniformly smaller p-values than the standard TWAS method for these eQTL set associations, suggesting that the weighted linear combination of eQTL for predicting these gene expressions may not always be powerful to capture genetic associations. For example, *EXOC3* is located in locus 5p15.33, 220kb away from the known risk SNP rs9918259, with its flanking region containing 28 selected *cis*-eQTLs in adipose tissue, explaining 4.2% of variability in its gene expression. When assessed by *SKAT*, this set of SNPs was significantly associated with BE/EAC with p-value  $8.24 \times 10^{-6}$  (FDR=0.0365). The standard TWAS analysis yielded a larger p-value,  $1.81 \times 10^{-5}$ . Figure 3 shows the regional plots of the three novel loci that were discovered in BEACON/Cambridge discovery set and validated in Bonn data. Specifically, Figure 3a shows a cluster of *cis*-eQTLs, located in the *EXOC3* gene, that were individually associated with BE/EAC at a moderate level of significance (p-value  $10^{-2} \sim 10^{-4}$ ). Previous meta-analysis identified rs9918259 as a risk SNP in *CEP72/TPPP*. Adjusting for rs9918259 in the *SKAT* regression model attenuated the p-value for *EXOC3* gene set association from  $8.24 \times 10^{-6}$  to  $2.98 \times 10^{-3}$ , suggesting that the SNP set of eQTL predicting *EXOC3* gene expression may add new evidence for association at this locus.

The remaining seven eQTL sets in Table 1 are all >10 Mb away from the closest existing GWAS SNP; adjusting for existing GWAS SNPs did not reduce the gene-set association significance. Of top interest is *HSP90AA1(65,66)*, which is located on chromosome 14, with no GWAS risk SNPs previously identified. This gene was identified by association of 94 eQTLs in blood with BE. The regional plot in Figure 3e shows widespread individual eQTL associations over a 1-Mb window around this gene,

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exemplifying the power of gene-set association in aggregating signals that may not reach genome-wide significance individually. Four eQTLs of *ZNF641(67)* gene were identified in esophageal junction, collectively associated with BE/EAC (p-value= $3.81 \times 10^{-6}$ ), and also individually associated with BE/EAC with less significance (Figure 3e). Table 2 shows loci identified by eQTL gene-set association analyses which have been previously linked to risk of BE and EAC. All are located within 0.5 Mb of an existing GWAS SNP; all but three (*CRTC1*, *SSBP4* and *JUND*) became non-significant when adjusting for the closest GWAS SNP(s). Six eQTL sets including *CRTC1*, *SSBP4* and *JUND* are from 19q13.11, a locus harboring three risk SNPs in *CRTC1* that have been consistently detected in previous GWAS efforts. The top association in this locus is *THEM161A* (p= $9.02 \times 10^{-10}$ ). Adjusting for the three known risk SNPs in the region (rs10423674, rs10419226, rs199620551) does not completely remove association significance for *CRTC1*, *JUND*, and *SSBP4*, suggesting that this region may have independent risk alleles other than the three known risk SNPs. The locus 3q27.1 contains rs9823696, the only previous risk SNP linked to EAC but not BE. Rather than *HTR3C* and *ABCC5*, the nearest genes to this GWAS variant, gene-set analysis implicated *YEATS2* and *ABCF3*. The remaining genes in Table 2 were reported in prior GWAS as candidate risk genes based on their proximity to index SNPs: *LDAH/GDF7* (2p24.1), *BARX1* (9q22.32), and *ALDH1A2/AQP9* (15q21.3).

***Three eQTL set associations in Table 1 were replicated using Bonn GWAS data***

We evaluated all association signals identified in Table 1 using the *SKAT* method and summary statistics from the Bonn GWAS (with 1000 Genomes EUR LD structure) (Table 3). Because the trait association analyses were based on HRC imputation, and

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the available Bonn summary statistics were imputed based on 1000 Genomes data (V3), a small number of SNPs were missing for some of the eight genes. eQTL SNP sets for three genes had evidence of replication in the Bonn data, using the Hochberg adjusted p-value  $<0.05$  as the threshold for confirming candidate associations: *EXOC3*, *ZNF641*, and *HSP90AA1*. In particular, *EXOC3* ( $p=0.0000185$ ) and *ZNF641* ( $p=0.00378$ ) remained significant even using the stringent Bonferroni correction. Twelve out of 26 eQTLs for *EXOC3* had a univariate p-value  $<0.05$  (minimum p-value  $1.69 \times 10^{-7}$ ). Three out of four eQTLs for *ZNF641* also have univariate p-values  $<0.05$ .

## Discussion

Advanced esophageal adenocarcinoma is a deadly disease with rising incidence in Western countries. International efforts including BEACON and other European studies have identified ~20 susceptibility SNPs, though collectively these risk SNPs explain only a small portion of the genetic heritability. Polygenic risk scores (PRS) based on GWAS hits have not been able to significantly improve prediction beyond environmental risk factors. In our view, current genetic studies have reached a plateau in discovery, largely due to the rarity of the cancer and the limited available sample sizes. In this work we conducted TWAS for EAC and BE, using data from six etiologically relevant GTEx tissues. The standard TWAS method using predicted gene expression was compared to a novel approach that assessed gene-level eQTL-set associations by *SKAT*. Individual-level genetic data from BEACON/Cambridge were used as the discovery set and summary statistics of genetic data from Bonn were used as the validation set. Using a



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significance threshold of  $FDR < 0.05$  in the discovery set, standard TWAS identified no associations, while the *SKAT* approach yielded 13 eQTL set associations in 11 loci. Among them, 8 eQTL set associations (5 loci) are novel findings, either representing novel susceptibility regions without previously identified risk SNPs, or in the case of *EXOC3*, a novel signal independent of known risk SNPs in the neighborhood. Among the genes from the known loci, our results suggest that there are potentially susceptibility genes at 19p13.11 independent of the three known risk SNPs. Notably, the loci identified by eQTL set associations largely did not overlap with loci previously reported for gastro-oesophageal reflux disease (GERD), a major risk factor for BE/EAC and a clinical trait by itself. For eQTLs in Tables 1 and 2, only one eQTL rs9636202 (*SYNA1*) was found to be a GERD risk locus previously reported (68). These results represent a significant advance in identifying novel inherited genetic risk associations for BE/EAC, since the publication of the first GWAS meta-analysis in 2016 (22). While we underscore the importance of discovering new candidate risk loci for a rare cancer with limited study samples available, we also acknowledge that caution is needed in interpreting eQTL set associations. Functional laboratory studies are essential to identify causal variants and genes that are driving observed associations.

One interesting observation is that there is no finding from the standard approach, while *SKAT-eQTL* produced 13 significant eQTL set associations. All set associations in Tables 1 and 2 have smaller p-values from *SKAT-eQTL*, some substantially more significant (e.g., *HSP90AA1* and *KRTAP5-8*, for which p-values of the standard TWAS are greater than 0.05). This analysis is a single observation of applying two methods applied to GWAS datasets, therefore does not establish the power comparison between

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the two methods. A formal power comparison using simulated genetic datasets is necessary and will be pursued in future work. Here we merely conjecture why an eQTL-set ensemble association may be advantageous in the context of BE/EAC studies. The prevailing theory for TWAS using imputed gene expression is that the genetic component of gene expression for genes relevant to disease etiology can be accurately predicted, using etiologically relevant tissue samples. As noted previously, however, a complicating factor is that most tissues are comprised of multiple cell types in varying abundance, and only a subset of these cell types – often representing a small fraction of overall cells – may be the most relevant to disease development. This indeed may be the case for BE/EAC, as candidates for the cell-of-origin include subpopulations of precursors in the GE junction or gastric cardia. That said, cancer development is a multi-faceted process involving not only stem-like precursor cells, but also the tissue microenvironment, comprised of and influenced by multiple constituent cell types. It is quite possible that for this reason, a more flexible gene-set association method, such as *SKAT*, may outperform standard TWAS. If this hypothesis is true, one may envision that the SKAT-eQTL association approach could be applied successfully on a wider scale and similarly help accelerate discovery of novel loci for other types of cancers.

Compared to the standard GWAS analysis for individual SNP associations, the eQTL set based aggregation approach provides better power in detecting loci that contain multiple eQTL association signals. In Supplementary Figure 1, we show the locus zoom plots for the individual SNP associations combining the discovery and validation set from the eight loci identified in Table 1. None of the SNPs in these regions reached the p-value cutoff of  $5 \times 10^{-8}$ . However, the set-based method can assemble and

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detect multiple eQTL associations in a locus with single variants reaching moderate levels of significance.

Rarely employed in TWAS, the discovery-validation design we adopted is partly driven by the multitude of analyses we intended to conduct using two analytical approaches and 6 potentially relevant tissues, and for 3 trait comparisons of interest, as well as the lack of access to individual-level genetic data in Bonn study. The latter is a limitation of current analysis. Confirmation of discovery-stage associations, using an independent validation dataset, reduces the possibility of false positive discoveries. As noted before, though motivated by the goal of deciphering causal pathways of disease etiology, the original TWAS approach is not immune to spurious findings. This is particularly true for the *SKAT-eQTL* approach – it is essentially a gene-set association method assembling risk associations that are individually unlikely to survive the multiple-testing penalty. Three novel eQTL set associations (*EXOC3*, *SENP6*, *HSP90AA1*) were validated by the Bonn summary data, each of which has multiple individual risk SNPs with a moderate level of association (Figure 3).

We end our discussion with several other limitations and future work. First, while delivering several novel susceptibility signals, the power of our TWAS is limited by the sample size of BE/EAC cases due to its rarity. Compared to existing TWAS for other cancers, our sample size is much smaller. New population-based genetic studies are needed to improve power and further advance EAC genetic research. Second, our analyses were restricted to European ancestry participants. Although the incidence of EAC in whites is much higher than that in African Americans, future studies are needed for eQTL and TWAS in broader populations. Third, although our genetic findings are

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promising, experimental studies are needed to understand the mechanisms of genetic risk predisposition.

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## Tables

Table 1. Novel eQTL set-based genetic associations with BE, EAC, or BE/EAC.

| Locus    | Gene            | Tissue   | Trait | # eQTLs | $R^2$ | Most significant GWAS SNP | Distance (Mb <sup>a</sup> ) | TWAS- $P^b$           | SKAT- $P^c$           | FDR <sup>d</sup> | $P$ -adj <sup>e</sup> |
|----------|-----------------|----------|-------|---------|-------|---------------------------|-----------------------------|-----------------------|-----------------------|------------------|-----------------------|
| 5p15.33  | <i>EXOC3</i>    | Adipose  | BE/EA | 28      | 0.042 | rs9918259                 | 0.22                        | $1.81 \times 10^{-5}$ | $8.24 \times 10^{-6}$ | 0.037            | $2.98 \times 10^{-3}$ |
| 6q14.1   | <i>SENP6</i>    | Blood    | BE/EA | 51      | 0.046 | rs76014404                | 13.92                       | $2.23 \times 10^{-3}$ | $7.12 \times 10^{-6}$ | 0.036            | $5.32 \times 10^{-6}$ |
| 11q13.4  | <i>KRTAP5-8</i> | Adipose  | EA    | 23      | 0.047 | rs4930068                 | 69.26                       | 0.27                  | $1.41 \times 10^{-5}$ | 0.049            | $3.28 \times 10^{-5}$ |
| 12q13.11 | <i>ZNF641</i>   | Junction | BE/EA | 4       | 0.014 | rs1247942                 | 65.90                       | $5.16 \times 10^{-6}$ | $3.81 \times 10^{-6}$ | 0.025            | $2.26 \times 10^{-6}$ |
| 14q32.31 | <i>HSP90AA1</i> | Blood    | BE    | 94      | 0.040 | --                        | --                          | 0.66                  | $8.49 \times 10^{-6}$ | 0.037            | --                    |
| 16q23.1  | <i>CFDP1</i>    | Stomach  | BE/EA | 5       | 0.011 | rs1979654                 | 11.07                       | $3.98 \times 10^{-3}$ | $4.29 \times 10^{-6}$ | 0.026            | $2.67 \times 10^{-6}$ |
| 16q23.1  | <i>CHST5</i>    | Junction | BE    | 21      | 0.026 | rs1979654                 | 10.84                       | 0.36                  | $2.45 \times 10^{-6}$ | 0.022            | $3.14 \times 10^{-6}$ |
| 16q23.1  | <i>BCAR1</i>    | Blood    | BE    | 335     | 0.021 | rs1979654                 | 11.14                       | 0.13                  | $6.08 \times 10^{-6}$ | 0.034            | $1.12 \times 10^{-5}$ |

<sup>a</sup> Distance between the gene and the most significant GWAS risk SNP identified from previous GWAS.

<sup>b</sup> P-value for association analyses in standard TWAS.

<sup>c</sup> P-value for eQTL gene-set SKAT association.

<sup>d</sup> FDR based on the pooled set of p-values for eQTL gene-set SKAT associations across three trait comparisons and six tissues.

<sup>e</sup> P-value derived from the SKAT model adjusting for GWAS risk SNPs.

Table 2. eQTL set-based genetic associations with BE, EAC, or BE/EAC at thirteen risk regions reported in prior GWAS.

| Locus    | Gene            | Tissue   | Trait | # eQTLs | $R^2$ | Most significant GWAS SNP | Distance (Mb <sup>a</sup> ) | TWAS- $P^b$           | SKAT- $P^c$            | FDR <sup>d</sup>      | $P$ -adj <sup>e</sup> |
|----------|-----------------|----------|-------|---------|-------|---------------------------|-----------------------------|-----------------------|------------------------|-----------------------|-----------------------|
| 2p24.1   | <i>LDAH</i>     | Junction | BE/EA | 25      | 0.179 | rs7255                    | 0.005                       | $1.94 \times 10^{-4}$ | $7.11 \times 10^{-8}$  | $2.37 \times 10^{-3}$ | 0.429                 |
| 2p24.1   | <i>GDF7</i>     | Blood    | BE/EA | 15      | 0.079 | rs3072                    | 0.012                       | 0.146                 | $3.38 \times 10^{-7}$  | $5.67 \times 10^{-3}$ | 0.641                 |
| 3q27.1   | <i>YEATS2</i>   | Stomach  | EA    | 6       | 0.018 | rs9823696                 | 0.368                       | 0.160                 | $5.18 \times 10^{-6}$  | 0.030                 | 0.942                 |
| 3q27.1   | <i>ABCF3</i>    | Adipose  | EA    | 43      | 0.063 | rs9823696                 | 0.120                       | 0.223                 | $1.45 \times 10^{-5}$  | 0.049                 | 0.339                 |
| 9q22.32  | <i>BARX1</i>    | Adipose  | BE/EA | 5       | 0.028 | rs11789015                | 0.002                       | $4.35 \times 10^{-6}$ | $3.03 \times 10^{-6}$  | 0.022                 | 0.386                 |
| 15q21.3  | <i>ALDH1A2</i>  | Adipose  | BE/EA | 19      | 0.023 | rs66725070                | 0.022                       | $3.58 \times 10^{-3}$ | $7.87 \times 10^{-7}$  | 0.012                 | 0.506                 |
| 15q21.3  | <i>AQP9</i>     | Blood    | BE/EA | 206     | 0.018 | rs2464469                 | 0.068                       | 0.573                 | $8.08 \times 10^{-6}$  | 0.037                 | 0.054                 |
| 19p13.11 | <i>JUND</i>     | Blood    | BE/EA | 347     | 0.011 | rs10419226                | 0.413                       | 0.221                 | $2.62 \times 10^{-6}$  | 0.022                 | 0.041                 |
| 19p13.11 | <i>SSBP4</i>    | Blood    | BE    | 44      | 0.011 | rs10419226                | 0.273                       | 0.904                 | $1.82 \times 10^{-6}$  | 0.019                 | 0.006                 |
| 19p13.11 | <i>ISYNA1</i>   | Blood    | BE/EA | 206     | 0.028 | rs10419226                | 0.258                       | 0.679                 | $1.35 \times 10^{-5}$  | 0.046                 | 0.085                 |
| 19p13.11 | <i>KLHL26</i>   | Blood    | BE/EA | 32      | 0.015 | rs10419226                | 0.055                       | 0.212                 | $3.21 \times 10^{-7}$  | $5.67 \times 10^{-3}$ | 0.428                 |
| 19p13.11 | <i>CRTC1</i>    | Adipose  | BE/EA | 21      | 0.061 | rs10419226                | 0.009                       | 0.130                 | $6.61 \times 10^{-8}$  | $2.37 \times 10^{-3}$ | 0.002                 |
| 19p13.11 | <i>TMEM161A</i> | Mucosa   | BE/EA | 4       | 0.013 | rs10423674                | 0.412                       | $4.25 \times 10^{-3}$ | $9.02 \times 10^{-10}$ | $1.07 \times 10^{-4}$ | 0.203                 |

<sup>a</sup> Distance between the gene and the most significant GWAS risk SNP identified from previous GWAS

<sup>b</sup> P-value derived from association analyses in standard-TWAS

<sup>c</sup> P-value derived from association analyses in SKAT-TWAS

<sup>d</sup> FDR based on p-value derived from association analyses in SKAT-TWAS

<sup>e</sup> P-value derived from association analyses in SKAT-TWAS after adjusting for GWAS risk SNP

Table 3. Association of eQTLs for the eight genes in Table 1 with BE, EAC or BE/EAC risk in Bonn GWAS.

| Locus    | Gene            | #eQTLs | Trait | #snps in Bonn | #snps ( $P < 0.05$ ) <sup>a</sup> | MinP <sup>b</sup>     | Bonn_ $P$ <sup>c</sup> | Hochberg_P            |
|----------|-----------------|--------|-------|---------------|-----------------------------------|-----------------------|------------------------|-----------------------|
| 5p15.33  | <i>EXOC3</i>    | 28     | BE/EA | 26            | 12                                | $1.69 \times 10^{-7}$ | $1.85 \times 10^{-5}$  | $1.48 \times 10^{-4}$ |
| 6q14.1   | <i>SENP6</i>    | 51     | BE/EA | 48            | 3                                 | $3.55 \times 10^{-4}$ | $5.01 \times 10^{-2}$  | 0.189                 |
| 11q13.4  | <i>KRTAP5-8</i> | 23     | EA    | 20            | 2                                 | $8.02 \times 10^{-4}$ | 0.162                  | 0.324                 |
| 12q13.11 | <i>ZNF641</i>   | 4      | BE/EA | 4             | 3                                 | $4.26 \times 10^{-3}$ | $3.78 \times 10^{-3}$  | 0.026                 |
| 14q32.31 | <i>HSP90AA1</i> | 94     | BE    | 89            | 14                                | 0.012                 | $8.28 \times 10^{-3}$  | 0.049                 |
| 16q23.1  | <i>CFDP1</i>    | 5      | BE/EA | 5             | 1                                 | 0.028                 | 0.053                  | 0.189                 |
| 16q23.1  | <i>CHST5</i>    | 21     | BE    | 20            | 2                                 | $9.81 \times 10^{-3}$ | 0.063                  | 0.189                 |
| 16q23.1  | <i>BCAR1</i>    | 335    | BE    | 328           | 11                                | $4.63 \times 10^{-3}$ | 0.464                  | 0.464                 |

<sup>a</sup> Number of SNPs with univariate p-value < 0.05 in Bonn data.

<sup>b</sup> Minimum univariate p-value in Bonn data.

<sup>c</sup> Validation SKAT p-value in Bonn data.

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## Figure Legends

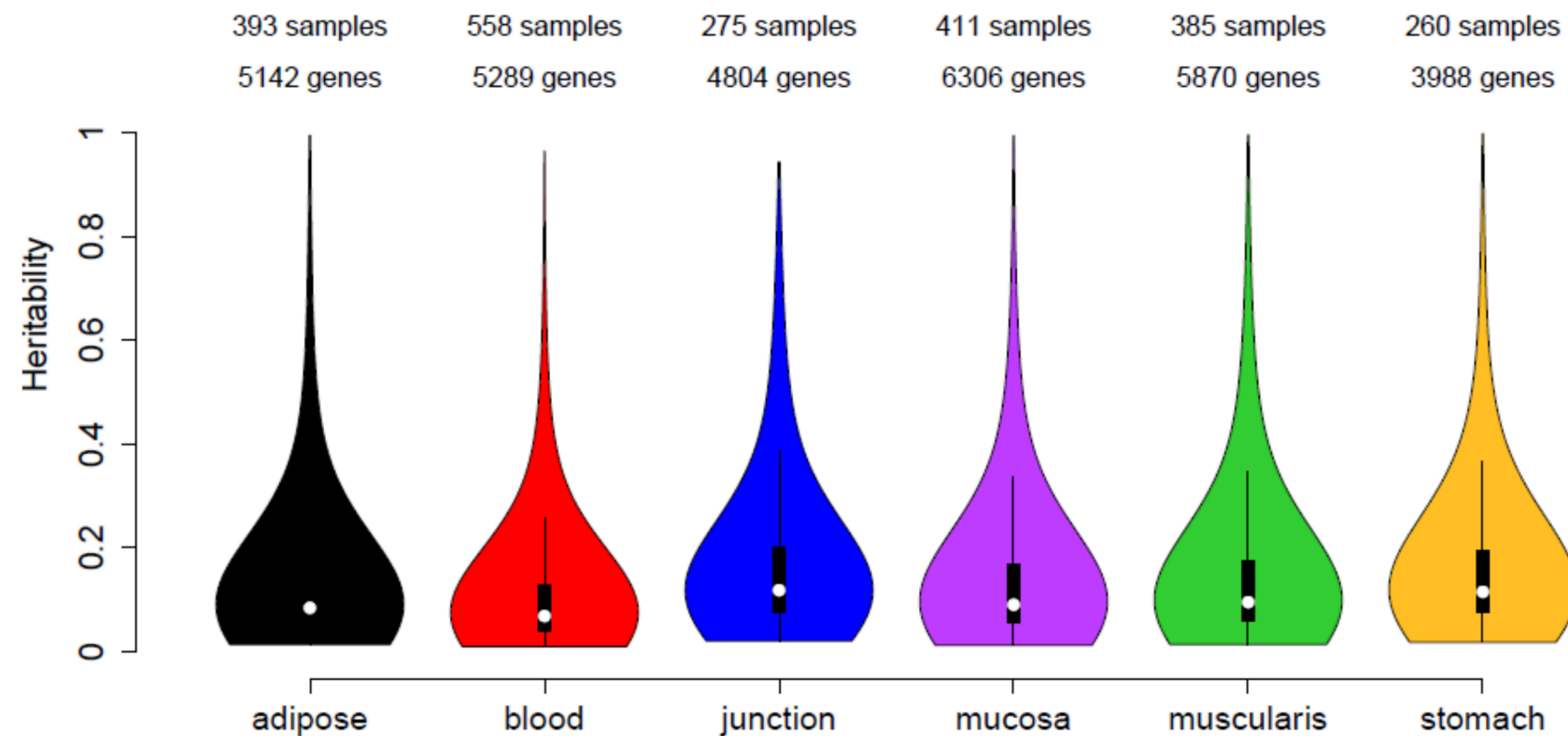
Figure 1. Predictive eQTL models for gene expression across 6 tissues in GTEx. (a) Violin plots for  $R^2$  estimates for genes with  $R^2 \geq 0.01$ . (b) Venn diagram of genes with  $R^2 \geq 0.01$  in three esophageal tissues in GTEx.

Figure 2. Manhattan plots for p-values derived from two methods: eQTL set-based association by SKAT (top); standard TWAS using predicted gene expression (bottom). (a) BE versus control. (b) EA versus control. (c) BE/EA vs control. The line for FDR=0.05 is based on all three trait associations.

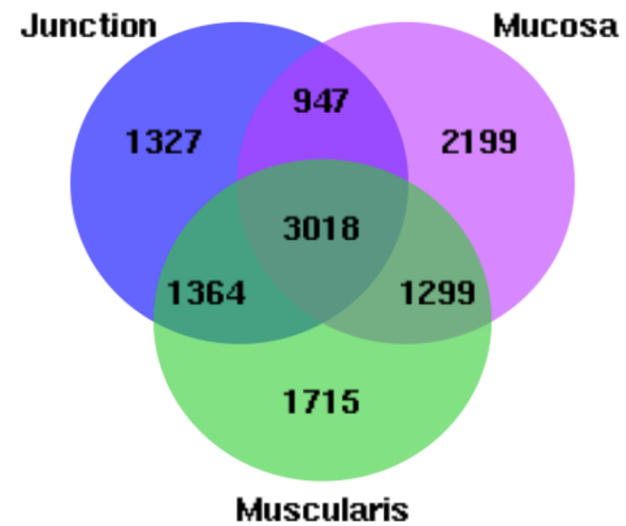
Figure 3. Regional plots for novel loci that were discovered in Beacon/Cambridge discovery set and validated in Bonn data. (a) eQTLs of *EXOC3* in discovery. (b) eQTLs of *EXOC3* in validation. (c) eQTLs of *ZNF641* in discovery. (d) eQTLs of *ZNF641* in validation. (e) eQTLs of *HSP90AA1* in discovery. (f) eQTLs of *HSP90AA1* in validation.

**Figure 1**

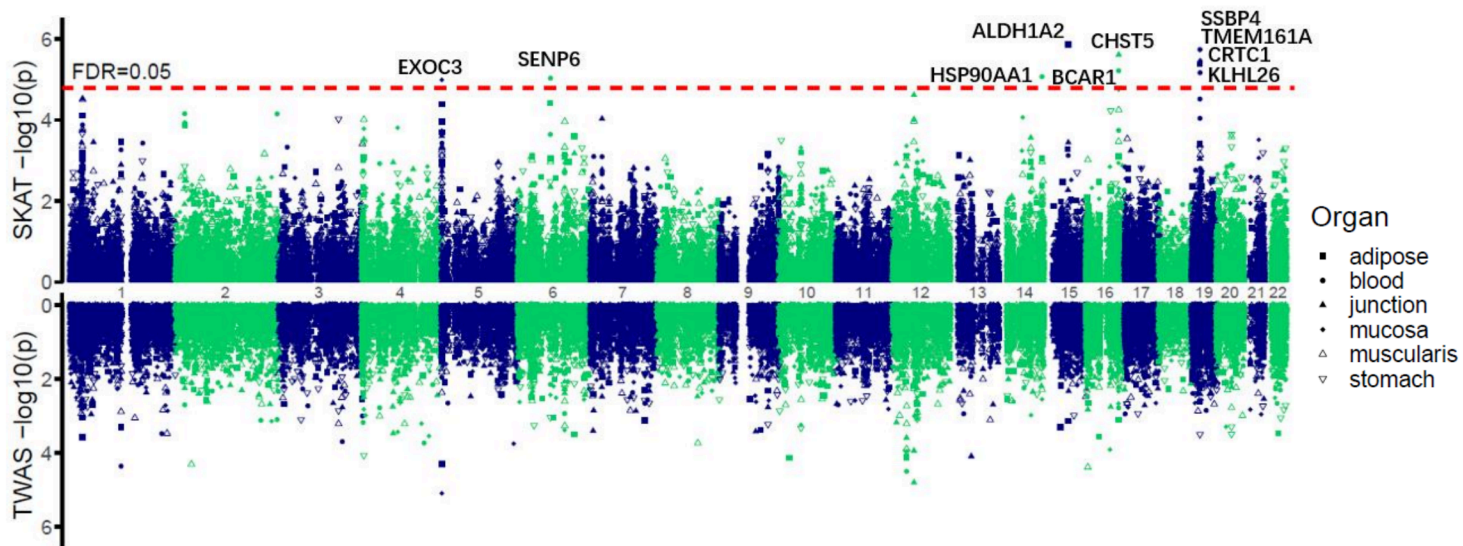
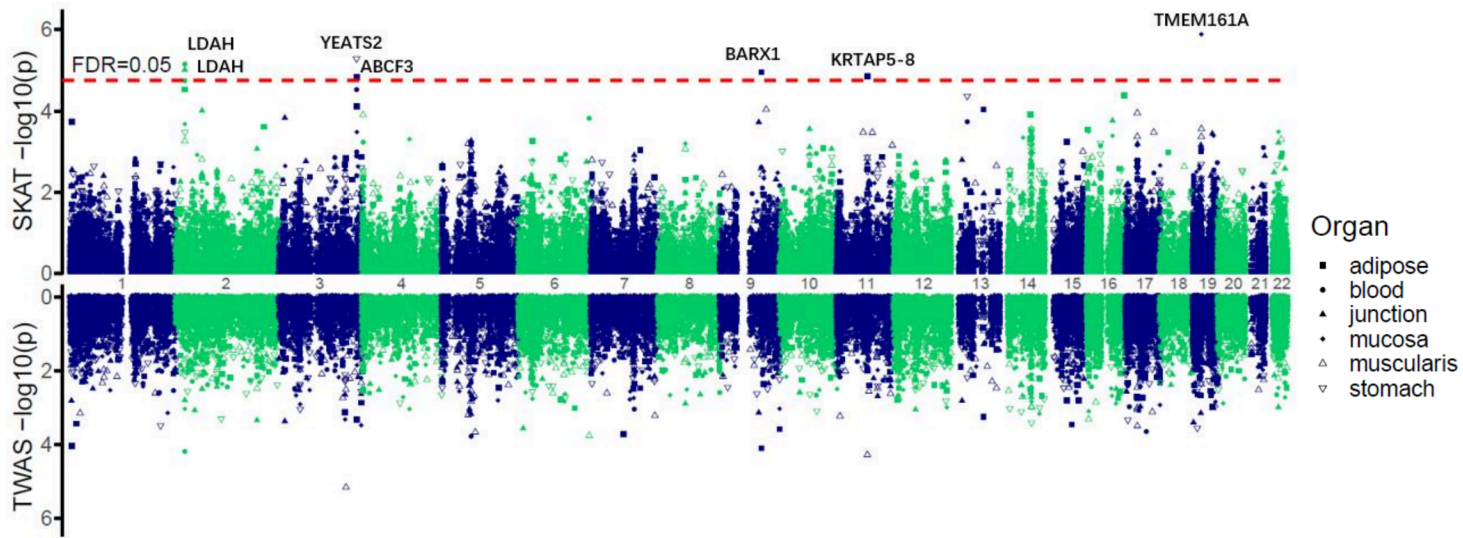
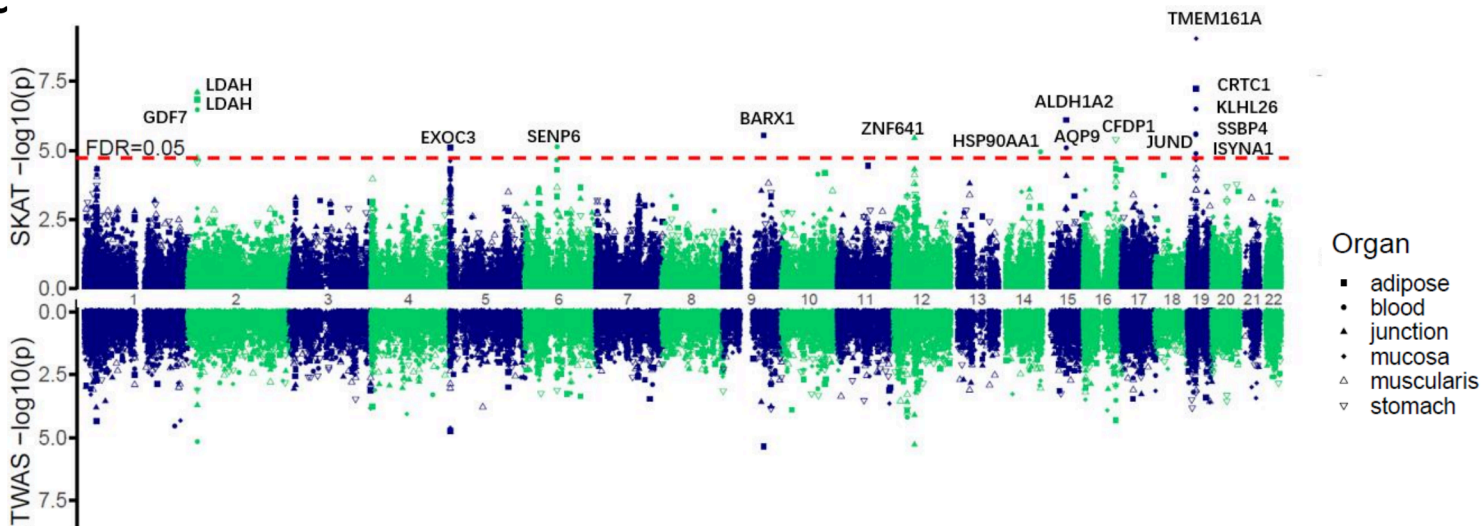
**a**

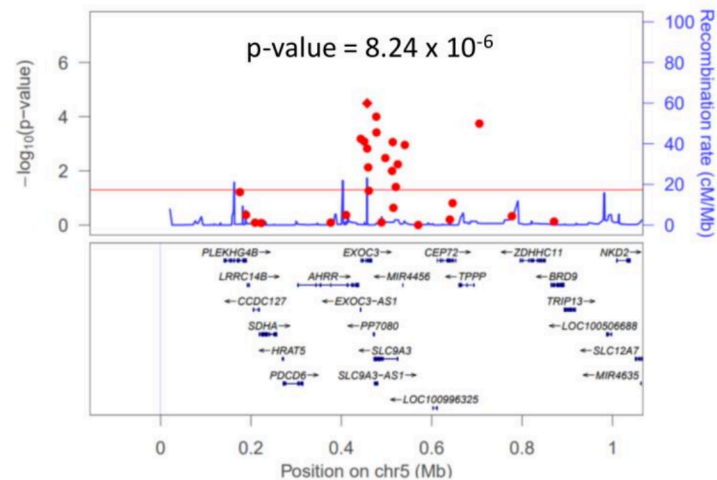
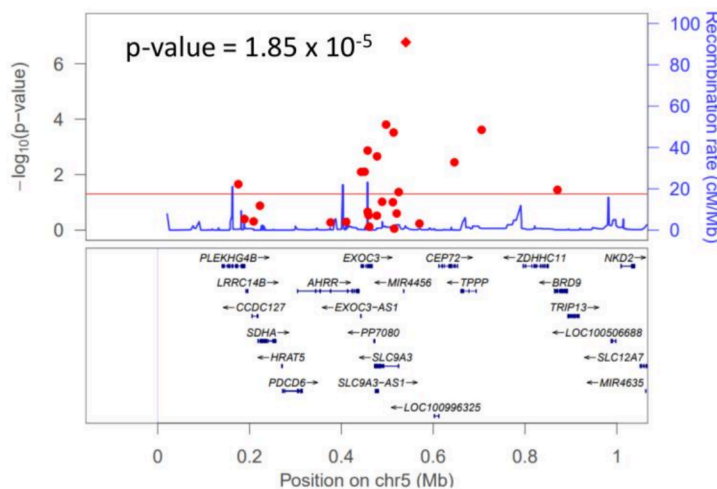
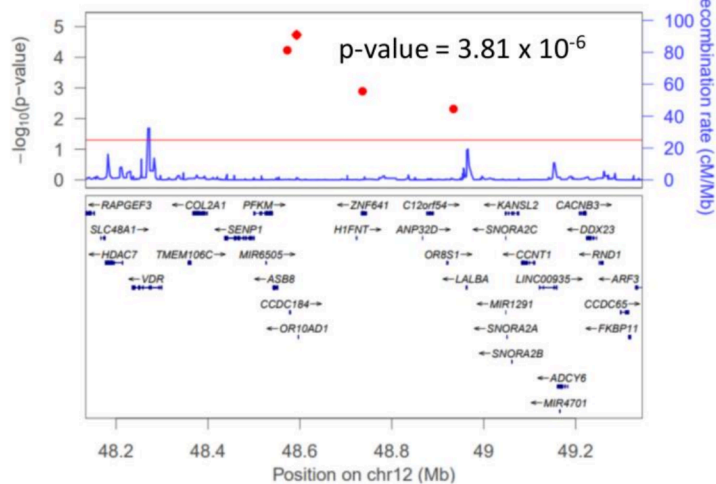
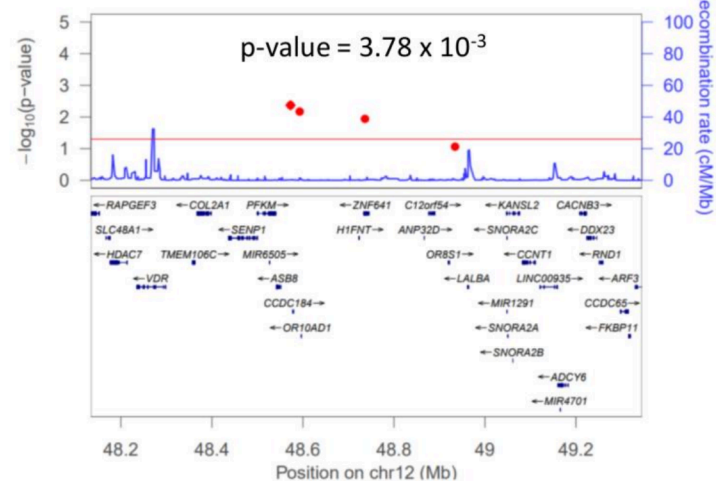
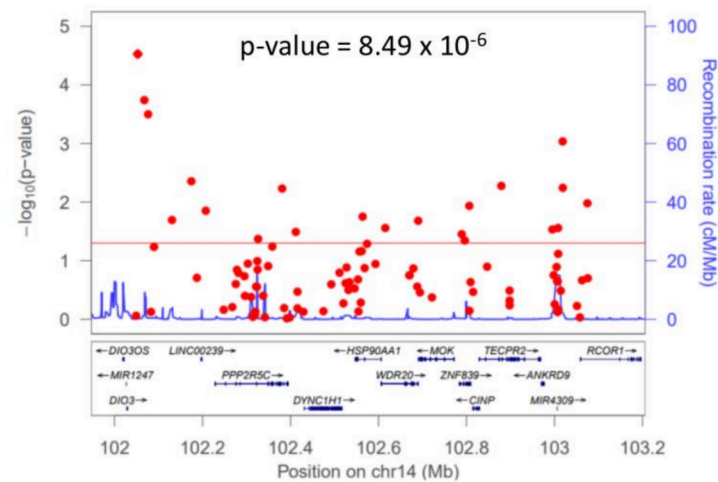


**b**





**Figure 2****a****b****c**

**Figure 3****a** adipose\_BEEA\_BC, EXOC3, 28 SNPs**b** adipose\_BEEA\_Bonn, EXOC3, 26 SNPs**c** junction\_BEEA\_BC, ZNF641, 4 SNPs**d** junction\_BEEA\_Bonn, ZNF641, 4 SNPs**e** blood\_BE\_BC, HSP90AA1, 94 SNPs**f** blood\_BE\_Bonn, HSP90AA1, 89 SNPs